

Chapter 12

Population Genetics of Increased Hybrid Performance between Two Maize Populations under Reciprocal Recurrent Selection

J. A. Labate, K. R. Lamkey, M. Lee, and W. L. Woodman

INTRODUCTION

Heterosis, the superiority in one or more characteristics of crossbred organisms relative to their inbred parents, is the basis of the modern cultivars utilized in maize (*Zea mays* L.). Heterosis is of interest in nondomesticated species due to its relevance to the question “how much polymorphism is maintained in natural populations due to selection?” (Berger, 1976). For maize and certain other domesticated species that employ inbred lines to produce commercial hybrids, knowledge of the mechanisms of gene action producing heterosis could contribute to advances in breeding techniques.

One method used to evaluate the existence of heterosis involves measuring multilocus heterozygosity levels in individuals sampled from a population with molecular genetic markers and correlating heterozygosity with a trait believed to reflect fitness, e.g., fecundity, viability, growth rate, or developmental stability. A vast number of these studies, many involving natural populations, have been published during the previous three decades. A general consensus is that a significant positive correlation between multilocus heterozygosity and fitness surrogates has been documented for a systematically wide range of organisms, although it is not a universal phenomenon (recently reviewed by Britten, 1996; Mitton, 1994; Zouros & Foltz, 1987).

The two genetic mechanisms most commonly invoked to explain heterosis are dominance and overdominance. The dominance hypothesis explains heterozygote superiority as a result of the masking of deleterious recessive alleles in an individual, whereas the hypothesis of overdominance postulates an advantage of heterozygosity per se, e.g., through differences in biochemical properties of homozygote vs. heterozygote encoded single-locus products (Berger, 1976). Dominance cannot be distinguished in practice from pseudooverdominance, associative overdominance, or dominance-correlation heterosis. These are all synonyms of heterosis due to the joint action of genes associated in negative gametic phase disequilibrium. Some causes of gametic disequilibrium are directional selection, recombination suppression, inbreeding, or small effective population size (see Houle, 1989 for references).

Heterosis is relevant to the study of several subdisciplines within biology (e.g., plant and animal breeding, mating system evolution, developmental genetics); many good reviews differing slightly in their emphases have been published recently. Sedcole (1981) reviewed examples from plant breeding from approximately 1930 to 1980. Tsafaris (1995) provided a review of recent molecular

techniques used to study heterosis in plants, e.g., looking at RNA amount polymorphism (RAP), protein amount polymorphism (PAP), or DNA methylation levels. A review of heterosis as it relates to plant inbreeding depression can be found in Ritland (1996).

Berger (1976) reviewed theoretical mechanisms for the superiority of heterozygosity per se for protein polymorphisms; many of these have intuitive appeal. Presently, there are only a few well-documented instances of overdominance as a mechanistic explanation of heterosis in natural populations (see Mitton, 1994). The same can be said for domesticated species. One maize example is often cited: Schwartz (1973) found that active and stable heterodimers of alcohol dehydrogenase (Adh) are made up of two monomers, one of which is inactive (but stable) and the other of which is labile (but relatively active). The paucity of examples of single-locus heterosis may not be due to its infrequency, but may be because it is difficult to study, and overdominance is infrequently the sole supporting hypothesis.

Crow (1993, p. 15) recently reviewed genetic evidence that has led to the disfavor of the overdominance hypothesis in lieu of simple dominance as an explanation of heterosis in maize. Most importantly, researchers have found positive evidence for pseudoverdominance. This came from experiments in which hybrid maize populations were advanced several generations and recombination broke up linkages between favorable dominant and deleterious recessive alleles (e.g., Gardner and Lonnquist, 1959). Additional reasons for accepting the dominance hypothesis, according to Crow (1993), are a larger deleterious mutation load than originally generally believed (measured in a few species) and successful selection for relatively high yielding maize inbred lines (compared to early hybrids). The mutation load can explain the observed 15 to 20% grain yield increases observed in maize hybrids over their panmictic base populations, and high yielding inbred lines would not be possible if overdominance was the mechanism underlying high yield.

QUANTITATIVE GENETIC EVIDENCE FROM THE BSSS X BSCB1 RECIPROCAL RECURRENT SELECTION PROGRAM

In spite of the general acceptance of dominance as the explanation for heterosis in maize today, this was not true 50 years ago. Comstock et al. (1949) proposed a breeding method for maize that they termed recurrent reciprocal selection (now known as reciprocal recurrent selection, RRS). Their motivation for developing the method was, as they stated, to discover a selection method that would be effective regardless of the level of dominant gene action. They proposed that RRS would be beneficial for instances in which overdominance, or situations analogous to overdominance (repulsion phase linkages), existed or when interactions of non-allelic genes (epistasis) were important; it would also exploit additive genetic effects. In theory, RRS is intended to improve the performance of an interpopulation cross of two genetically divergent populations. One cycle of RRS involves development of genetic units within populations (e.g., S_1 lines, first-generation progenies from self-fertilized individuals), reciprocal crosses of genetic units between populations, phenotypic evaluation of these testcrosses, and selection of progenies based on testcross results. Selected progenies are then mated within each population. The next cycle of selection is initiated from these. RRS is designed to allow for genetic recombination within populations to maintain quantitative genetic variation, while minimizing inbreeding. The maintenance of two separate gene pools allows a different allele to be fixed within each population. For loci where this is achieved, interpopulation hybrids are assured to be heterozygous.

Two maize populations, Iowa Stiff Stalk Synthetic (BSSS) and Iowa Corn Borer Synthetic #1 (BSCB1), are currently in their 14th cycle of RRS in the Cooperative Federal-State maize breeding program at Iowa State University. Increased grain yield of the interpopulation cross has been the primary target of selection, with reduced grain moisture at harvest and increased resistance to root and stalk

lodging as secondarily selected traits. Selection has been highly successful; mean grain yield of the interpopulation cross improved 77% by Cycle 11, relative to Cycle 0, with concurrent favorable responses in the other traits (Keeratinijakal & Lamkey, 1993a).

Midparent heterosis for BSSS(R) and BSCB1(R) was estimated as the difference between the mean of the interpopulation cross and the mean of the two parental populations. Inbreeding depression (the reduction in the mean value of a character produced by inbreeding) was measured for the interpopulation cross by selfing their F_1 . Steady increases in heterosis and inbreeding depression for grain yield over 11 cycles were found (Keeratinijakal & Lamkey, 1993a). These were interpreted as resulting from an increase, over time, in heterozygosity of the interpopulation cross. Using Smith's (1983) model Keeratinijakal & Lamkey (1993b) partitioned the genetic response to selection of BSSS(R) and BSCB1(R) into components due to additive and dominance effects and looked for evidence of overdominance. They found (partial to complete) dominance effects to be more important than additive effects in the interpopulation cross, with no evidence for overdominance. Diversity analysis (Moll & Hanson, 1984) of the two populations supported this interpretation. Directional dominance for grain yield and a difference in the frequencies of alleles affecting grain yield between the original populations were also inferred.

Similar results have been reported for other maize RRS programs. Eyherabide and Hallauer (1991a,b) reported on reciprocal full-sib recurrent selection in the BS10 and BS11 populations. They found significant increases in midparent heterosis and inbreeding depression for grain yield in the interpopulation cross over eight cycles of selection. They also detected directional dominance and different frequencies of alleles with dominance effects for grain yield between the Cycle 0 populations. They suggested that selection had caused changes in frequencies of alleles with dominant effects in a different set of loci for each population or that different isoalleles with dominant effects had been selected in each population. Hanson and Moll (1986) also concluded that overdominant gene effects were not evident in the Jarvis and Indian Chief populations after 10 cycles of RRS; alleles having additive or dominant effects were selected.

MOLECULAR MARKER EVIDENCE FROM THE BSSS X BSCB1 RECIPROCAL RECURRENT SELECTION PROGRAM

We have genotyped samples from three populations within BSSS(R) and BSCB1(R), representing three different stages in their selective history (see Labate et al., 1997 for complete details). BSSS and BSCB1 synthetic populations trace back to 16 and 12 inbred lines, respectively. These collections of inbred lines are herein referred to as progenitor (P) populations. Cycle 0 populations were formed by several generations of random-mating bulked seed obtained from a series of crosses between progenitor inbred lines. These BSSS(R) and BSCB1(R) Cycle 0 populations were the starting material for RRS. Finally, we have genotypes from samples from both populations after twelve cycles of RRS (Cycle 12).

The molecular markers used were 82 nuclear genomic restriction fragment length polymorphism (RFLP) loci randomly distributed across all 20 chromosomal arms. The markers were assumed to be selectively neutral, i.e., the alleles at a locus would not differ measurably in their effects on the selected traits. The probes were chosen for their high levels of polymorphism and extensive coverage of the genome. One-hundred individuals from each Cycle 0 and Cycle 12 population were chosen at random for genotyping, as well as single individuals from each of 28 progenitor inbred lines (two of the BSSS progenitor inbred lines had been lost; however, the two parental lines of one of these were included). Each of the 82 RFLP probes was considered to be a single locus, and variants at each locus were assumed to be allelic.

Genetic Diversity

We found that mean gene diversity, expected heterozygosity under random-mating, was initially quite high within BSSS(R) and BSCB1(R). This also can be thought of as the probability of obtaining a heterozygote when two alleles are sampled at random from the population. This probability was around 60% in both progenitor populations. After 12 cycles of RRS, mean gene diversity had decreased to near 30% in each. Coinciding with this, the mean number of alleles per locus in BSSS(R) and BSCB1(R) dropped from about four to less than three. A further question was of interest. Looking at the *total* gene pool of BSSS(R) and BSCB1(R), what happened to genetic diversity over 12 cycles of RRS? If two alleles were sampled at random, one from each population, what would be the probability of obtaining a heterozygote? The increases in heterosis and inbreeding depression of the interpopulation cross seen in the quantitative genetic analyses suggested that the interpopulation cross was becoming more heterozygous. The pooled mean genetic diversity for the progenitor populations was estimated to be 63% and for the Cycle 12 populations, approximately 66%. The two estimates were not significantly different based on their standard errors.

Because of the assumption of selective neutrality of the RFLP markers, the lack of increase in interpopulation gene diversity was not completely unexpected. In fact, the estimated loss of mean genetic diversity *within* each population conformed to theoretical expectations (Nei, 1987, Eq. 13.12) of genetic drift of neutral alleles (i.e., random changes in allele frequency caused by gametic sampling each generation). We could see that, in the face of substantial loss of diversity within each population, the between population genetic diversity had remained high. Genetic diversity is a function of the numbers of alleles at a locus and allelic frequencies. This implied that, in general, different alleles had reached high frequencies in BSSS(R)C12 and BSCB1(R)C12.

Results from a principal components analysis (PCA) (Rohlf, 1994) of the 428 individuals sampled from BSSS(R) and BSCB1(R) populations are shown in Fig. 12-1. Each point represents an individual separated in a three-dimensional space based on the presence/absence of 391 alleles (genotypes for 82 loci). The progenitor lines do not form two discrete clusters according to which population they formed. BSSS(R) and BSCB1(R) were initially nearly genetically identical. By Cycle 0, BSSS(R) and BSCB1(R) seem to be distinct from each other. In the absence of genetic drift and selection, the Cycle 0 populations should have remained clustered with the progenitors. We have inferred that maintenance for several decades of BSSS(R)C0 and BSCB1(R)C0 has altered their genetic constitutions. This was especially evident in BSSS(R), for which it seemed that many rare alleles present in P were not sampled in the modern representatives of Cycle 0 (Labate et al. 1997). By Cycle 12, BSSS(R) and BSCB1(R) were substantially diverged. The separation between the Cycle 0 and Cycle 12 populations include a component due to genetic drift, because a limited number of lines (10 to 20) were selected and recombined each cycle, and a component due to selection, because the recombined lines were not chosen at random.

So far, the results presented have focused on *mean* diversity, and genetic changes across *all* loci. By examining the data, it was clear that some of the loci had experienced extreme changes in allele frequencies over the course of selection. The pertinent question became, "Have any of the loci experienced allele frequency changes that were too large to be explained by genetic drift?" Even though the markers fit a neutral model based on mean levels of gene diversity, this did not preclude that some of the allele frequency changes had been influenced by selection. This could have come about directly through selection or, more probably, through genetic hitchhiking. The hitchhiking effect is seen when selection at a locus changes the frequencies of neutral alleles at closely-linked loci and is conditioned on initial linkage disequilibrium between the loci.

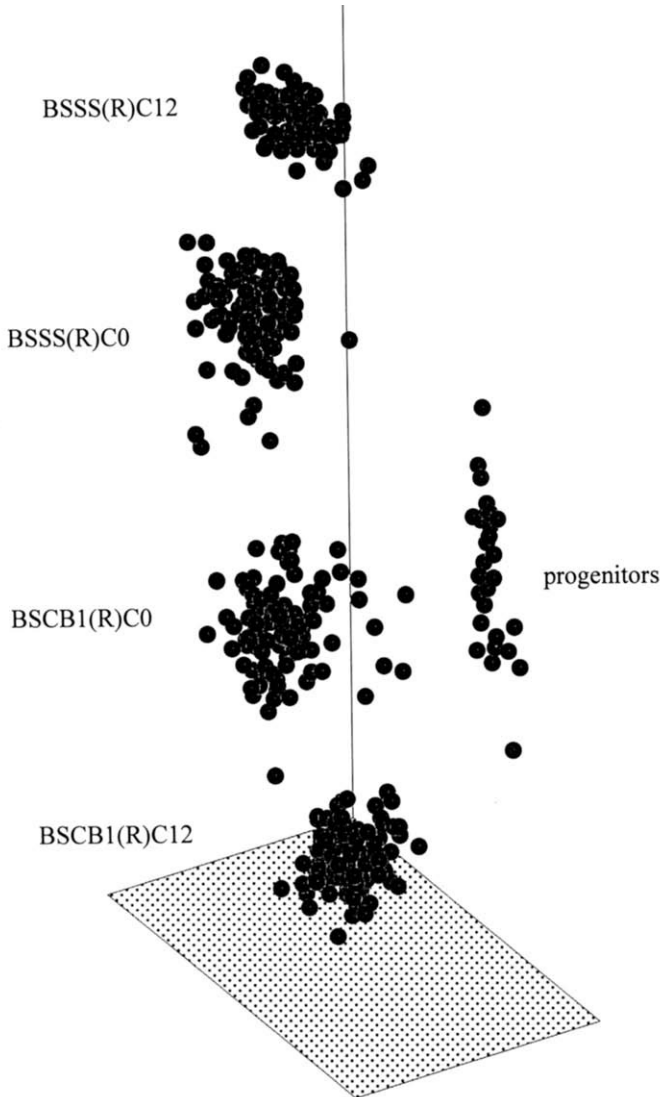


Fig. 12-1. Principal components analysis of Iowa Stiff Stalk Synthetic (BSSS) and Iowa Corn Borer Synthetic #1 (BSCB1) based on genotypes of sampled individuals at 82 RFLP loci. The six sampled populations include progenitor inbred lines, populations before RRS (C0 populations), and populations after 12 cycles of RRS (C12 populations). Progenitor populations do not form two distinct groups.

Effective Population Size

Accurate knowledge of effective population size (N_e) is a key to discerning genetic changes brought about by drift from those that result from selection. Effective population size is defined as the number of individuals in an idealized (i.e., random mating) population that would undergo genetic drift at the same rate as the observed population. Under RRS, N_e is thought to be equal to the number of selected lines each cycle (Vencovsky, 1978). If all parents leave exactly the same number of offspring, N_e is expected to equal $2N - 1$ (Kimura, 1983, p. 41). When the number of selected lines has varied, N_e can be calculated as the harmonic mean of the number of selected lines over all cycles. Our empirical estimates based on the loss of mean genetic diversity between Cycle 0 and Cycle 12 supported an N_e equal to the harmonic mean of the number of selected lines, $N_e = 12$ (Labate et al., 1997). A second method (Waples, 1989a), based on allele frequency changes across all loci, was used to estimate N_e for BSSS(R) and BSCB1(R) populations (Labate et al., 1999). The two methods agreed; N_e is approximately the harmonic mean of the number of selected lines over all cycles. The 95% confidence intervals obtained for N_e using Waples' (1989a) method approached, but did not overlap with, $(2N - 1)$.

Neutrality Tests

Given our estimates of N_e , we applied a test of selective neutrality (Waples, 1989b) to each of the 82 RFLP loci in BSSS(R) and BSCB1(R) populations. The null hypothesis was: the observed variation in allele frequency between two time points can be sufficiently explained as arising from the sampling of a population, of size N_e , that has undergone t generations of genetic drift.

We used estimated frequencies at time points Cycle 0 and Cycle 12 as initial and final allele frequencies, assumed $N_e = 12$ (or 23), and $t = 12$ generations (cycles). Because allele frequency changes at many of the loci between Cycle 0 and Cycle 12 were too large to be explained by genetic drift alone, we interpreted these changes as positive evidence for directional selection and/or genetic hitchhiking. The null hypothesis of drift was rejected for 11 and 17 loci in BSSS(R) and BSCB1(R), respectively, using Waples' test at a probability level of 5%. The loci were found on all chromosomes and were spread throughout the genome. These nonneutral loci fit a pattern of complementary genetic changes between the two populations. Only one was shared between BSSS(R) and BSCB1(R), and at that locus a different allele was reaching high frequency within each population.

The observed allele frequencies at the 27 loci are illustrated in Fig. 12–2. Frequencies of nonneutral alleles are shown at Cycle 0 and Cycle 12 for both populations. Looking within a population at nonneutral alleles identified for that population, rejection of the null hypothesis was associated with an approximately 60% change in an allele's frequency.

We then estimated gene diversity of the interpopulation cross, comparing the 55 neutral loci to the 27 nonneutral loci (Labate et al., 1999). The nonneutral loci increased in mean expected heterozygosity of the interpopulation cross between Cycle 0 (0.664 ± 0.0352) and Cycle 12 (0.776 ± 0.0537) whereas the 55 neutral loci did not (Cycle 0 = 0.603 ± 0.0243 , Cycle 12 = 0.595 ± 0.0384). Comparing the two populations, the 11 nonneutral loci in BSSS(R) contributed to the increase in interpopulation heterozygosity more than the 17 nonneutral loci in BSCB1(R). A partial explanation for this can be found by studying Fig. 12–2, parts c and d. Many of the 17 nonneutral BSCB1(R) alleles were at high frequencies in BSSS(R) at Cycle 0 and remained high in BSSS(R) at Cycle 12 (e.g., bnl835, bnl749, umc155). These loci underwent marked *decreases* in interpopulation expected heterozygosity.

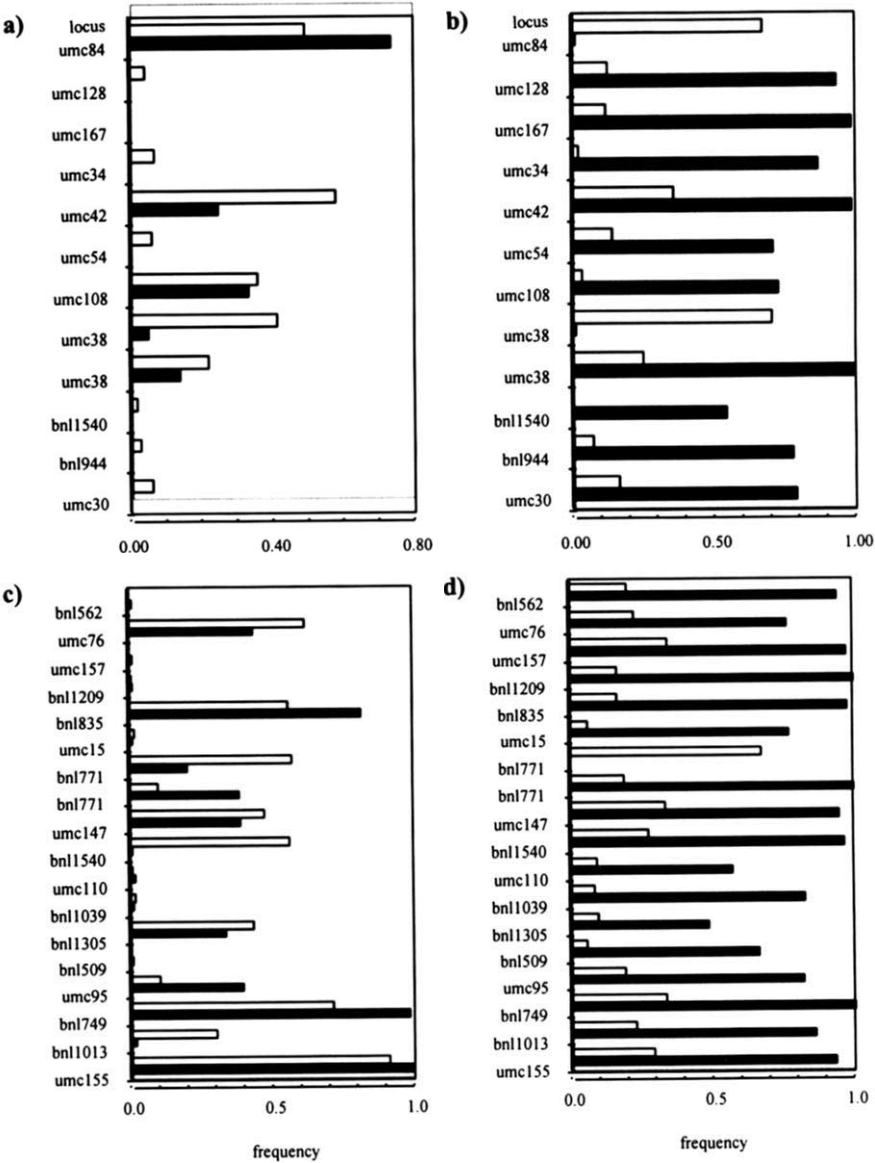


Fig. 12–2. Allele frequencies at Cycle 0 (white bars) and Cycle 12 (filled bars) for 27 nonneutral loci identified in the BSSS(R) and BSCB1(R) populations. a) frequencies in BSCB1(R) for 11 nonneutral loci in BSSS(R), b) frequencies in BSSS(R) for 11 nonneutral loci in BSSS(R), c) frequencies in BSSS(R) for 17 nonneutral loci in BSCB1(R), d) frequencies in BSCB1(R) for 17 nonneutral loci in BSCB1(R).

One prediction under RRS is that if a favorable allele exists in both populations, selection will be more effective for that allele in the population within which it is more common (Cress, 1967). At about one-half of the nonneutral loci, the favored allele was at an initial frequency of less than 10% in the reciprocal population and remained low. The other loci didn't conform to this predicted pattern (Fig. 12-2). Possible reasons for this are (i) at most loci, there were more than two alleles in BSSS(R) and BSCB1(R), so the dynamics of selection were not predicted by this simple model; (ii) intralocus, complete dominance was not the genetic mechanism for increasing the selected allele; or (iii) in the instance of genetic hitchhiking, interlocus correlation (two-locus disequilibrium) patterns were different within BSSS(R) and BSCB1(R).

CONCLUSIONS

Heterosis for grain yield in the interpopulation cross has increased in the BSSS(R) and BSCB1(R) RRS program, and the two populations have become quite genetically diverged from each other. The use of molecular markers has provided some insight into the roles of selection and genetic drift in BSSS(R) and BSCB1(R). Theoretical studies (Li, 1978) have shown that the absolute value of the selection coefficient for an allele must be greater than $1/N_e$ for selection to overcome genetic drift. This assumes a Wright-Fisher model of random genetic drift of neutral alleles (see Hartl & Clark, 1989, p. 351). The selection coefficient is the relative gametic contribution of a particular genotype compared with the most favored genotype in the population (Falconer & Mackay, 1996, p. 26). Our findings imply that a large fraction of loci in the maize genome, about 33% of those surveyed, were affected by selection. If $N_e = 12$ as estimated, then selection coefficients were at least 8%.

Although yield has not been the only agronomic trait selected, it has been emphasized. If yield is affected by many loci that are densely distributed throughout the genome and that carry large phenotypic effects, it is easy to understand why fixation of the most favored genotype in an inbred line derived from an improved population is difficult. Other population genetic studies where molecular markers were used also found that a large fraction of scored loci affected yield (Stuber et al., 1980, 1992), although some studies (e.g., Brown & Allard, 1971; Kahler, 1983) have found that genetic drift could explain observed allele frequency changes. The earlier studies used allozyme loci; DNA-based markers are much more informative in maize.

Stuber et al. (1992), using 67 RFLP loci and nine isozyme loci, genotyped sets of lines descending from a cross originating between two maize inbred lines. When they regressed mean trait value on percent heterozygous marker loci, they found a high correlation between grain yield and proportion of heterozygous markers. A large fraction of the genome was found to affect yield (markers significantly associated with yield were found on all 10 chromosomes), even though this experimental design was limited to detecting regions polymorphic between the two original inbreds.

Reciprocally selected populations should continue to provide a suitable experimental system within which to study relationships between multilocus heterozygosity and phenotype. In this genetic system recombination is prohibited at the interpopulation level, allowing fixation of balanced intralocus or interlocus gene action in the interpopulation cross. Testing theories of gene action requires estimation of parameters such as mutation rates, selection pressure, recombination distances, and inbreeding coefficients (Zouros & Foltz, 1987). It should be possible to obtain more accurate measures of these parameters in maize selection programs than in natural populations.

ACKNOWLEDGMENTS

This study was supported in part by a grant from Pioneer Hi-Bred International, Inc. to K. R. Lamkey and M. Lee. J. A. Labate is funded by the USDA-ARS Postdoctoral Research Associate Program. This work is a joint contribution of the Corn Pest and Crop Genetics Research Unit, USDA-ARS, Dep. of Agronomy, Iowa State Univ. and Journal Paper J-17614 of the Iowa Agriculture and Home Economics Experiment Station, Ames, Iowa, 50011-1010. Project No. 3082 and 3134, and supported by Hatch Act and State of Iowa Funds.

REFERENCES

- Berger, E. 1976. Heterosis and the maintenance of enzyme polymorphism. *Am. Nat.* 110:832-839.
- Britten, H.B. 1996. Meta-analysis of the association between multilocus heterozygosity and fitness. *Evolution* 50:2158-2164.
- Brown, A.H.D., and R.W. Allard. 1971. Effect of reciprocal recurrent selection for yield on isozyme polymorphisms in maize (*Zea mays* L.). *Crop Sci.* 11:888-893.
- Comstock, R.E., H.F. Robinson, and P.H. Harvey. 1949. A breeding procedure designed to make maximum use of both general and specific combining ability. *J. Am. Soc. Agron.* 41:360-367.
- Cress, C.E. 1967. Reciprocal recurrent selection and modifications in simulated populations. *Crop Sci.* 7:561-567.
- Crow, J.F. 1993. Mutation, mean fitness, and genetic load. *Oxford Surv. Evol. Biol.* 9:3-42.
- Eyherabide, G.H., and A.R. Hallauer. 1991a. Reciprocal full-sib recurrent selection in maize: I. Direct and indirect responses. *Crop Sci.* 31:952-959.
- Eyherabide, G.H., and A.R. Hallauer. 1991b. Reciprocal full-sib recurrent selection in maize: II. Contributions of additive, dominance, and genetic drift effects. *Crop Sci.* 31:1442-1448.
- Falconer, D.S., and T.F.C. Mackay. 1996. Introduction to quantitative genetics. 4th ed. Longman. Essex, England.
- Gardner, C.O., and J.H. Lonnquist. 1959. Linkage and the degree of dominance of genes controlling quantitative characters in maize. *Agron. J.* 51:524-528.
- Hanson, W.D., and R.H. Moll. 1986. An analysis of changes in dominance-associated gene effects under intrapopulation and interpopulation selection in maize. *Crop Sci.* 26:268-273.
- Hartl, D.L. and A.G. Clark. 1989. Principles of Population Genetics. 2nd ed. Sinauer Associates, Inc., Sunderland, MA.
- Houle, D. 1989. Allozyme-associated heterosis in *Drosophila melanogaster*. *Genetics* 123:789-801.

- Kahler, A.L. 1983. Effect of half-sib and S1 recurrent selection for increased grain yield on allozyme polymorphisms in maize. *Crop Sci.* 23:572–576.
- Keeratinijakal, V., and K.R. Lamkey. 1993a. Responses to reciprocal recurrent selection in BSSS and BSCB1 maize populations. *Crop Sci.* 33:73–77.
- Keeratinijakal, V., and K.R. Lamkey. 1993b. Genetic effects associated with reciprocal recurrent selection in BSSS and BSCB1 maize populations. *Crop Sci.* 33:78–82.
- Kimura, M. 1983. The neutral theory of molecular evolution. Cambridge Univ. Press, Cambridge.
- Labate, J.A., K.R. Lamkey, M. Lee, and W.L. Woodman. 1997. Molecular genetic diversity after reciprocal recurrent selection in BSSS and BSCB1 maize populations. *Crop Sci.* 37:416–423.
- Labate, J.A., K.R. Lamkey, M. Lee, and W.L. Woodman. 1999. Temporal changes in allele frequency in two directionally selected maize populations. *Theor. Appl. Genet.* (accepted).
- Li, W.-H. 1978. Maintenance of genetic variability under the joint effect of mutation, selection and random drift. *Genetics* 90:349–382.
- Mitton, J.B. 1994. Molecular approaches to population biology. *Annu. Rev. Ecol. Syst.* 25:45–69.
- Moll, R.H., and W.D. Hanson. 1984. Comparisons of effects of intrapopulation vs. interpopulation selection in maize. *Crop Sci.* 24:1047–1052.
- Nei, M. 1987. Molecular evolutionary genetics. Columbia Univ. Press, New York.
- Ritland, K. 1996. Inferring the genetic basis of inbreeding depression in plants. *Genome* 39:1–8.
- Rohlf, F.J. 1994. NTSYS-pc: Numerical taxonomy and multivariate analysis system. Ver. 1.80. Exeter Software, Setauket, NY.
- Schwartz, D. 1973. Single gene heterosis for alcohol dehydrogenase in maize: The nature of the subunit interaction. *Theor. Appl. Genet.* 43:117–120.
- Sedcole, J.R. 1981. A review of the theories of heterosis. *Egypt. J. Genet. Cytol.* 10:117–146.
- Smith, O.S. 1983. Evaluation of recurrent selection in BSSS, BSCB1, and BS13 maize populations. *Crop Sci.* 23:35–40.
- Stuber, C.W., S.E. Lincoln, D.W. Wolff, T. Helentjaris, and E.S. Lander. 1992. Identification of genetic factors contributing to heterosis in a hybrid from two elite maize inbred lines using molecular markers. *Genetics* 132:823–839.
- Stuber, C.W., R.H. Moll, M.M. Goodman, H.E. Schaffer, and B.S. Weir. 1980. Allozyme frequency changes associated with selection for increased grain yield in maize (*Zea mays* L.). *Genetics* 95:225–236.

- Tsaftaris, S.A. 1995. Molecular aspects of heterosis in plants. *Physiol. Plant.* 94:362–370.
- Vencovsky, R. 1978. Effective size of monoecious populations submitted to artificial selection. *Brazil J. Genetics* 1:181–191.
- Waples, R.S. 1989a. Temporal variation in allele frequencies: Testing the right hypothesis. *Evolution* 43:1236–1251.
- Waples, R.S. 1989b. A generalized approach for estimating effective population size from temporal changes in allele frequency. *Genetics* 121:379–391.
- Zouros, E., and D.W. Foltz. 1987. The use of allelic isozyme variation for the study of heterosis. *Isozymes* 13:1–59.

CONTRIBUTORS

J. Axtell	Professor, Department of Agronomy, Purdue University, 1150 Lilly Hall, West Lafayette, IN 47907-1150
D. Andrews	Professor, Department of Agronomy, 328 Keim Hall, University of Nebraska, Lincoln, NE 68583-0915
R. Bernardo	Assistant Professor, Department of Agronomy, Purdue University, 1150 Lilly Hall of Life Sciences, West Lafayette, IN 47907-1150
N.E. Borlaug	Senior Consultant to the Director General, CIMMYT, Lisboa 27, Apdo. Postal 6-641, 06600 México, D.F., México
J.L. Brewbaker	Professor, Department of Horticulture, University of Hawaii, 3190 Malle Way, Rm. 102, Honolulu, HI 96822
M. Cooper	Senior Lecturer, School of Land and Food, St. Lucia Campus, University of Queensland, Brisbane, Qld. 4072, Australia
J.G. Coors	Professor, Department of Agronomy, University of Wisconsin, Madison, WI 53706
H. Cordova	Leader of Tropical Maize Program, CIMMYT, Lisboa 27, Apdo. Postal 6-641, 06600 México, D.F., México
J.F. Crow	Professor, Department of Genetics, University of Wisconsin, Madison, WI 53706
A. Dhopte	Professor, Department of Botany, Punjabrao Krishi Vidyapeeth, Akola 444 104, Maharashtra, India
D.N. Duwick	Affiliate Professor of Plant Breeding, Department of Agronomy, Iowa State University, P.O. Box 446, 6837, N.W. Beaver Drive, Johnston, IA 50131-0446
J.D. Eastin	Professor, Department of Agronomy, University of Nebraska, P.O. Box 830817, Lincoln, NE 68583-0817
J.W. Edwards	Graduate Research Assistant, Department of Agronomy, Iowa State University, Ames, IA 50011-1010
G.O. Edmeades	Interim Director of Maize Program, CIMMYT, Lisboa 27, Apdo. Postal 6-641, 06600 México, D.F., México
G. Ejeta	Professor, Department of Agronomy, Purdue University, West Lafayette, IN 47907-1150
S.A. Engelbrecht	General Manager of Operations, Sensako, P.O. Box 3295, Brits 9700, South Africa

G.I. Gandoul	Department of Agronomy, University of Nebraska, P.O. Box 830817, Lincoln, NE 68583-0817
T.J. Gerik	Blackland Research Center, 808 E. Blackland Road, Temple, TX 76502
H.H. Geiger	Professor of Population Genetics, University of Hohenheim, 350 Institute of Plant Breeding, Seed Science and Population Genetics, D-70593 Stuttgart, Germany
M.V. Ginkel	Head, Bread Wheat Program, CIMMYT, Lisboa 27, Apdo. Postal 6-641, 06600 México, D.F., México.
I.L. Goldman	Associate Professor, Department of Horticulture, University of Wisconsin, 1575 Linden Drive, Madison, WI 53706-1597
J.M. González	Head, Departamento de Coordinación y Desarrollo, Centro de Investigaciones Agrarias de Mabegondo, Apdo. 10, 15080 La Coruña, Spain
M.M. Goodman	William Neal Reynolds and Distinguished University Professor, Crop Science Department, North Carolina State University, Box 7620, Raleigh, NC 27695
C.J. Goodnight	Associate Professor, 115 Marsh Life Science Building, Department of Biology, University of Vermont, Burlington VT 05405-0086
A. Grunst	Consultant, 120 S. 32nd Street, West Des Moines, IA 50265
A.R. Hallauer	C.F. Curtiss Distinguished Professor in Agriculture Professor, Department of Agronomy, Iowa State University Ames, IA 50011-1010
W. Hanna	Research Geneticist, USDA-ARS-SAA, Coastal Plain Exp. Stn., P.O. Box 748, Tifton, GA 31793-0748
V.G. Hernandez	Director, Centro de Genetica, Colegio de Postgraduados, Montecillo, México 56230
D. Hess	Retired Director, CIMMYT Maize Program, #7 Merion Street, Abilene, TX 79606
K. Hoard	Research Analyst, 13247 NW 121st Place, Madrid, IA 50156
M.R.A. Hovney	Agricultural Research Center, Sorghum Research Department, Shandowell Station, Sohag, Egypt
R.B. Hunter	Manager of Product Development, Novartis Seeds, R.R. #1 Plattsville, Ontario N0J 1S0, CANADA
Y. Ibrahim	Graduate Research Assistant, Department of Agronomy, Purdue University, West Lafayette, IN 47907-1150

J. Janick	James Troop Distinguished Professor of Horticulture, Department of Horticulture and Landscape Architecture, Purdue University, West Lafayette, IN 47907-1165
J.P. Jordaan	General Manager of Cereal Grain Research, Sensako, P.O. Box 556, Bethlehem 9700, South Africa
I. Kapran	Sorghum Breeder and INTSORMIL Niger Country Coordinator, Institute National de Recherches Agronomiques du Niger, B. P. 429, Niamey, Niger
M. Kafka	Graduate Research Assistant, Department of Genetics and Plant Breeding, Aristotelian University of Thessaloniki, P.O. Box 261, 540 06 Thessaloniki, Greece
H.A. Knobel	Wheat Breeder, Sensako, P.O. Box 556, Bethlehem 9700, South Africa
J.A. Labate	Research Geneticist, Department of Agronomy, Iowa State University, Ames, IA 50011-1010
K.R. Lamkey	Research Geneticist, USDA-ARS, Department of Agronomy, Iowa State University, Ames, IA 50011-1010
M. Lee	Professor, Department of Agronomy, Iowa State University Ames, IA 50011-1010
A.B. Maunder	Senior Vice President (retired), Dekalb Genetics, 4511 9th Street, Lubbock, TX 79416
J.H. Malan	Wheat Breeder, Sensako, P.O. Box 556, Bethlehem 9700, South Africa
A.E. Melchinger	Professor, Institute of Plant Breeding, Seed Science and Population Genetics, University of Hohenheim, 70593 Stuttgart, Germany
W.R. Meredith, Jr.	Research Geneticist, Crop Genetics and Production Research, Box 314, Stoneville, MS 38776
T. Miedaner	Senior Scientist, University of Hohenheim, Landessaatzuchtanstalt (720), D-70593 Stuttgart, Germany
J.F. Miller	Research Geneticist, USDA-ARS, Northern Crop Science Laboratory, P.O. Box 5677, Fargo, ND 58102
J.B. Miranda Filho	Professor, Departamento de Genética, Universidade de São Paulo/ESALQ, Caixa Postal 83, 13400-970, Piracicaba, São Paulo, Brazil
M.L. Munoz	Centro de Genetica, Colegio de Postgraduados, Montecillo México 56230
V.B. Ogunlea	Professor, Institute for Agricultural Research, Ahmadu Bello University, P.M.B. 1044, Samaru-Zaria, Nigeria

L. M. Onofre	Centro de Genetica, Colegio de Postgraduados, Montecillo México 56230
P. Ozias-Akins	Associate Professor, Coastal Plain Exp. Stn., P.O. Box 748 Tifton, GA 31793-0748
S. Pandey	Maize Program Director, CIMMYT, Lisboa 27, Apdo. Postal 6-641, 06600 México, D.F., México
R. Pandya-Lorch	Coordinator, 2020 Vision for Food, Agriculture and the Environment Initiative, International Food Policy Research Institute, 2033 K Street N.W., Washington, DC 20006
C.L. Petersen	Department of Agronomy, University of Nebraska, P.O. Box 830817, Lincoln, NE 68583-0817
P.A. Peterson	Professor, Departments of Agronomy, Zoology, and Genetics, Iowa State University, Ames, IA 50011-1010
R.L. Phillips	Regents' Professor, Department of Agronomy and Plant Genetics, University of Minnesota, St. Paul, MN 55108
P.L. Pingali	Director, Economics Program, CIMMYT, Lisboa 27, Apdo. Postal 6-641, 06600 México, D.F., México
P. Pinstrup-Andersen	Director General, International Food Policy Research Institute, 2033 K Street N.W., Washington, DC 20006
D.W. Podlich	Quantitative Geneticist, School of Land and Food, St. Lucia Campus, University of Queensland, Brisbane, Qld. 4072, Australia
A. Polidoros	Postdoctoral Fellow, Department of Genetics and Plant Breeding, Aristotelian University of Thessaloniki, P.O. Box 261, 540 06 Thessaloniki, Greece
T.G. Reeves	Director General, CIMMYT, Lisboa 27, Apdo. Postal 6-641, 06600 México, D.F., México
D. Roche	Research Geneticist, USDA-ARS-SAA, Coastal Plain Exp. Stn., P.O. Box 748, Tifton, GA 31793-0748
F. Shaw	Research Associate, Ecology, Evolution and Behavior Department, 1987 Upper Buford Circle, St. Paul, MN 55108
R. Shaw	Associate Professor, Ecology, Evolution and Behavior Department, 1987 Upper Buford Circle, St. Paul, MN 55108
J.S.C. Smith	Germplasm Security Coordinator–Research Fellow, Pioneer Hi-Bred International, P.O. Box 1004, 7300 NW, 62nd Ave., Johnston, IA 50131-1004

O.S. Smith	Research Fellow, Pioneer Hi-Bred International, P.O. Box 1004, 7300 NW, 62nd Ave., Johnston, IA 50131-1004
C.L. Souza, Jr.	Professor, Departamento de Genética, Universidade de São Paulo/ESALQ, Caixa Postal 83, 13400-970, Piracicaba, São Paulo, Brazil
G. Srinivasan	Leader of Subtropical Maize Program and Head of International Testing, CIMMYT, Lisboa 27, Apdo. Postal 6-641, 06600 México, D.F., México
C.W. Stuber	Professor (Emeritus), Department of Genetics, North Carolina State University, Raleigh, NC 27695-7614
W.G. Sun	Researcher, Hawaii Agricultural Research Center, 99-193 Aiea Heights Rd., Aiea, HI 96701
E. Tani	Postgraduate Assistant, Department of Genetics and Plant Breeding, Aristotelian University of Thessaloniki, P.O. Box 261, 540 06 Thessaloniki, Greece
A.S. Tsaftaris	Professor, Department of Genetics and Plant Breeding, Aristotelian University of Thessaloniki, P.O. Box 261, 540 06 Thessaloniki, Greece
S.K. Vasal	Distinguished Scientist and Liaison Officer, CIMMYT Asian Regional Maize Program, P.O. Box 9-188, Bangkok 10900, Thailand
P.K. Verma	Proagro Seed Company Ltd., B-1-39 Toli Chowki, Hyderabad AP 500 008, India
S.S. Virmani	Plant Breeder cum Deputy Head, Plant Breeding, Genetics and Biochemistry Division, International Rice Research Institute, P.O. Box 933, Manila, Philippines
S.J. Wall	Senior Research Associate, Pioneer Hi-Bred International P.O. Box 1004, 7300 NW, 62nd Ave., Johnston, IA 50131 1004
T.C. Wehner	Professor, Department of Horticultural Science, North Carolina State University, Box 7509, Raleigh, NC 27695-7609
M.W. Witt	Kansas Agric. Exp. Stn. Eminence Rt., Garden City Branch Garden City, KS 67846
W.L. Woodman	Research Associate, Department of Agronomy, Iowa State University, Ames, IA 50011-1010
F. Zavala-Garcia	Facultad de Agronomía U.A.N.L., Apartado Postal #358, 66450 San Nicolas de los Garza N.L., Ubicacion de la Facultad, Carretera Zuazua-Marin Km. 17, 66700 Marin N.L., México