

# Hydrophobic Laser-Induced Graphene Potentiometric Ion-Selective Electrodes for Nitrate Sensing

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## ABSTRACT

Current solid-contact ion-selective electrodes (ISEs) suffer from signal-to-noise drift and short lifespans partly due to water uptake and the development of an aqueous layer between the transducer and ion-selective membrane. To address these challenges, we report on a nitrate ISE based on hydrophobic laser-induced graphene (LIG) coated with a poly(vinyl) chloride-based nitrate selective membrane. The hydrophobic LIG was created using a polyimide substrate and a double lasing process under ambient conditions (air at  $23.0 \pm 1.0$  °C) that resulted in a static water contact angle of  $135.5 \pm 0.7^\circ$  (mean  $\pm$  standard deviation) in wettability testing. The LIG–ISE displayed a Nernstian response of  $-58.17 \pm 4.21$  mV dec<sup>-1</sup> and a limit-of-detection (LOD) of  $6.01 \pm 1.44$   $\mu$ M. Constant current chronopotentiometry and a water layer test were used to evaluate the

potential (emf) signal stability with similar performance to previously published work with graphene-based ISEs. Using a portable potentiostat, the sensor displayed comparable ( $p > 0.05$ ) results to a U.S. Environmental Protection Agency (EPA) accepted analytical method when analyzing water samples collected from two lakes in Ames, Iowa. The sensors were stored in surface water samples for five weeks and displayed nonsignificant difference in performance (LOD and sensitivity). These results, combined with a rapid, and low-cost fabrication technique make the development of hydrophobic LIG-ISEs appealing for a wide range of long-term *in situ* surface water quality applications.

## Introduction

Ion-selective electrodes (ISEs), which convert ionic activity to electric potential (emf), are a common non-destructive method for ion detection in a wide range of industries. The pH meter, a selective glass electrode to the hydrogen ion ( $H^+$ ), is an early example of an ISE [1]. Researchers soon expanded beyond pH monitoring with the advent of selective membrane ISEs [2], and by the late 1960's, several were available for various ions [3]. However, the use of a liquid junction in these ISEs limited their applications due to high maintenance and complications with miniaturization [4]. Solid-contact ISEs remove the inner liquid solutions allowing the ion-selective membrane to make direct contact with the transducing layer. The first example of a solid-contact ISE was the coated-wire electrode (CWE) [5]. Early CWEs had a simple design but possessed a problem with emf drift due to thermodynamically ill-defined ('blocked') interfaces, formation of oxygen half-cells, and/or the presence of a thin water layer between the electrode and ion-selective membrane [4, 6]. These issues with CWEs led to significant research into solid-contact ISEs including various conducting polymers [7, 8] and nanomaterials [9].

Nanostructured conducting allotropes of carbon have proven a promising material for energy and sensing applications due to their mechanical strength, electrochemical properties, and inert nature [10]. Carbon materials such as carbon nanotubes [11], graphene [12], and graphene oxide [13] have been applied as solid contacts for ISEs. Graphene in particular is highly conductive with high mechanical strength that provides an excellent platform for sensing and biosensing [14]. In 2014, Lin et al. [15] first reported on a method to produce a 3D graphitic-like structure commonly referred to as laser-induced graphene (LIG). In this procedure, the  $sp^3$ -carbon atoms in polyimide (PI, Kapton) are photothermally converted to  $sp^2$ -carbon by a  $CO_2$  infrared laser ( $\lambda = 10.6\ \mu m$ ) producing a black porous 3D structure [16]. Further research has demonstrated the ability to form similar material with various lasers [17] and on carbon precursors such as cloth, paper, and food [18]. This maskless, direct writing, one-lasing process eliminates the need for annealing procedures and/or precious metal deposition commonly used on graphene produced through ink-jet printing [19], aerosol-jet printing [20], and chemical vapor deposition (CVD) [21] methods to improve the electrical conductivity and electrocatalytic nature. LIG has become a popular substrate for sensing purpose due to its rapid and scalable production method and has been applied towards electrochemically sensing a variety of analytes including pathogens [22] and various molecules and proteins [23–26]. Ion sensing with LIG was first demonstrated by Garland et al. [27] in soil columns with the goal of deploying these in farm fields for fertilizer monitoring ( $NO_3^-$  and  $NH_4^+$ ). Results from this study show a rapid Nernstian response, and recoveries in soil of 96% and 95% for ammonium and nitrate, respectively. LIG has also been applied to sensing ions in urine [28] ( $K^+$  and  $NH_4^+$ ). Both potassium and ammonium LIG–ISEs accurately assessed hydration levels in real urine samples and displayed a shelf-life of up to three months under dry storage. Recently, LIG was utilized for multi-analyte sensing in sweat including potentiometrically monitoring ions

(K<sup>+</sup>) [29]. This study also reported using LIG as a pseudo reference electrode with electrochemically deposited Ag/AgCl, however, due to influence of Cl<sup>-</sup> ions on the pseudo reference electrode, results show a super-Nernstian sensitivity of 95 mV dec<sup>-1</sup> for potassium. These previous studies have demonstrated the feasibility of LIG (fabricated via one-lasing process) as a transducer for ISEs; however, detailed fabrication of the LIG for this particular function is still lacking, and in particular, studies focused on developing hydrophobic LIG–ISEs for long-term environmental monitoring and the possible pitfalls associated with it (e.g., signal drift and sensor longevity) are still needed.

Herein, we report on a nitrate ISE using hydrophobic LIG fabricated by a double lasing process under ambient conditions (air at  $23.0 \pm 1.0$  °C). A CO<sub>2</sub> laser and PI film were used to create the hydrophobic LIG in a direct writing process without the need for extraneous reagents, post-fabrication annealing processes, or catalyst deposition. This LIG fabrication technique circumvents the need to perform additional chemical modification or lasing in an inert gas environment, demonstrating a more scalable method to create hydrophobic LIG than previous reports. Material characterizations show a 3D porous structure dominated by sp<sup>2</sup>-carbon with Raman spectra showing typical D, G, and 2D peaks of LIG. Wettability testing was performed with five liquids, and a modified Zisman plot was created to estimate the critical surface tension of the hydrophobic LIG. A PVC-based nitrate selective membrane that uses the quaternary ammonium salt tridodecylmethylammonium nitrate (TDMAN) was drop-casted onto the LIG to form a solid-contact at the interface. The hydrophobic LIG–ISE was calibrated using open circuit potentiometry (OCP) and was subjected to selectivity tests, stability characterization, and sensing nitrate in environmental water samples from two lakes in Ames, Iowa with comparable performance to a U.S. Environmental Protection Agency (EPA) accepted method. The results

obtained in this study demonstrate the potential hydrophobic LIG–ISEs have for long-term *in situ* monitoring of ions in surface waters.

## Materials and Methods

All ion-selective membrane chemicals, potassium ferro/ferricyanide, sodium nitrite, and sodium sulfate were purchased from Millipore Sigma (Darmstadt, Germany). Sodium nitrate, sodium chloride, and sodium bicarbonate were purchased from Thermo Fisher Scientific (Massachusetts, USA). Commercial polyimide (PI) film with 5 mil thickness was purchased from McMaster-Carr (Ohio, USA).

### Laser-Induced Graphene Fabrication

Hydrophobic LIG electrodes were designed in CorelDraw (Corel Corporation, Canada) and fabricated with a 75 W M2 Fusion Epilog CO<sub>2</sub> laser (Epilog Laser, Colorado, USA). The first lasing (i.e., hydrophilic LIG) was performed at 15% speed and 7% power followed immediately by a secondary lasing (i.e., hydrophobic LIG) at 20% speed and 3% power. Both laser treatments were performed in air ( $23.0 \pm 1.0$  °C) with a +1 mm offset from the laser focal point and a total lens to material distance of ~41 mm at 1200 DPI. The working area of the electrodes (diameter of 5 mm) was separated from the connection point with a fast-drying lacquer. Contacts of the LIG–ISE were protected with a conductive silver ink (CI-1001, Engineering Materials Systems Inc., Ohio, USA). After fabrication, the LIG electrodes were briefly rinsed with ethanol and dried under nitrogen gas. The nitrate-selective membrane was prepared by mixing 1.5 wt% tridodecylmethylammonium nitrate (TDMAN), 0.5 wt% methyltriphenylphosphonium bromide, 5.75 wt% polyvinyl chloride (PVC), 16.25 wt% 2-nitrophenyl octyl ether (2-NPOE), and 76 wt% THF for a total mass of 1000 mg. After vortex mixing for ~1 min and sonicating for ~3 min, two

applications of 10  $\mu$ L was drop-casted onto the freshly fabricated LIG and allowed to dry overnight in a vacuum desiccator. The hydrophobic LIG-ISEs were then conditioned in 1 mM NaNO<sub>3</sub> for 24 h prior to calibration.

### **Surface Characterization**

X-ray photoelectron spectroscopy (XPS) measurements were performed using a Kratos Amicus/ESCA 3400 instrument. The samples were irradiated with 240 W unmonochromated Mg  $K_{\alpha}$  X-rays. Photoelectrons emitted at 0° from the surface normal were energy analyzed using a DuPont type analyzer. The pass energy was set at 150 eV and a Shirley baseline was removed from all reported spectra. CasaXPS was used to process raw datafiles. Scanning electron microscopy (SEM) analysis was carried out using a FEI Quanta 250 field emission microscope and an accelerating voltage of 10 kV. The samples were coated with 2 nm of iridium to enhance conductivity.

Raman measurements were performed by a Horiba XploRA Plus confocal Raman microscope. A 532 nm laser operating at 1.2 mW and a 50x objective (0.5 NA) was used to check the LIG with 30 s acquisition time and 3 accumulations. Raman spectra at 12 random locations on the LIG were collected. All Raman peaks were fitted to a Lorentzian function in Igor Pro 6.37. The  $I_D/I_G$  and  $I_{2D}/I_G$  ratios were calculated from the fitting results.

A ramé-hart model 90 goniometer equipped with an automated dispensing system (ramé-hart p/n 100-22, ramé-hart instrument co., Succasunna, NJ) was used for all static contact angle measurements using a 3  $\mu$ L droplet. Images were generated and analyzed using DROPImage Pro software where the contact angle was estimated. The wettability of the hydrophobic LIG was tested using five solvents (DI water, methanol, dimethyl sulfoxide (DMSO), formamide, and glycerol (Sigma-Aldrich)). The static contact angles were plotted versus the surface energy of each liquid

to create a modified Zisman plot used to estimate the critical surface tension ( $\gamma_c$ ) of the hydrophobic LIG.

### **Electrochemical Characterization and Nitrate Sensing**

Constant current chronopotentiometry, cyclic voltammetry (CV), and electrochemical impedance spectroscopy (EIS) measurements were performed on a CH Instruments electrochemical analyzer (model CHI7018E, CH Instruments Inc., Texas, USA). Open-circuit potentiometry (OCP) for calibration, selectivity testing, water layer testing, temperature and pH sensitivity, and surface water analysis were performed on a PalmSens 4 potentiostat (PalmSens, Netherlands) equipped with a MUX8-R2 multiplexer. Hydrophobic LIG working electrodes and an external Ag/AgCl electrode (3 M KCl liquid junction) as the reference electrode for all experiments. For chronopotentiometry, CV, and EIS analysis a platinum wire electrode (99.95% Pt, 0.5 mm dia.) functioned as the counter electrode.

The LIG–ISE was calibrated from  $10^{-7}$  to  $10^{-1}$  by 0.5 log steps in DI water under agitation from a small magnetic stir bar (200 rpm) using stock solutions of  $\text{NaNO}_3$  added by pipette. Activities were calculated with the extended Debye–Hückel equation (Equation S3). Selectivity testing was performed using the fixed-interference method (FIM) [30] for four interfering ions ( $\text{NO}_2^-$ ,  $\text{Cl}^-$ ,  $\text{HCO}_3^-$ , and  $\text{SO}_4^{2-}$ ) and a constant concentration of 1 mM while varying the nitrate concentrations. The selectivity coefficients were calculated using Equation S5.

CV and EIS analysis of the bare hydrophobic LIG were performed with 5 mM ferro/ferricyanide in 0.1 M KCl. CV scans were performed from  $-0.4$  to  $0.6$  V at five scan rates from 10 to 200  $\text{mV s}^{-1}$ . EIS frequency range was 1 MHz to 1 Hz with an AC amplitude of 0.2 V and a DC voltage of 0 V. After drop casting the ion-selective membrane, EIS spectra were collected

with unconditioned, 24 h conditioned, 72 h conditioned hydrophobic LIG-ISEs (all in 0.1 M  $\text{NaNO}_3$ ) using the above parameters.

### **Water Layer and Chronopotentiometry**

The potential stability of the ion-selective membrane and hydrophobic LIG substrate interface was studied using constant current chronopotentiometry and a water layer test [31]. For chronopotentiometry measurements, the LIG-ISEs were conditioned in 0.1 M  $\text{NaNO}_3$  for 24 h followed by applying a  $\pm 1$  nA current for 60 s each while recording the emf signal. For the water layer test, LIG-ISEs were conditioned in 10 mM  $\text{NaNO}_3$  for 24 h followed by monitoring the emf signal for 5 h in  $\text{NaNO}_3$ , then 10 mM  $\text{NaCl}$  for 5 h, and finally 10 mM  $\text{NaNO}_3$  for 14 h. Electrodes were briefly rinsed with DI water in between transfers.

### **Temperature and pH Sensitivity**

Temperature influence on the sensitivity of the LIG-ISEs were studied at three temperatures (23.0, 30.0, and  $45.0 \pm 1.0$  °C) controlled by a hot plate and monitored with a glass thermometer ( $\pm 0.5$  °C accuracy). The pH influence was tested at two nitrate concentrations levels (1 mM and 100 mM) using National Institute of Standards and Technology (NIST) pH buffer solutions of 5.02, 7.00, 8.95, and 10.01 pH (Thermo Fisher Scientific, Massachusetts, USA).

### **Surface Water Analysis and Sensor Longevity**

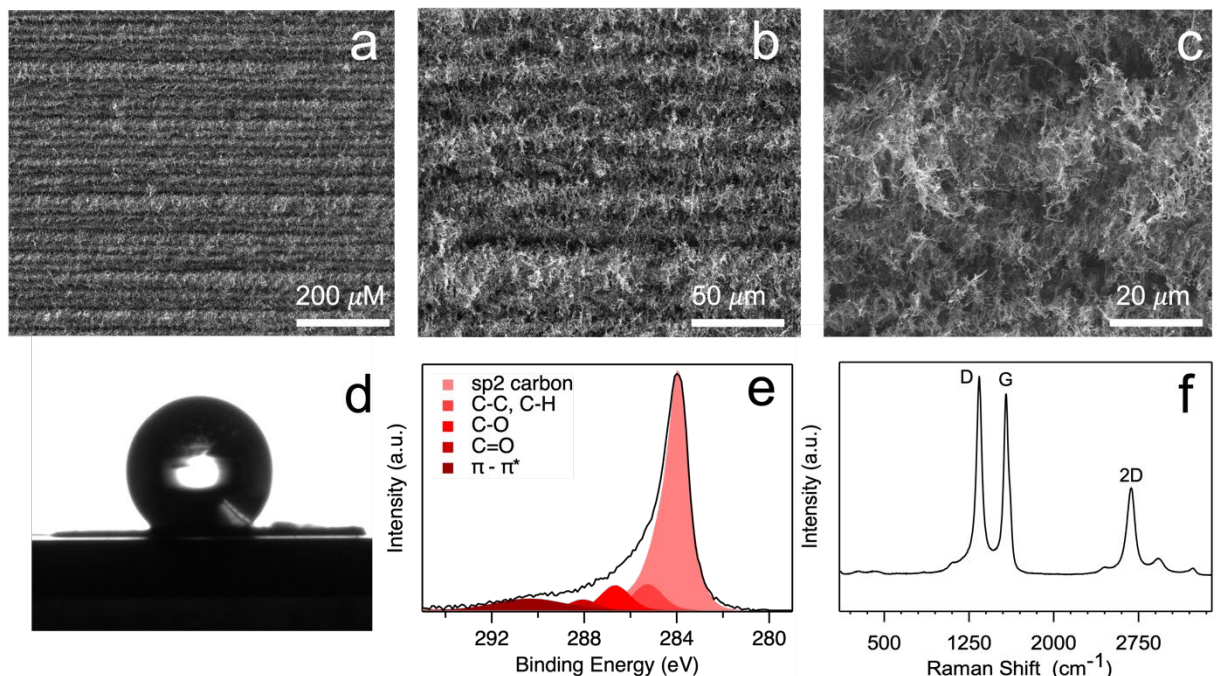
Samples from two local lakes in Ames, Iowa were collected, stored at 2 – 8 °C, and tested within 48 h of collection. For potentiometric ion measurements, U.S. EPA method 9210a was followed. Potentiometric ion sensing results were confirmed by a secondary laboratory with a combined nitrate and nitrite detection Seal Analytical AQ2 method (EPA-114A Rev. 11) where nitrite levels were measured (EPA-116A Rev. 5) and subtracted from the combined measurement results to estimate nitrate concentration.

The LIG–ISEs were submerged in collected water samples for 5 weeks and tested weekly to monitor the change in LOD and sensitivity over time. The samples were stored in ambient lab conditions and sensors were calibrated in whole log steps from  $10^{-6}$  to  $10^{-2}$  with similar conditions as stated above. Cross sectional SEM images were obtained of fresh LIG–ISEs and those stored in water samples to visually compare the effect on membrane adhesion.

### **Data Analysis**

All measurements were made in triplicate and were reported as mean  $\pm$  standard deviation. Data analysis was performed using MATLAB (R2020a, The MathWorks Inc., Natick, MA). Mean response signals from the linear portion of the calibration curve were used to determine the sensitivity and the limit-of-detection (LOD) was calculated using the  $3\sigma$  method. The response time was calculated as the time required to reach 95% of the steady-state signal ( $t_{95}$ ). One-way analysis of variance (ANOVA) and Student's t-test was performed with JMP Pro v.15 statistical software (SAS Institute, Cary, NC) using a significance level of 0.05. All figures were generated in SigmaPlot 14 (Systat Software Inc., San Jose, CA).

## **Results and Discussion**



**Figure 1.** (a), (b), and (c) SEM image after second laser treatment showing the rough flattened hydrophobic LIG. (d) Static contact angle photograph of DI water displaying the hydrophobic nature of the LIG. (e) XPS spectra showing predominately  $sp^2$ -carbon. (f) Average Raman spectra displaying the typical D, G, and 2D peaks of LIG.

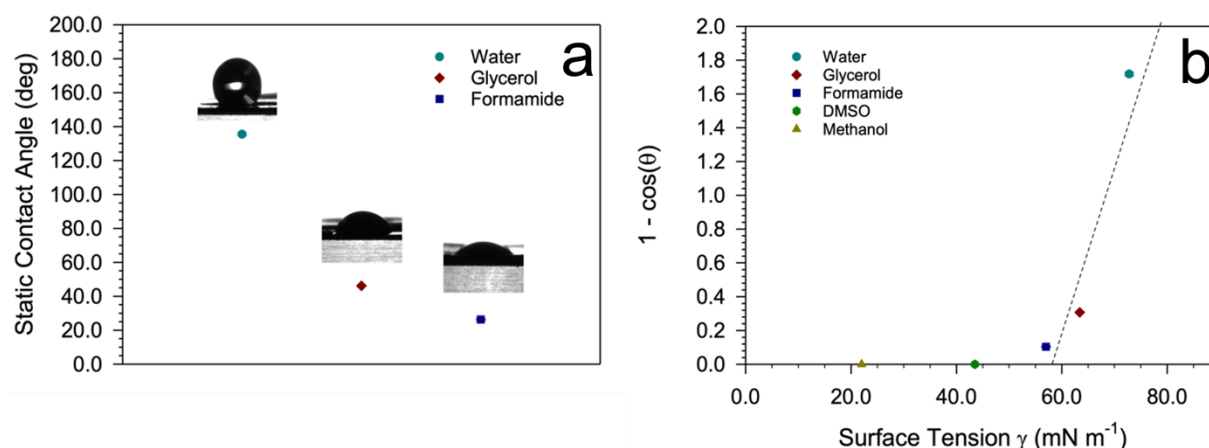
## Surface Characterization

The XPS analysis showed that LIG samples after second laser treatment (Figure 1e) are mainly consisted of  $sp^2$ -carbon evidenced by the asymmetric main peak at 284.0 eV and the broad  $\pi - \pi^*$  peak at 290.0 eV. The  $sp^3$  type hydrocarbon and oxidized carbon are also present on the surface shown by the symmetric, slightly broader peaks ranging from 285.0 to 288.0 eV. Assuming that the carbon content of the polyimide film is 50 – 60% [32], it is likely that the other bonds (i.e., H, N, and O) were broken by the high temperatures and released as gases [16]. XPS analysis after the second laser treatment of the PI film resulted in 92.8% carbon.

Raman spectra of the hydrophobic LIG (Figure 1f) displayed prominent D, G, and 2D bands, which are typical for LIG [15]. The D band ( $\sim 1339 \text{ cm}^{-1}$ ) originates from graphene lattice

defects sites, the G band ( $\sim 1579 \text{ cm}^{-1}$ ) arises from in-plane vibrations of  $\text{sp}^2$ -carbon atoms and is present in all graphitic material, and the 2D band, or G' band, ( $\sim 2682 \text{ cm}^{-1}$ ) originates from a two-phonon double resonance process [33]. The  $I_D/I_G$  ratio was calculated as  $1.1 \pm 0.1$ , which indicates high disorder in the LIG structure [34]. Additionally, the  $I_{2D}/I_G$  ratio was calculated as  $0.5 \pm 0.03$  indicating a multilayer graphitic-like structure [35]. The Raman results were spatially uniform over different sampling locations. (See Table S1 for more details.)

A sufficiently hydrophobic surface is one method of preventing an aqueous layer from developing between the interface of the solid-contact transducing layer and ion-selective membrane [36]. The hydrophobic LIG in this study displayed a static contact angle of  $135.5 \pm 0.7^\circ$  for DI water. The static contact angle was measured for five solvents and results are shown in Figure 2. If these measurements are plotted against the surface energy of each liquid, a modified Zisman plot (Figure 2b) can be used to estimate the critical surface tension ( $\gamma_c$ ) of the hydrophobic LIG by fitting a model using linear regression analysis and extrapolating to the intersection with the x-axis [37]. This value can then be used to quickly assume the behavior of a given liquid on the LIG surface. If  $\gamma < \gamma_c$ , then the liquid should wet the LIG surface and vice versa for  $\gamma > \gamma_c$  [38]. For the hydrophobic LIG,  $\gamma_c$  was estimated as  $57.7 \pm 0.03 \text{ mN m}^{-1}$  suggesting that water ( $72.75 \text{ mN m}^{-1}$ ) [39] should not wet the surface and methanol ( $22.95 \text{ mN m}^{-1}$ ) [39] should, as experimental results confirm.

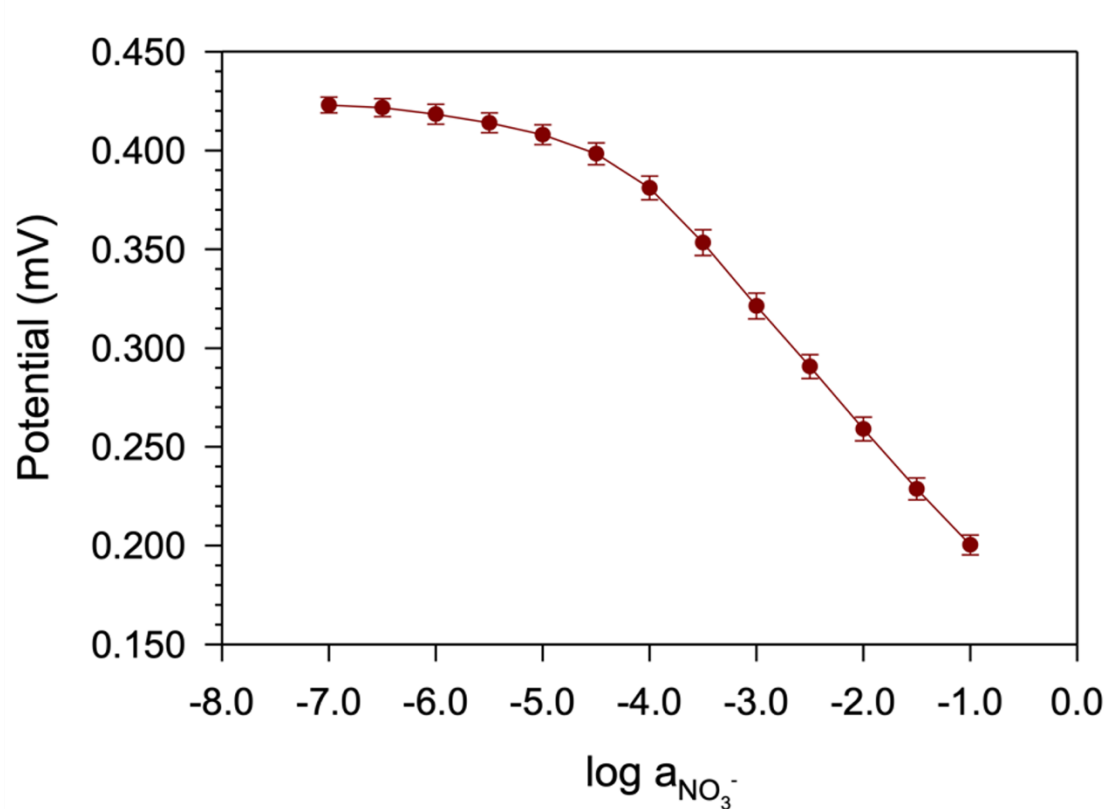


**Figure 2.** (a) Static contact angle measurements and corresponding photographs of water ( $135.5 \pm 0.7^\circ$ ), glycerol ( $46.1 \pm 0.3^\circ$ ), and formamide ( $26.3 \pm 0.5^\circ$ ) droplets. (b) Modified Zisman plot used to estimate the critical surface tension ( $\gamma_c$ ) of the hydrophobic LIG at the intersection of the linear regression line and the x-axis.  $\gamma_c$  was estimated as  $57.7 \pm 0.08 \text{ mN m}^{-1}$ . Data represents mean  $\pm$  standard deviation ( $n = 3$ ).

## Nitrate Calibration and Selectivity

Open circuit potentiometry (OCP) was used to create calibration curves for the nitrate sensors in DI water by sequentially increasing the nitrate concentration in the electrochemical cell (Figure 3 and Figure S5). The slope of the linear portion of the calibration curve was reported as the sensitivity using the average value of the steady-state signal for each concentration. According to the Nernst equation, at 25 °C for a monovalent anion, the sensitivity for an ISE is  $-59.16 \text{ mV dec}^{-1}$  [40]. For the hydrophobic LIG–ISEs, the average sensitivity was  $-58.17 \pm 4.21 \text{ mV dec}^{-1}$ . The slight sub-Nernstian response is possibly due to incomplete membrane conditioning or the diffusion of small molecules (e.g.,  $\text{H}_2\text{O}$ ,  $\text{CO}_2$ ,  $\text{O}_2$ ) across the polymeric ion-selective membrane [41]. The LOD ( $3\sigma$ ) was calculated as  $6.01 \pm 1.44 \mu\text{M}$  and the linear range was  $\sim 10^{-3.5}$  to  $10^{-1} \text{ M}$  with a response time ( $t_{95}$ ) of  $12.52 \pm 3.28 \text{ s}$ . A slight improvement in sensitivity and LOD is demonstrated here with the hydrophobic LIG relative to other LIG-based nitrate ISEs [27], however the LOD is higher than nitrate ISEs based on conducting polymers or organosulfur

compounds such as tetrathiafulvalene (TTF) with similar emf signal drift (See Table S3 for more information). This is likely due to the larger bulk capacitance and smaller resistance of the hydrophobic LIG as compared to the slightly hydrophilic LIG produced from a diode laser ( $\lambda = 405\text{ nm}$ ) [42].



**Figure 3.** Calibration curve for  $\text{NO}_3^-$ . The average sensitivity was  $-58.17 \pm 4.21\text{ mV dec}^{-1}$  and LOD was  $6.01 \pm 1.44\text{ }\mu\text{M}$  with a response time ( $t_{95}$ ) of  $12.52 \pm 3.28\text{ s}$ . Data represents mean  $\pm$  standard deviation ( $n = 3$ )

The fixed interference method (FIM), with a constant interfering ion concentration of 1 mM, was used for all interference testing. The selectivity coefficients (Table 1) are similar to previously reported work intended for agricultural applications [43, 44] with  $\text{Cl}^-$  and  $\text{NO}_2^-$  showing the largest interference.  $\text{NO}_2^-$  is typically not present at large concentrations in surface waters due to multiple rapid oxidation pathways [45] and may not be a cause for concern. However, the

presence of  $\text{Cl}^-$  can create challenges for nitrate ISEs especially intended for environmental monitoring. In areas with high concentrations of  $\text{Cl}^-$  ions, desalination techniques [46] may need to be implemented to accurately estimate nitrate levels *in situ*.

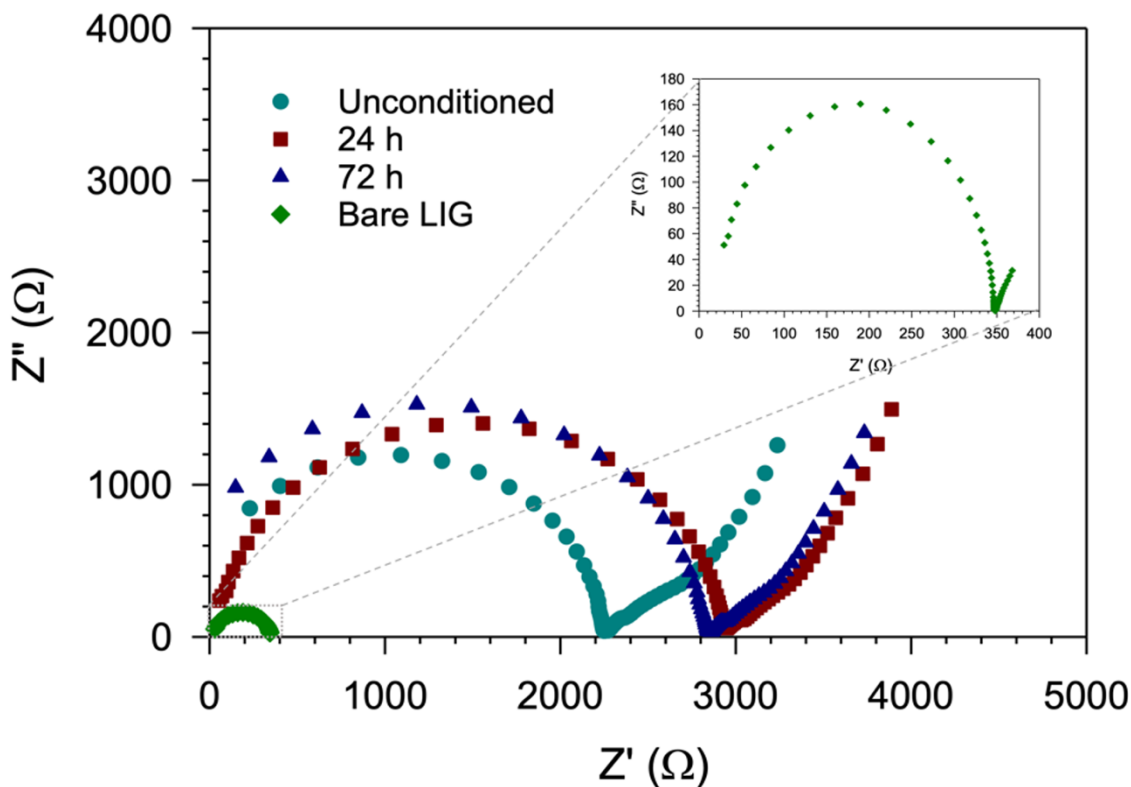
**Table 1.** Selectivity coefficients ( $\log K_{\text{NO}_3^-,j}^{\text{POT}}$ ) at 1 mM interfering ion concentration. Values given are mean  $\pm$  standard deviations (n = 3).

Ion, $j$	$\log K_{\text{NO}_3^-,j}^{\text{POT}}$
$\text{Cl}^-$	$-1.70 \pm 0.09$
$\text{NO}_2^-$	$-1.61 \pm 0.05$
$\text{HCO}_3^-$	$-2.93 \pm 0.08$
$\text{SO}_4^{2-}$	$-2.43 \pm 0.16$

### Electrochemical Impedance Spectroscopy and Cyclic Voltammetry

EIS analysis of ISEs has been proposed to determine physical damage, biofouling, and leaching of membrane components offering a simple and rapid way to characterize ISE functionality *in situ* [47]. This is particularly beneficial for ISEs for wide usage in, possibly remote, environmental settings where issues with physical damaging and biofouling are prevalent. EIS has also been used to show the water uptake of polymeric ion-selective membranes [48]. The high water uptake of PVC-based is a drawback that can lead to water layer formation, emf signal instability, and membrane delamination [49]. Samples of unconditioned, 24 h conditioned, and 72 h conditioned LIG-ISEs were used to evaluate the water uptake during conditioning of the electrodes. Figure 4 shows the resulting EIS spectra and the change in bulk resistance from unconditioned ISEs (2.222

$\pm 0.028 \text{ k}\Omega$ ) to conditioned ( $3.254 \pm 0.363 \text{ k}\Omega$  for 24 h,  $3.208 \pm 0.323 \text{ k}\Omega$  for 72 h). The inset shows the bare hydrophobic LIG charge transfer resistance ( $R_{ct}$ ,  $0.291 \pm 0.057 \text{ k}\Omega$ ). A nonsignificant ( $p = 0.8778$ ) change is shown between the 24 h and 72 h conditioning step. These results are somewhat expected based on in-depth studies of water uptake by polymeric membranes showing uptake beginning only after 10 min upon contact with the aqueous solutions with the majority occurring in the first 10 h [50]. CV analysis of bare hydrophobic LIG (Figure S4) displays quasi-reversible behavior with a peak separation of  $167.0 \pm 9.5 \text{ mV}$  at  $10 \text{ mV s}^{-1}$  and an electroactive surface area (ESA) of  $0.278 \pm 0.038 \text{ cm}^2$ . The ESA is approximately 70% larger than the geometric area ( $0.196 \text{ cm}^2$ ) and is likely due to the unordered porous nature of LIG, allowing for more exposure of edge plane sites to the redox solution [51].



**Figure 4.** Representative EIS spectra of unconditioned, 24 h conditioned, and 72 h conditioned hydrophobic LIG-ISEs in 0.1 M  $\text{NaNO}_3$  with a nonsignificant difference between the 24 h and

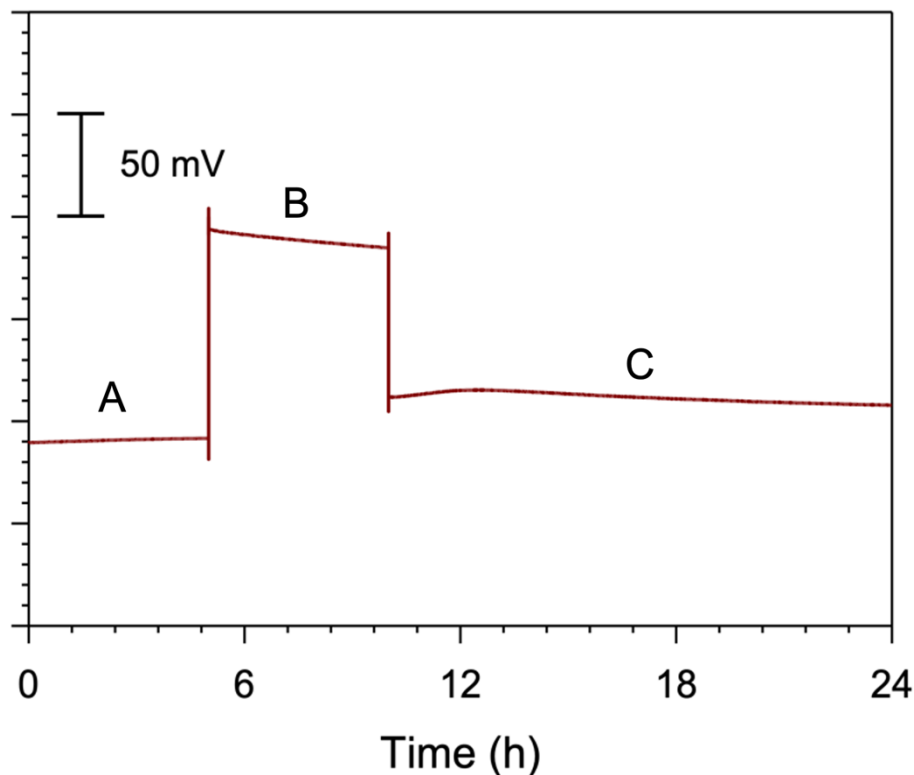
72 h conditioned LIG–ISEs. Inset shows a representative EIS spectra of bare hydrophobic LIG in 5 mM ferri/ferrocyanide in 0.1 M KCl with an  $R_{ct}$  of  $0.291 \pm 0.057$  k $\Omega$ .

### Water Layer Test and Chronopotentiometry

The water layer test (or aqueous layer test) proposed by Fibbioli et al. [31] has become a common validation step in the development of solid-contact ISEs. Results from this test can help determine if a truly solid contact has formed between the ion-selective membrane and the under lying transducer. The formation of an inner water layer can develop upon contact with the aqueous sample, and conditions of the fabrication process (e.g., humidity) have also shown to influence the possibility of an aqueous layer to develop [41]. When a water layer forms, ISEs will experience long equilibration times, substantial emf signal drift, and/or sensitivity to CO<sub>2</sub> partial pressure. Over time the later can continue to spread over the interface affecting the adhesion of the membrane and ultimately lead to delamination. For this study, LIG–ISEs were conditioned for 24 h in 10 mM NaNO<sub>3</sub>, followed by monitoring the emf for 5 h. The electrodes were then transferred to 10 mM NaCl for 5 h, and finally back to 10 mM NaNO<sub>3</sub> for 14 h, all while monitoring the emf signal. Figure 5 shows the resulting time versus emf response with a drift upon return to the 10 mM NaNO<sub>3</sub> of  $0.328 \pm 0.040$  mV h<sup>-1</sup>. The initial jump when changing solutions is due to the change in phase boundary potential at the membrane/solution interface and is a function of the selectivity coefficient and the concentrations of the primary and interfering ions. The hydrophobic nature of the double lased LIG appears to hinder the development of a water layer and shows smaller drift relative to previously reported solid-contact nitrate ISEs intended for long-term use [52].

Potential stability of the emf response can also be influenced by the degree to which the solid contact can be polarized [53]. The small input current (typically < pA) from the high impedance voltmeter should have minimal effect for emf stability. As shown in Equation S4, the

emf signal drift is related to the capacitance of the solid contact where an increase in capacitance will lead to less drift in the emf response. This is particularly prudent when designing solid contact ISEs for continuous monitoring of ions over an extended period of time. Following Bobocka [54], which has become a common method in solid-contact ISE characterization, an applied current of  $\pm 1$  nA should be sufficient to polarize an electrode and evaluate the emf stability. For these tests, each current was applied for 60 s while recording the emf response (Figure S6). The potential drift was  $10.5 \pm 0.96 \mu\text{V s}^{-1}$  and the calculated capacitance was  $95.24 \pm 8.24 \mu\text{F}$ , which are similar to previously published work with graphene-based ISEs [13, 55], however, it is much lower than other carbon-based solid contacts [56, 57]. Increasing the capacitance with conducting polymers or nanoparticles could further improve the emf stability of the LIG-ISEs and may have the added benefit of achieving  $E^\circ$  reproducibility, eventually leading to calibration free LIG-ISEs [58] and will be the focus of future work.



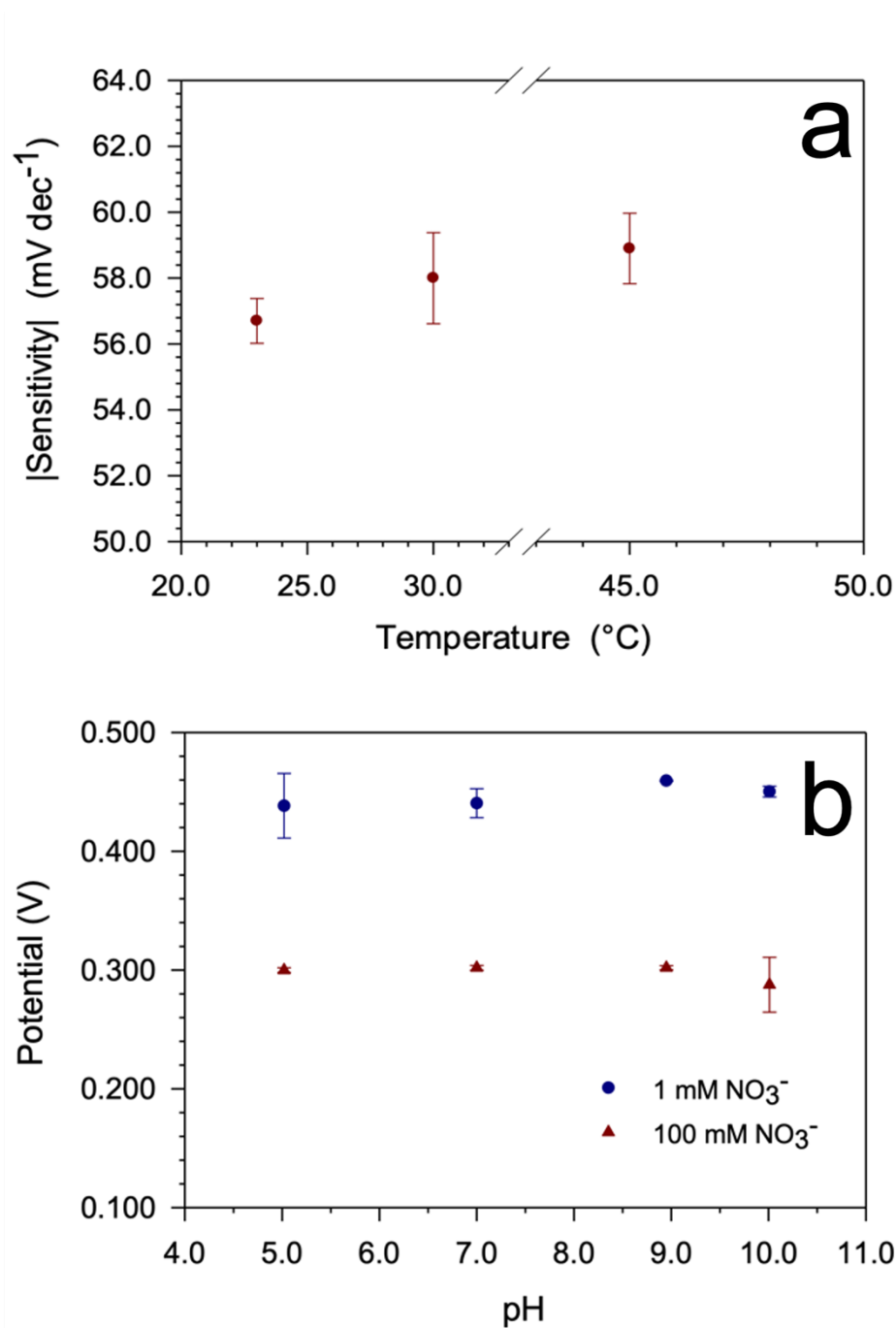
**Figure 5.** Representative 24 h water layer test. Solutions were 10 mM (A) NaNO<sub>3</sub>, (B) NaCl, and (C) NaNO<sub>3</sub>. The drift was  $0.328 \pm 0.040 \text{ mV h}^{-1}$  upon return to 10 mM NaNO<sub>3</sub>.

### Temperature and pH Dependence

Calibration curves were performed at three different temperatures ( $23.0$ ,  $30.0$ , and  $45.0 \pm 1.0 \text{ }^{\circ}\text{C}$ ) in DI water to evaluate the effects on the LIG-ISEs sensitivity (Figure 6a). The increase of temperature resulted in a higher sensitivity which was expected as the Nernst equation has a positive linear dependence on temperature. Similar to previous works [59], the ion-selective membrane here has shown a temperature dependence of  $0.11 \text{ mV } ^{\circ}\text{C}^{-1}$ .

Ion-selective membranes can be affected by pH by altering the membrane charge leading to signal interference. Furthermore, gases such as CO<sub>2</sub> can penetrate into developed water layers, causing change of the pH inside the layer and ultimately alter the emf signal [60]. Water pH is

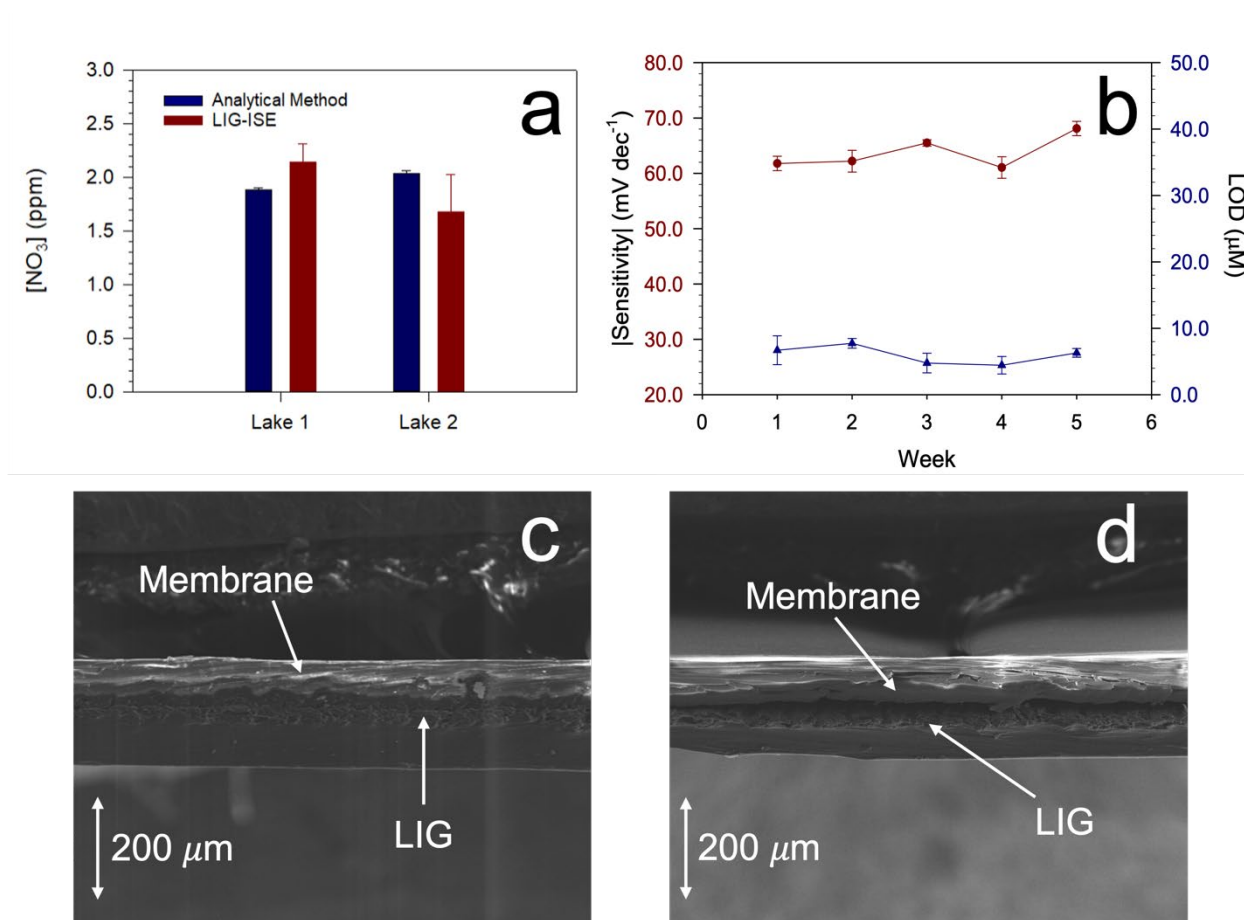
typically within a narrow range, for example, river and ground water ranges from pH 6 to pH 8 [61]. The effect of pH was tested at two different nitrate levels (1 mM and 100 mM) in buffer solutions of pH 5.02, 7.00, 8.95, and 10.01. Figure 6b shows the response for the LIG-ISEs, a nonsignificant difference between the responses was observed for 1 mM ( $p = 0.3487$ ) and 100 mM ( $p = 0.4231$ ) nitrate concentration levels.



**Figure 6.** (a) Temperature effect on the sensitivity at 23.0, 30.0, and  $45.0 \pm 1.0$   $^{\circ}C$ , showing a sensitivity dependance of  $0.11\ mV\ ^{\circ}C^{-1}$ . (b) Emf response at pH levels 5.02, 7.00, 8.98, and 10.01 showing a nonsignificant difference between responses at each concentration level. Data represents mean  $\pm$  standard deviation ( $n = 3$ ).

### Surface Water Analysis and Sensor Longevity

Samples from two lakes in Ames, Iowa were collected and tested according to U.S. EPA method 9210a with the hydrophobic LIG-ISEs to validate the sensors performance in real samples. The LIG-ISEs were calibrated in the water samples and the emf response was monitored for 5 min in each sample. Results are comparable with an EPA accepted analytical method performed by a secondary laboratory, as the concentrations are within the standard deviation of the LIG-ISEs measurement. The LIG-ISEs were then stored in water samples for five weeks with calibration performed weekly to monitor the effect on the LOD and sensitivity, which demonstrate nonsignificant change on these performance parameters over a 3-week period, and the sensors remained operational for up to five weeks. A statistically significant change in the sensitivity of the sensor was observed beyond week three while LOD showed nonsignificant change. This is a slightly shorter storage time compared to some nitrate ISEs; however, these reports do not clarify storage and testing conditions or use unfeasible testing protocols for ISEs intended for *in situ* monitoring of nutrients (Table S3). SEM cross-sectional images were also taken of freshly prepared LIG – ISEs, and those stored in water samples for five weeks (Figure 7c and 7d, respectively). Membrane detachment can be seen clearly in the LIG-ISEs that were stored in surface water samples and could possibly explain the statistically significant change in membrane sensitivity observed after week 3 of storage. Further studies are investigating how to improve the LIG-ISE longevity to monitor nutrients in surface waters *in situ*.



**Figure 7.** (a) Results from analysis of surface water from two lakes showing comparable ( $p > 0.05$ ) performance between the analytical method (blue bars) and LIG-ISEs (red bars). (b) Sensitivity and LOD monitoring over five weeks of storage in surface water samples displaying minimal change ( $p > 0.05$ ) over the five weeks. (c) SEM cross-sectional image of an unconditioned LIG-ISE and (d) SEM cross-sectional of an LIG-ISE after storage in surface water sample for five weeks showing the beginning of membrane delamination. Data represents mean  $\pm$  standard deviation ( $n = 3$ ).

## Conclusion

Higher temporal and spatial *in situ* monitoring of ions, particularly N, P, and K, would allow accurate and precise application of fertilizers reducing costs to farmers and offering researchers a tool to study nutrient run-off and the consequences associated with it. The present study explored implementing hydrophobic LIG fabricated via a double lasing process for monitoring ions in surface water samples. Nitrate was chosen as the ion of interest for this study due to the importance

in fertilizer application, plant growth, and nutrient run-off research. The development of sufficiently selective anion selective membranes is still a challenge, and with the current platform, selectivity studies show notable interference from  $\text{Cl}^-$ , suggesting the possible necessity for desalinization in certain situations. Stability and longevity experiments displayed favorable results comparable to an EPA standard analytical method and demonstrates the potential applicability that hydrophobic LIG-ISEs have for low-cost long-term *in situ* monitoring of ions in water sources. The technique does not require post-fabrication annealing process nor the deposition of expensive metals to increase the electrical conductivity or electrocatalytic nature commonly used with other graphene-based sensors. The maskless lithography-free double lasing fabrication process, performed in an ambient environment, utilizes a laser commonly found in machine shops, demonstrating the feasibility of implementing a facile and scalable roll-to-roll manufacturing procedure for hydrophobic LIG. Future development of LIG-ISEs will focus increasing emf signal stability and work towards calibration-free sensors and the integration with wireless networks to perform long-term in-field studies. This could eventually lead to low-cost solid-contact ISEs for precision agriculture and surface water quality monitoring.

## Associated Content


### Supporting Information

Additional SEM images; XPS and Raman spectra of LIG after the first laser treatment (e.g., hydrophilic LIG); Table of Raman spectroscopy  $I_D/I_G$  ratios after second laser treatment; Representative CV figure of bare hydrophobic LIG; Representative emf response figure; Representative chronopotentiometry response figure; Equations used for electroactive surface area, capacitance, selectivity coefficients, activity values calculations; Nitrate calibration curves using LIG-ISE in surface water samples; Table with results from surface water sample analysis using EPA analytical method and LIG-ISEs measurements, and Table of comparison of recently published reports on nitrate ISE. (PDF)

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## Notes

The authors declare no competing financial interests

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## **ABSTRACT FIGURE**

