# SIB-COMPATIBILITY IN S<sub>1</sub> LINES

## OF MEDICAGO SATIVA L.

by

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## INTRODUCTION

Full utilization of hybrid vigor in alfalfa has not yet been successfully put into commercial practice. Due to the inherent nature of the crop, and especially the absence of desirable inbred lines, commercial hybrid seed production has entailed much effort and expense, thereby rendering the cost of production of hybrid seed to be high.

Commercial production of hybrid seed of alfalfa can be achieved, however, without the necessity of inbreeding. Alfalfa has a perennial nature of growth and can easily be propagated asexually. Furthermore, production of large numbers of inbred lines is not feasible due to the limits imposed by the loss of self-fertility which accompanies inbreeding. Non-inbred clones possessing high combining ability, as determined on the basis of their polycross progeny performance, can be used as parents in hybrid seed production. In order to assure a consistent and high level of performance of the commercial hybrid seed, vegetative propagation of parental clones adequate to plant the entire acreage desired for the production of hybrid seed, is necessary.

This need for extensive vegetative propagation of parental clones has been a definite obstacle in the production of hybrid seed of alfalfa. The additional costs of producing seed in this manner have not as yet been justified

based on the comparative performance of hybrids and the best yielding synthetic varieties.

The necessity for vegetative propagation could be obviated through the use of inbred lines with restored selffertility. Results of numerous studies point to the fact that inbreeding in alfalfa is accompanied by a drastic reduction in self-fertility and as a direct consequence, the number of generations of selfing possible is highly limited. With brother-sister mating, which is another form of inbreeding, it is believed that some of the sterility problems encountered in straight selfing may be overcome, or at least postponed.

The primary objectives of the present study were: (a), to determine the nature of sib-compatibility among  $S_1$  plants of alfalfa derived from clones of low, intermediate, and high self-fertility, (b), to compare fertility of  $S_1$  plants from sib-crossing, continued selfing, and backcrossing to the parental clones, and (c), to determine the possibility of using advanced generation  $S_1$  lines for the commercial production of hybrid alfalfa seed.

#### REVIEW OF LITERATURE

#### Mode of Pollination of Alfalfa

Alfalfa is a naturally cross-pollinated crop. Pollination studies conducted over a wide range of environmental conditions have confirmed that a high degree of natural crosspollination occurs in this crop. Burkhart (1937), in Argentina, found an average of 84.5 per cent cross-fertilization; in Canada Knowles (1943) and Bolton (1948) obtained 94.2 per cent and 90 per cent, respectively. In the U. S. Tysdal <u>et al</u>. (1942) reported an average of 89.1 per cent cross-pollination whereas Pedersen (1953) obtained 86.1 per cent.

There is also a considerable amount of self-fertilization in alfalfa but studies have shown that generally there is a marked reduction in seed-setting following self-pollination. Knowles (1943) working with self-fertile selections and random Grimm plants found an average of 1.65 seeds and 0.56 seeds per flower selfed, respectively, whereas after cross-pollination 4.60 seeds and 3.70 seeds per flower, in the same order, were obtained. Similarly Bolton (1948) obtained an average of 1.58 seeds per flower following self-pollination as opposed to the average cross-fertility of 5.54. Tysdal and Kiesselbach (1944) have reported that in open-pollinated plants 12.6 per cent of the population were found to be between zero and 20 per cent self-fertile and among selfed plants 60.2 per cent

had less than 20 per cent self-fertility. The results just presented all point to the fact that cross-pollination in alfalfa is more potent than self-pollination.

There is also a wide variation in the degree of selffertility among individual plants as reported by Williams (1931). Wilsie (1951) observed a range in self-fertility of single plant selections ranging from 0.12 to 1.84 seeds per flower tripped. Other studies which show remarkably wide range in self-fertility are those of Wilsie (1948), McAllister (1950), and Koffman (1959). Sandal (1946) found a marked difference in self-fertility between low-fertility and highfertility groups of clonal lines. On the average 0.15 seeds and 1.25 seeds per flower tripped, respectively, were obtained. Cooper <u>et al</u>. (1937) obtained an average of 1.25 seeds per flower tripped for the nigh seed-setting plants and 0.07 seed per flower for the low seed-setting group.

"Tripping" as described by Bolton (1948) is a term which refers to the process whereby the staminal column is released from the tissues attaching it to the keel and wing petals and strikes against the standard petal. A consideration of this process in relation to the effective pollination of the alfalfa flower has been dealt with repeatedly. Kirk and White (1933) were of the opinion that self-fertilizing plants of alfalfa do not necessarily require tripping of the flowers for fertilization to occur. He observed that anthers of

ordinary alfalfa shed their pollen in the bud stage of development and pollen grains are deposited on the stigma in abundance. The same observation was made by Armstrong and White (1935). The abnormal positional relationship of the stigma to the anther does not prevent pollination of the untripped flower (Brink and Cooper, 1936). Piper et al. (1914), Hay (1925) and Tysdal (1946) observed that about 5 to 6 per cent of the flowers in some plants can achieve self-fertilization in the absence of tripping. Carlson (1928) and Engelbert (1932) made similar observations, but it is generally agreed that selfpollination with the aid of tripping would result in a much greater increase in seed-setting of the plants. This is substantiated by the results obtained by Piper et al. (1914), Southworth (1928), Carlson (1928, 1930), Clarke and Fryer (1930) and Brink and Cooper (1936). Armstrong and White (1935) were of the opinion that in the act of tripping the stigmatic surface is ruptured and that a stigmatic content is released which initiates pollen germination. Pollen penetration is also facilitated. Pollen germination was found to have occurred in 84 per cent of the tripped flowers and in less than 1 per cent of the untripped flowers.

Piper et al. (1914) and Knowles (1943) have reported cases of automatic tripping. Armstrong and White (1935) maintain that the high seed-setting capacity of certain selected autogamous plants is due largely to the spontaneous

tripping of their flowers.

There are many factors that may induce automatic tripping. Knowles (1943) has found that temperature is the most important of the weather factors in bringing about mechanical tripping. Piper et al. (1914) stressed the importance of intense Tysdal (1946) stated that rain, sun, and wind may sunshine. cause some tripping although Gray (1925) found wind to be a very ineffective agent. Stewart (1926) believed that alternating spells of cloudy and bright weather with moderate wind and occasional showers are favorable for automatic tripping. But the most important factor is the visitation by insects. Insect agents which cause tripping of flowers are mainly responsible for self- and cross-pollination of alfalfa. The role of leaf cutter bees, bumble bees and honey bees and alkali bees has been emphasized (Dwyer and Allman, 1933; Hadfield and Calder, 1936; Jones and Olson, 1943; Hare and Vansell, 1946; Vansell and Todd, 1946; Pedersen and Todd, 1949).

Self-sterility and Cross-compatibility in Alfalfa

There are a number of factors inherent in the alfalfa plant which may be associated with the sterility problems usually encountered in this crop.

## Polyploid nature of alfalfa

Fryer (1930), in a survey of chromosome numbers of 17 species, has shown that the genus <u>Medicago</u> includes species with 14, 16 and 32 chromosomes, but that only tetraploids are present in <u>Medicago sativa</u>. Meiotic studies made by Reeves (1930) and Cooper (1935) revealed that 16 bivalents at Metaphase I are most frequent in tetraploid alfalfa, strongly suggesting a condition of allotetraploidy. On the other hand in a study of diakinesis cells from 12 plants Grun (1949), as reviewed by Atwood and Grun (1951), found an average of 0.62 quadrivalents per cell. Individual plant averages ranged from 0.21 to 0.89 quadrivalents per cell. As many as four quadrivalents per cell were noted, which is suggestive of an autotetraploid origin.

Armstrong (1954) has advanced the theory that tetraploid alfalfa is an alloploid originating from closely related diploid species. The theory is based on the following results obtained. A hexaploid was obtained from crossing a tetraploid and an induced octoploid. Meiotic behavior in the hexaploid showed a low frequency of univalents indicating that the A and B genomes are partially homologous. It was observed also that the quadrivalent frequency in the octoploid was more than three times as high as in the tetraploid, suggesting a lack of complete homology between the two genomes.

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Atwood and Grun (1951) argued that the presence of laggard chromosomes frequently observed would indicate that aneuploids could be expected to occur in relatively high frequency, especially true if the species is highly autoploid, by virtue of the fact that a metabolic balance could be maintained better in an aneuploid from an autoploid than that from an alloploid. The results of the studies of Ledingham (1940) also are cited as evidence of autoploidy. Since an intermediate condition between true alloploidy and autoploidy seems to exist in M. sativa, the term "segmental alloploidy" as first described by Stebbins (1947) is suggested. This condition may account for the non-consistency in the nature of inheritance of characters. All 36 genetic studies involving 25 characters reviewed by Atwood and Grun (1951) indicated a disomic pattern of inheritance. However, in most of these studies F2 and backcross segregations were used and tetrasomic inheritance could not be confirmed until  $F_3$  and later generations were observed. The first convincing evidence of tetrasomic inheritance was reported by Stanford (1951) involving purple-flower color, confirmed later by the work of Twamley (1955), Dudley and Wilsie (1956), Davis (1956), and Markus and Wilsie (1957).

The inherent polyploid nature of alfalfa which results in meiotic irregularity where plants are rendered both poor pollen and seed parents and which may perpetuate an unstable

chromosome condition in the hybrids, contributes to the selfsterility nature of the plant (Armstrong, 1952, 1954; Farley and Hutchison, 1941).

## Somatoplastic sterility

The collapse of ovules during the early stages of postfertilization development has been observed in alfalfa, particularly after self-pollination. According to Brink and Cooper (1939) ovule collapse is frequent in 48 to 72 hours after pollination. Brink and Cooper (1938, 1939, 1940) observed that about 34.4 per cent of the fertile ovules in the selfed series collapsed in the ovary at 72 to 144 hours after pollination in comparison to the 7.1 per cent found in the crossed series. In Medicago falcata, more than 80 per cent of the self-fertilized ovules and less than 12 per cent of the cross-fertilized ovules were aborted, according to Miller (1960). Cooper et al. (1937) found also that while on the average 3.1 ovules per flower in a high-seed setting group were fertilized, only 1.25 seeds per flower developed to maturity. In a low seed-setting group the average number of fertile ovules found was 2.5 but only 0.07 seed per flower was formed. It was observed also that the embryos of the low seed-producing group developed more slowly.

Histological studies made by Brink and Cooper (1939) showed that an abnormal growth took place in the somatic

tissues adjacent to the embryo sac prior to the collapse of the ovule. Such growth was characterized by the finely vacuolate and densely staining nature of the cytoplasm of the cells immediately adjacent to the endosperm, and the resemblance of such cells to the tapetal cells was gradually lost. This condition was first observed on the funicular side of the ovule in the region of the vascular bundle before the breakdown of the embryo sac became noticeable. The breakdown of the endosperm ensued, starting at the chalazal region and it continued toward the embryo.

Ovule collapse, which is commonly referred to as "somatoplastic sterility", is explained by Brink and Cooper (1939) on a basis associated with food translocation. It is believed that the reserved food being translocated to the inner integument and the endosperm, a dominant tissue of the embryo sac, can be equally shared if parallel growth takes place between the two competing tissues. Failure of the endosperm to keep pace with growth offers the maternal tissues a competitive advantage. Consequently the endosperm starves and the ovule eventually collapses. The visible food reserves immediately available are observed to gradually disappear just after fertilization and this is the most critical stage in ovule development. It is maintained also that the reason for ovule collapse occuring in lesser frequency in crosses is the fact that the endosperm of crosses grows at a relatively

faster rate than that of selfs. Farley and Hutchison (1941) found that in the hybrids of a British Columbia variety the proliferation of the endoflexial portion of the integument normally occurs before the initiation of growth of the endosperm thereby the competition which otherwise would lead to somatoplastic sterility is minimized.

## Partial self-incompatibility

The greater potency of cross-fertilization over selffertilization has been indicated earlier. Stevenson and Bolton (1947) have shown that cross-fertilization often occurs when foreign pollen is applied one hour or longer after tripping without emasculation. Brink and Cooper (1938) stated that in partially self-incompatible plants the male gametophytes, although not necessarily impotent on the individual from which they arise, are found to be less effective in effecting fertilization than unrelated male gametophytes. Cooper and Brink (1940) found that with selfpollination only about 14.6 per cent of the eggs in the ovules had become fertile, whereas 66.2 per cent fertile eggs were obtained following cross-pollination.

Partial self-incompatibility is, to a great extent, the result of the inability of the growing male gametophyte to reach the eggs in the ovules of the same plant. Cooper and Brink (1938, 1940) observed that after selfing, few pollen

tubes were able to advance beyond the mid-region of the ovary so that most of the fertile ovules were concentrated in the apical half of the ovary. After cross-pollination the tubes usually reached the base of the ovarian cavity and were able therefore to accomplish fertilization in a greater number of ovules. After self-pollination there was a uniform gradient in fertility which occurred from the apex of the overy downwards, the ovules in the most basal positions rarely showing signs of fertilization. A similar gradient was observed following cross-pollination, but there was a greater proportion of fertilized ovules in each position in the ovary in comparison with that observed after selfing. In diploid alfalfa there was an equal frequency of cross-fertilized ovules in the basal and apical portions of the ovary, according to the observations of Miller (1960). It is believed that in selfing, pollen tubes may seldom reach the basal ovules in the ovary. It was observed that pollen tube growtn, seen at 30 hours and at 48 hours after pollination, was more rapid in cross matings than in the selfs. Cooper and Brink (1940) concluded that partial self-incompatibility is due only in part to restricted pollen tube growth. There was evidence that frequently after selfing the pollen tubes also by-passed the micropyles of the ovules containing the unfertilized egg.

#### Pollen sterility

Pollen sterility also is a factor contributing to low self-fertility. Non-viability of pollen may be brought about by abnormalities in meiosis (Armstrong, 1952, 1954; Farley and Hutchison, 1941) or the physiological disturbances in the maturing pollen (Armstrong, 1952). Bolton and Fryer (1937) noted two classes of sterile pollen; one consisted of clear, empty-appearing grains and the other apparently normal grains which did not germinate. Brink and Cooper (1936) state that occasionally the amount of aborted pollen may be large enough to limit seed formation. Studies conducted by Armstrong and White (1935) and Clarke and Fryer (1930) have shown similar results. Engelbert (1932) has shown a case in which one plant was found to possess sterile anthers and did not produce seed. He further concludes that plants producing a large percentage of sterile pollen are generally poor seed producers.

In a test of pollen viability on an artificial medium Bolton and Fryer (1937) obtained wide variations among different plants, similar to the results found by Sexsmith and Fryer (1943). The pollen viability ranged from 13.3 to 64.0 per cent for the sterile group in contrast to the 50.5 to 87.9 per cent obtained for the fertile plants.

In summary, the incompatibility associated with selfing or crossing is due to the failure of pollen to fertilize the

egg and secondly, the collapse of fertilized ovules resulting from somatoplastic sterility. Cooper and Brink (1940) believe that the observed 98 per cent of the difference between the potential and actual fertility following self-pollination and the 67 per cent difference in cross matings are accounted for by the frequency of fertilization and the collapse of fertile ovules within the first 6 days. In one instance Cooper (1935) observed that in spite of the abundance of growing pollen tubes, less than half of the ovules in the ovary contained fertilized eggs. There were observed in heavy setting lines, for example, 10 to 12 ovules present in the ovary, yet an average of only 3 or 4 seeds per pod developed.

## Genetics of Incompatibility Known in Other Forage Legume Species

The genetics of self- and cross-incompatibility has been studied extensively in other forage legumes, especially the clovers. The "diploid personate type of multiple oppositional allelomorphs" mechanism similar to that found in <u>Nicotiana</u> by East and Manglesdorf (1925), and East and Yarnell (1929) has been found to operate also in red clover, sweet clover, alsike clover and white clover. The theory is that if a pollen grain carries the same allelomorph as one of those in the pistil the resulting pollen tube fails to grow far enough to allow fertilization. On the other hand, if a pollen grain bears an allelomorph different from either of those in the pistil.

pollen tube growth is normal and fertilization is brought about. Williams and Williams (1947a, 1947b) have confirmed its action in red clover (<u>Trifolium pratense</u>) and found 37 out of 40  $S_x$  and  $S_y$  alleles tested to be different. The same mechanism has been found to operate in alsike clover (<u>Trifolium hybridum</u>) as shown by Brewbaker (1953).

Atwood (1940), found that multiple oppositional alleles are also in operation in white clover (<u>Trifolium repens</u>). The same worker (1942) found that of the 26 alleles tested in the first series, 25 (or 96 per cent) proved to be different and in the second series, 34 (or 83 per cent) of the 41 alleles were different. It is believed that the total number existing in nature is much larger. In another study, Atwood (1944), working with 2 series, found 36 (73 per cent) out of 49 alleles tested were different. In the second series, 39 (80 per cent) out of 49 proved to be different.

In sweet clover (<u>Melilotus officinalis</u>), Brink (1934) first observed that the cause of self-incompatibility seemed to be the reduced rate of germination of the pollen on its own stigma, failure of many of the tubes formed to become established in the style, and the slow growth of those which penetrate the pistil. With cross-pollination the pollen germinated freely in large numbers at the base of the style after 24 hours. Evidence has been furnished by Gettys and Johnson (1944) that the oppositional factor type of sterility operates.

In addition to the  $S_X S_Y$  multiple allelomorphic series controlling self- and cross-incompatibility, a fertility factor  $(S_{f})$  has been discovered in the clovers. Atwood (1942, 1945) has shown its occurrence in Trifolium repens. The single factor,  $S_{f}$ , is a member of the multiple allelic series and acts as a dominant or partly dominant allele to the other members of the series in affecting pollen tube growth. As an illustration, in a heterozygous  $S_f S_X$  plant used as a female the  $S_{f}$  functions as a partly dominant factor when all pollen normally would have been inhibited to germinate. But the  $S_{f}$ factor in the pollen has no effect in stimulating other pollens to function in the pistil which bears a similar allele. Furthermore, although the pollen bearing the Sf factor penetrates a style bearing the same allele, it does not compete equally with other pollen bearing entirely different alleles from those in the pistil.

Gettys and Johnson (1944) have demonstrated the presence of an  $S_f$  gene in sweet clover. Its occurrence in red clover also has been shown by Williams and Williams (1947a). The practical use of the fertility gene in a breeding program for red clover was outlined by Rinke and Johnson (1941). Atwood (1942) suggested the applicability of its use in a white clover breeding program.

It is not known whether the genetics of self- and crossincompatibility occuring in the clovers applies directly to

alfalfa. The study in alfalfa is made complicated by the polyploid nature of this crop. Miller (1960), using diploid alfalfa (<u>M. falcata</u>) in the study, hypothesized that selfincompatibility is controlled by two sets of genes derived from the duplication of a single locus. The competitive interaction of two different S genes in the pollen was believed to overcome self-incompatibility. Dean (1942) has advanced an hypothesis based on the autotetraploid inheritance of oppositional alleles in M. sativa.

Genetic studies in other species have shown that selfincompatibility genetically controlled by the multiple oppositional alleles in the diploid may be overcome at the polyploid level. Lewis (1943) illustrated this in Oenothera organensis. Where inhibition of pollen tubes is weak in the diploid it disappeared altogether in the tetraploid. The incompatibility reaction in tetraploids is dependent not on the style being a tetraploid but entirely on the pollen being diploid. He further concluded that growth of heterogenic pollen grain with 2 different S genes on styles with one or with both genes is due to a competition of the S genes for a basic antigenic substance. Atwood and Brewbaker (1953) found that in autoploid white clover, heterogenic pollen may not always be functional where there is a dominance relationship involved among the different S allelles. For example, a dominance relationship  $S_{2} \gg S_{3} S_{4} > S_{1}$  was observed.

Improvement in self-fertility from the diploid to the tetraploid level has been observed in the following species: <u>Solanum spp (Cadman, 1942; Johnstone, 1939), Pyrus communis</u> (Crane and Lewis, 1942), <u>Trifolium repens</u> (Atwood and Brewbaker, 1953), <u>Trifolium hybridum</u> (Brewbaker, 1953), and <u>Trifolium nigrescens</u> (Brewbaker, 1955). On the other hand, no apparent improvement in self-fertility was made in the autoploids of <u>Brassica</u> spp and <u>Raphanus sativus</u> (Howard, 1942), <u>Taraxacum Koksaghyz</u> (Bannan, 1946), <u>Secale cereale</u> (Lundqvist, 1947), <u>Ananus sativus</u> (Kerns and Collins, 1947), and <u>Parthenium</u> <u>argentatum</u> (Gerstel and Riner, 1950). It needs to be proven whether or not tetraploid alfalfa has an improved selffertility over its diploid relatives.

## Environmental Factors Affecting Pollen Tube Growth

Specific factors have been demonstrated to exert influence on pollen tube growth. According to Bolton and Fryer, (1937) relatively high temperature conditions accelerate pollen tube growth, although in their results certain individual plants reacted somewhat differently at two different temperature treatments. A linear relationship was obtained by Sexsmith and Fryer (1943) between temperature and pollen tube growth, the length increasing with an increase of temperature within the range from 70 to 100° F. At 50° F. no germination occurred in one half hour although a slight bulge at the germ

pores of the pollen grains was produced. Grandfield (1945) found that the alfalfa flowers developing into pods decreased in number as temperature went above  $100^{\circ}$  F. The optimum was  $80^{\circ}$  F.

Pollen tube growth reactions to temperatures in alfalfa may not always be constant (Bolton and Fryer, 1937). In <u>Oenothera organensis</u>, Lewis (1943) observed that the reactions of the pollen genotypes to temperature were different. In the diploid, tube growth of pollen possessing a supposedly incompatible genotype had an optimum of  $15^{\circ}$  C. but the compatible tube growth had an optimum at  $30^{\circ}$  C. In the tetraploid, the S<sub>3</sub>S<sub>6</sub> and S<sub>3</sub>S<sub>4</sub> pollen had an optimum growth at  $30^{\circ}$  C. The optimum for the S<sub>4</sub>S<sub>6</sub> genotype was at 15 to  $20^{\circ}$  C.

Moisture and relative humidity also have been found to influence pollen tube growth. Martin (1915) stated that the water requirement for pollen germination was dependent upon a certain ratio between the moisture delivered by the stigma and the moisture of the air surrounding the stigma. Grandfield (1945) found that the number of flowers setting pods increased as the relative humidity decreased from 90 to 10 per cent. At 80° F. there was a difference of 24 per cent between the percentage of pods setting at 10 and 90 per cent relative humidity.

Environmental Factors Affecting Seed Setting

It has been indicated previously that factors which affect pollen tube growth such as temperature and moisture, and flower-tripping agents like rain, sunshine, wind and pollinating insects play important roles in influencing seed setting, due to both self- and cross-fertilization. The amount of water supplied to the crop also may have a decided effect on seed setting. Stewart (1926) stated that the more important alfalfa seed producing areas are located in the arid and semiarid regions of the U.S. He believed that with an abundant supply of moisture the alfalfa plant grows rapidly, giving a high yield of forage but low seed yield. Engelbert (1932) suggested that in Ontario, Canada the limiting climatological factor for seed yield was the amount and distribution of rainfall in summer. Excessive rainfall in July appeared to be the cause of failure in seed production whereas a limited amount of rainfall in the same month offered the best condition for seed production. In Colorado, Blinn (1920) observed an instance in which the abnormally wet weather in spring prevented setting of seeds. Grandfield (1945) reported that high plant reserves tend to increase seed production especially if there is a relatively low supply of soil moisture. Moderate air temperature, low humidity, and soil moisture below the optimum amount tend to produce the type of vegetative growth which stimulates the storage of high organic reserves, resulting in

a physiological condition conducive to seed setting. Tysdal (1946) found that at a thick rate of planting (plants spaced 8 inches each direction) the plants growing in the soil with a relatively low moisture supply produced more seed than those under a high moisture condition. At 32-inch spacing, conditions of high and low soil moisture resulted in practically the same yield.

Other factors influence seed setting. Aicher (1917) believed that there was a close correlation between the number of days of sunshine in the summer and the amount of seed set. Blinn (1920) expressed the opinion that the amount of heat and light influenced seed setting. As Dwyer and Allman (1933), Hadfield and Calder (1936), hare and Vansell (1946) and Vansell and Todd (1946) have shown, the success of alfalfa seed production rests mainly on the activities of pollinating insects, especially wild bees and honey bees in the area.

## Inbreeding in Alfalfa

Results of a number of studies have shown that the process of inbreeding in alfalfa is accompanied by a drastic reduction in seed yield and also by a decrease in forage yield.

Kirk (1927) reported that, on the average, a pronounced and progressive reduction in vigor of growth was observed for each generation of selfing and he noted that the mean yield of seed in the second generation of selfing was markedly less

than that of open-pollinated strains. Williams (1931) found that the yield of seed from the  $S_1$  plants was only 12.4 per cent of that of the parent plants from which they were derived. A reduction of 46.1 per cent in seed set among  $S_1$ lines was obtained by Sandal (1946). Skory (1947) obtained a decrease of 84 per cent in self-fertility from the first to the second selfed generation. Similarly, Koffman (1959) reported a drop in self-fertility of 80 to 90 per cent in the second and third generations of selfing compared to that of the parental clones. It was observed by Bolton (1948) that while seed yield of  $S_1$  to  $S_3$  lines was exceedingly low, one line in the  $S_3$  was found to produce 60 to 69 pounds of seed per acre.

Wilsie (1958) showed that selfing for one generation resulted in a loss of self-fertility of from 80 to 90 per cent and a loss of vegetative vigor of from 20 to 30 per cent. Loss of vigor and the reduction in seed yield were greater in the  $S_2$  than in the  $S_1$ . Results obtained by Tysdal <u>et al</u>. (1942) indicated that forage yield was reduced on an average of 32 per cent in the first selfed generation, with additional decreases in the succeeding selfed generations until a level of 26 to 30 per cent of the original yield was maintained in the seventh and eighth generations. Koffman (1959) found that selfed progenies yielded only 55 to 75 per cent of the forage yield of non-inbred parental clones. Similar results were

obtained by McAllister (1950).

Bolton (1948) showed that progenies of crosses among second generation selfed plants derived from the same parents were intermediate in seed and forage yield between comparable inbred lines and outcross progenies. A partial restoration of self-fertility in sib-matings of  $S_1$  plants was reported by Koffman (1959).

There are other effects associated with inbreeding in alfalfa. Kirk (1927) found a reduction in variability of some important morphologic and physiologic characters. Some chlorophyll deficient seedlings appeared and winter injury damage to some inbreds occurred. Stewart (1934) found a significantly lower variability in selfed progenies for plant height, plant width, stem diameter, leaflet length and width, blossom color, and foliage color.

## Breeding Procedures

Progress made in alfalfa breeding has been reviewed by Bolton (1948) and White (1949).

## Improvement of seed yield by selection

Kirk (1927) obtained a significant correlation (r = .46) in seed setting between the first and second generations of selfed lines and concluded that the high and low seed producing characteristics are inherited. Selection within self-

fertilized lines was suggested as a means of increasing seed yield. This idea was abandoned later because of the loss of vigor resulting from selfing. Tysdal and Kiesselbach (1944) reported a significant positive correlation (r = 0.3781) between self-fertility and seed production under poor seedsetting conditions, but a negative significant correlation (r = -0.3874) was obtained under good seed-setting conditions. The positive association under poor seed-setting conditions was attributed to the fact that selfing was more prevalent and since selfing results in reduced vigor, the progeny of these plants would be low in forage productivity. They believed that selection for self-fertility was not advisable.

Another approach to improving seed yield in alfalfa is through "maternal line selection" as proposed by Fryer (1939). This method consisted of 4-year cycles following each other and continued indefinitely.

In the first year 80 progenies, each containing 50 plants, were grown from seeds. In the second year each plant was scored on the basis of the density of pods, and poor plants and poor progenies were rogued. Plants were scored for fertility again in the third year and only 100 plants, which represented from 30 to 40 different progenies were selected late in the fall. In the fourth year further selection was made and 80 out of the 100 selections made in the previous year, were allowed to set seed under open-pollination. These were

used to initiate the next cycle of selection. In 3-year tests, the alfalfa stock improved through 10 years of maternal-line selection showed a superiority in yield over three unimproved standard varieties and a selfed line. Pedersen (1953) found that over a 3-year period nine selections based on maternal performance were 38.8 per cent better in seed production than the average of six checks.

## Utilization of hybrid vigor for forage yield

Tysdal et al. (1942) showed evidence of considerable hybrid vigor from hand-pollinated crosses between inbred lines, depending on the combining ability of the parents. Single crosses yielded from 60 to 139 per cent of the average of Grimm, Hardistan and Ladak, but the average of 28 hybrids was only 96 per cent of the average yield of the check varieties. Tysdal and Kiesselbach (1944) obtained results from forage yield tests of 31 different  $F_1$  hybrids which showed marked hybrid vigor resulting from crossing. Some double crosses produced by crossing two F<sub>1</sub> hybrids exhibited as much vigor as the single crosses from inbred lines. In the order of yield, the top five hybrids were those built up from 3, 4, 2, 5, and 8 lines respectively. The average yield of the 5 hybrids exceeded that of the highest yielding variety by 23 per cent. Waldron (1920) found 47.5 per cent increase in forage yield in a cross between M. sativa and M. falcata,

as compared to that of the parents.

Tysdal and Kiesselbach (1944) suggested a procedure for the commercial production of  $F_1$  hybrid seed in alfalfa similar to that being followed in corn. Clonal lines used for the production of the hybrid are chosen on the basis of their combining ability. While the best measure of combining ability is average single cross performance, Tysdal and Crandall (1948) concluded that polycross progeny performance was a satisfactory method of testing the combining ability of clones. Wilsie and Skory (1948) obtained a correlation of r = 0.37 between general combining ability as measured by the yield of open-pollination progenies and that determined by the average yield of single crosses. Open-pollination progenies and S<sub>1</sub> progenies appeared to be of equal value in measuring the breeding potentialities of the parental clones. Bolton (1948) presented data to show that where large numbers of clones are to be tested, a relatively small number of tester plants may be used for making test crosses to be used in the estimation of combining ability.

Tysdal and Kiesselbach (1944) stressed that selection of parental clones at the start of a breeding program should be directed toward plants that are highly self-sterile. Eventual use of highly sterile materials in the production of  $F_1$  hybrids will insure maximum degree of outcrossing rather than selfing. Furthermore, it was believed that self-sterility is

associated with high cross-fertility. Using hybrid materials in the study, these authors found a correlation between the self-sterility of the hybrids and increased seed production resulting from crossing. Two of the most highly self-sterile hybrids showed the greatest increase in seed production when outcrossed. All hybrids showed a greater seed production when sibbed than when selfed, but self-sterile hybrids showed a greater increase when outcrossed than did the self-fertile hybrids. Other workers have obtained contradicting results. McAllister (1950) found that high self-fertility clones produced approximately three times as many seeds when selfed or crossed as did clones of low self-fertility, indicating a positive relationship between self- and cross-fertility. Bolton (1948) earlier had obtained a positive correlation (r =0.288) between self- and cross-fertility.

Tysdal and Kiesselbach (1944) presented evidence, from a relatively small population of selected plants, that selfsterility was associated with higher forage yield. Selection of plants high in self-sterility was suggested as a means of maintaining a high level of productivity. In contrast, Wilsie (1951) showed that self-fertility appeared to be relatively independent of combining ability. There was no correlation between self-fertility and open-pollinated progeny forage yields and only a slight correlation was obtained between selffertility and S1 progeny yields. The lack of association

between self-sterility and forage yield is further substantiated by the results obtained by Fryer (1939) and Pedersen (1953).

The method of producing double crosses in alfalfa proposed by Tysdal and Kiesselbach (1944) rests on the following main points: (1) selection of parental plants based on superior polycross progeny performance; (2) the use of selfsterility to ensure cross-pollination; (3) vegetative propagation of the parental plants in order to obtain sufficient amounts of single-cross seed; and (4) production of double cross seed.

Armstrong (1952) suggested the following for the commercial production of double-cross seed. Firstly, there should be available two superior lines possessing a sterility mechanism which is due to physiologic disturbances in the maturing pollen but which prove to be good combiners, and another two good combining lines that are self-incompatible but which produce an abundance of pollen. Secondly, the  $F_1$ generations (or single crosses) should be made within the types and not across the types, in order that self-sterility may be retained and crossing between related  $F_1$  plants be reduced to a minimum. The difficulty with this system would be in finding any four clones which would meet the aforementioned requirement.

An alternative plan to that proposed by Tysdal and

Kiesselbach (1944) was outlined by Bolton (1948) for the production of commercial hybrids of alfalfa. It suggested the use of "non-self-tripping, self-fertile" inbred plants in order to avoid the necessity for vegetative propagation. Crossing for the production of both single cross and commercial double cross seed will depend on insect pollination.

The use of synthetic varieties or seed of advanced generations of a double cross, or that originating from a composite of four or more parental clones with good general combining ability, has been highly successful. Tysdal <u>et al</u>. (1944, 1948) showed that clones selected for high combining ability by the polycross method produced a synthetic variety having a significantly higher forage yield than the standard variety or a synthetic produced using clones of poor-combining ability.

#### MATERIALS AND METHODS

## Plant Materials

In the present study the S<sub>1</sub> progenies used were obtained from four clones of alfalfa varying in the degree of selffertility as determined on the basis of previous observations. The clones may be described as follows:

- (1). Clone 585 B-2, which was obtained from the cross C84xV-2 and selected for alfalfa spotted aphid resistance. It was chosen to represent a high self-fertility group. The S<sub>1</sub> plants obtained from this clone were numbered 1 to 42 inclusive (Group I).
- (2). Clone 618, a second cycle selection from Synthetic 2187, was selected for resistance to <u>Cercospora</u> <u>medicaginis</u>. It was classified as having a low self-fertility. The S<sub>1</sub> plants were numbered from 101 to 142 (Group II).
- (3). Clone 631-101, a selection from Vernal, obtained from x-ray irradiation studies and selected for resistance to <u>Cercospora</u>. It was classified as intermediate in self-fertility but in this study it turned out to be of low self-fertility. The S<sub>1</sub> plants were assigned the numbers from 201 to 242 (Group III).

(4). Clone 5, a low-crown selection from an old golf course near Des Moines, Iowa. It was chosen to be representative of a low self-fertility group, but in this study proved to be intermediate in self-fertility. The S<sub>1</sub> plants were numbered from 301 to 342 (Group IV).

#### Procedure

A total of  $42 S_1$  plants, together with the parental clone, constituted each of the four groups. The  $S_1$  plants of each group were chosen at random from a larger  $S_1$  progeny growing in the breeding nursery. Stem cuttings were taken from the field in the fall of 1958 and were propagated in vermiculite in the greenhouse. When the cuttings had rooted, they were transplanted into 4-inch pots. Three cuttings were transplanted for each  $S_1$  plant. The growing medium used was of a sterilized mixture of 2 parts field soil, 1 part peat, and 1 part fine sand.

Each group or family of plants, was arranged on a single greenhouse bench in the order of the assigned number. Twelve plants of the parental clone also were included.

Nutrient solution was applied to the plants at monthly intervals. As the plants grew the elongating stems were supported by bamboo stakes held in an upright position. In order to accelerate flowering, the photoperiod was extended to 18 hours each day, using 200-watt incandescent lamps.

Determination of self-fertility of each of the S<sub>1</sub> plants and parental clones was made by artificial selfing of at least 300 flowers. Selfing of flowers was accomplished by applying pressure on the keel (which holds the staminal column of the flower) with the tip of a toothpick, and drawing the tip across the exposed stigma. The number of flowers that were selfed, and the date of selfing were indicated. Selfing was performed whenever flowers were available, over a period of time in winter and spring of 1958-59 and 1959-60.

In order to study sib-mating compatibility relationships, cyclic crosses (e.g.  $1 \ge 2, 2 \ge 3, 3 \ge 4 \cdots 42 \ge 1$ ) among the 42 S<sub>1</sub> plants of each group were made by hand pollination. There were 42 cross combinations for each group and including reciprocal crosses, a total of 84 different crosses were actually involved. In addition, check crosses with 2 unrelated clones also were made.

For each cross, ten flowers were used. Two racemes, borne on the same plant, were selected and only 5 flowers from a raceme were used, the rest of the flowers being clipped off.

In making a cross the standard petal of each flower used as female was first cut off close to the base, after which the flowers were tripped. An adequate quantity of pollen was collected from the exposed staminal column of each plant used

as a parent. Emasculation of flowers was achieved by suction, using a finely drawn glass tube connected to an electricallydriven vacuum pump. Reciprocal crosses were made, pollen being applied immediately after emasculation. The racemes bearing the hand pollinated flowers were appropriately tagged.

In each group four series of cyclic crosses were made, usually at intervals of from 7 to 10 days. Each series of crosses served as a replicate and in order to keep environmental variability as low as possible all crosses in a series were completed within a period of one or two days. To provide a measure of the differences in seed setting between days during which a series was completed, check crosses were made each day. After making each series of sib crosses, the pots were re-arranged on the bench in order to eliminate the possible positional effects of the pots on seed-setting. Repotting of plants within the entire group was done whenever necessary in order to complete the desired number of replicated crosses.

Each S<sub>1</sub> plant in each group also was backcrossed to the parental clone, the latter being used as the pollinator. Four replicated series of backcrosses were made. A replication was completed in one day.

The dates during which all the crosses were made are indicated as follows:

Group		Rep I	Rep II	Rep III	Rep IV	
A. Cyclic crosses						
Group I	(1-42)	3/7/59	3/17-18/59	4/9 <b>-1</b> 0/59	4/23-24/59	
Group II	(101-142)	3/24 <b>-</b> 25/59	3/18-19/60	-	-	
Group III	(201-242)	4/15-16/59	6/22-23/60	-	-	
Group IV	(301-342)	3/12/59	4/2-3/59	4/8-9/60	4/12-13/60	
B. Backcrosses						
Group I	(1-42)	1/9/60	1/16/60	4/4/60	4/6/60	
Croup II	(101-142)	4/11/60	-	-	-	
Group III	(201-2l <sub>1</sub> 2)	3/23/60	3/26/60	4/2/60	4/5/60	
Group IV	(301-342)	1/23/60	1/30/60	4/15/60	4/16/60	

Pods were harvested when fully mature, usually about four weeks after selfing or crossing. The total number of pods formed was recorded, and when the pods were threshed the total number of seeds set was determined by counting only those that were well-filled.

For comparative purposes, the fertility data were expressed on the basis of the number of seeds set per flower pollinated.

### Results of Selfing

The results of selfing of the  $S_1$  progenies and  $S_0$  parental clones used in this study are shown in Tables 1, 2, 3 and 4, in the Appendix. The self-fertility data for the parental clones obtained in the present study did not coincide exactly with data obtained previously.

Table 1. Self-fertility of four S<sub>o</sub> clones of alfalfa as determined in two separate experiments

C.	1956-1957		1958-1960	
S <sub>o</sub> parental clone	Seeds per flower selfed	Self-fertility classification		Self-fertility classification
585 B-2 C1	2.20	High	2.582	High
618 CI	0.20	Low	0.498	Low
631 <b>-1</b> 01 Cl	1.50	Intermediate	0.538	Low
5 01	0.24	Low	0.910	Intermediate

Considering the average self-fertility of the  $S_1$ progenies and corresponding non-inbred parents, the four groups of alfalfa plants used in the present study may be reclassified as follows: Group I (1-42) as a high self-fertility group; Group II (101-142) and Group III (201-242) as low self-fertility groups; Group IV (301-342) as intermediate in self-fertility.

Generally, there was a marked reduction in self-fertility among the  $S_1$  plants as compared with the self-fertility of the  $S_0$  parental clone (Table 2b). The self-fertility of the  $S_1$ plants in Groups I, II, III and IV, expressed as an average of the group, was 0.482, 0.051, 0.106 and 0.246 seed per flower tripped, respectively. The self-fertility of the parental clone of each group, in the same order, was 2.582, 0.498, 0.538 and 0.910 seed per flower selfed.

There was a remarkable range in self-fertility among the  $S_1$  plants within a group. This is especially true among those in the high and intermediate self-fertility groups (Table 2a). It is of interest to note that one  $S_1$  plant in Group III and two  $S_1$  plants in Group IV had a self-fertility higher than that of their non-inbred parents.

The most drastic reduction in self-fertility occuring in the low fertility groups seems to indicate that the parental clones for these groups had already a preponderance of the genes associated with the expression of self-incompatibility, and that the intensity of self sterility is increased after one generation of inbreeding, owing to the fact that the individuals were approaching homozygosity.

In the high self-fertility group, 18 S<sub>1</sub> plants out of 42 studied were observed to be autogamous or capable of

Classes (Seed per	Frequency distribution of S <sub>1</sub> plants				
flower selfed)	Group I (1-42)	Group II (101-142)	Group III (201-242)	Group IV (301-342)	
$\begin{array}{c} 0.000 & = & 0.050 \\ 0.051 & = & 0.100 \\ 0.101 & = & 0.200 \\ 0.151 & = & 0.200 \\ 0.201 & = & 0.250 \\ 0.251 & = & 0.300 \\ 0.301 & = & 0.350 \\ 0.351 & = & 0.400 \\ 0.401 & = & 0.450 \\ 0.451 & = & 0.500 \\ 0.501 & = & 0.550 \\ 0.551 & = & 0.600 \\ 0.651 & = & 0.550 \\ 0.651 & = & 0.700 \\ 0.651 & = & 0.700 \\ 0.651 & = & 0.700 \\ 0.751 & = & 0.800 \\ 0.800 & = & 0.850 \\ 0.851 & = & 0.900 \\ 0.951 & = & 1.000 \\ 1.001 & = & 1.050 \\ 1.051 & = & 1.100 \\ 1.101 & = & 1.150 \\ 1.151 & = & 1.200 \\ \end{array}$	25 54631222 12 21	28 6 2 1 1	17 8 7 3 2 2 1	2 8 8 8 3 4 1 2 1 1 1 1 1 1 1 1	
1.201 - 1.250 1.251 - 1.300 1.301 - 1.350	2				
$1.351 - 1.400 \\ 1.401 - 1.450 \\ 1.451 - 1.500 \\ 1.501 - 1.550 \\ 1.551 - 1.600$	1				
Total	lµ2	40	41	42	

Table 2a. Frequency distribution of S<sub>1</sub> plants on the basis of self-fertility as measured by the number of seeds per flower selfed

Average self- fertility	Group I (1-42)	Group II (101-142)	Group III (201-242)	Group IV (301-342)
Parental S <sub>o</sub> clone	2,582	0.498	0.538	0.910
S <sub>l</sub> plants	0.482	0.051	0.106	0.246
Average S <sub>1</sub> sel fertility ex- pressed as a percentage of self-fertilit the parental clone	the y of	10 <b>.</b> 24	19.70	27.03

Table 2b. Average self-fertility of  $S_0$  and  $S_1$  plants

setting seeds without the aid of artificial tripping. The parental clone of this group possessed the same autogamous characteristic. Practically all the plants in this group produced a large number of flowers per plant and an abundance of pollen. In Group II, the group lowest in self-fertility, eleven  $S_1$  plants out of a total of 42 included were observed to have produced a very small quantity of pollen. Generally, the  $S_1$  plants produced limited numbers of flowers per plant. In Group III, a low self-fertility group, four plants did not produce adequate pollen. Flower production was not a problem. In Group IV, five  $S_1$  plants produced very little pollen but the others were good pollen producers. Flowers were borne in large numbers except for two plants.

#### Results of Cyclic Sib-mating

The data on seed setting following sib-mating in the high fertility group are presented in Table 5 in the Appendix. Cross-fertility among sib-matings ranged from 0.125 to 2.550 seeds per flower crossed. The analysis of variance is shown in Table 3a. Significant differences in seed set were obtained among sib-crosses as shown by the significance of the mean square for crosses at the 1 per cent level. Evidently, genetic factors are involved which determine the compatibility relationship of any two sib lines. It is believed that the S<sub>1</sub> individuals were segregating and were therefore different in genetic make-up for factors determining self- or cross-incompatibilities. Using the L.S.D. value, comparisons were made in seed set of two sib-crosses involving a common female parent but differing in the pollen source. Out of

Table	3a.	Analysis of variance for seed setting obtained
		from sib-crosses among S <sub>1</sub> plants in Group I

Source of variation	Degree of freedom	Mean sq <b>uare</b>	Computed F- value
Crosses	85	134.35	6.71**
Replications	3	42.82	2.14
Error	255	20.03	

\*\* Exceeds the 1 per cent level

Source of variation	Degree of freedom	Mean square	Computed F- value
Sib-crosses	83		
Cross combinations	41	152.29	3.26**
Reciprocals	1	2.86	
Combinations x reciprocals	41	46.75	2.55**
Error	249	18,32	

Table 3b. A partitioning of the sum of squares for seed yield among sib-crosses

\*\* Exceeds the 1 per cent level

42 possible sets of comparisons, two showed a statistically significant difference at 5 per cent level, whereas six pairs showed significant difference at the 1 per cent level. Nonsignificance obtained in the remaining sets of comparisons indicates a similarity in the compatibility factors contained by the fertilizing pollen of the two sources involved. Similarly, comparisons in seed set were made between two sibcrosses involving two different female plants hand-pollinated with pollen coming from the same plant. Ten pairs of comparisons showed significant differences at the 5 per cent level whereas four sets had differences significant at the 1 per cent level. The results obtained point to the fact that cross compatibility is dependent on the genotype of both the maternal and paternal lines which complement each other. Statistical differences between reciprocal crosses were obtained in eight cases, out of 42 sets of reciprocal crosses, five of which had a significance at 1 per cent level. A significant difference was also obtained between reciprocals of the check crosses. It would seem that in some combinations, the order of pairing in making the cross is very important.

It is of interest to note that eight compatible sibcrosses even produced as much seed as that of a check cross which involves two unrelated  $S_0$  clones. While many crosses were highly compatible, other crosses, on the other hand, were not as compatible and produced very limited amounts of seed.

The mean square for replication is non-significant. The four replicated series of crosses were all made in the spring of 1959.

The sum of squares representing the sib-crosses is further partitioned into orthogonal comparisons (Table 3b). The between-reciprocals component represents two groups, one of which includes the crosses in which all the 42 S<sub>1</sub> plants were used as female parents while the other contains all crosses involving the same 42 S<sub>1</sub> plants but used as pollen sources. The sib-crosses in one group are therefore the reciprocals of the entries in the other group. The new error term used is also a re-calculated value after eliminating the entries for

the check crosses.

The mean square for cross combinations is significant at the 1 per cent level. A combination represents the average seed set of its reciprocal crosses. Certain combinations were certainly more compatible than others. The mean square for combinations x reciprocals interaction is also significant at the 1 per cent level, indicating that, generally, the same cross combination did not perform consistently in reciprocal crosses.

The average cross-fertility for the sib-crosses is 0.970 seed per flower crossed. The average self-fertility of the  $42 S_1$  plants and parental clone is 0.482 and 2.582 seeds per flower, respectively. Although a comparison of seed setting between sibbing and selfing may not be statistically valid. by virtue of the fact that the averages were obtained from two separate experiments, yet there is reason to believe that the process of sib-mating in this particular group would result in a greater quantity of seed setting in comparison to straight selfing. In the first generation of selfing the majority of the plants would have obtained some degree of duplication for the genetic factors associated with the selfincompatibility mechanism and selfing of these plants would result in a reduction in seed setting, as has been actually demonstrated. The S, population, however, is still highly heterogeneous so that sib-mating among plants, due to

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complementary gene action, would overcome such sterility problems accruing from straight selfing.

The results of the sib-mating study in Group IV are presented in Table 8 in the Appendix. The analysis of variance is shown in Table 4a. The degrees of freedom are diminished by 5, a figure corresponding to the number of missing data. It will be noted that the mean square for crosses is significant at the 1 per cent level. Cross-fertility ranges from 0.075 to 3.175 seeds per flower crossed. It is indicated that while some sib-crosses are highly compatible, other combinations are not. Eight sib-crosses gave as much seed set as one of the reciprocals of the check crosses. Comparisons are made between two sib-crosses in which a common female parent and two different male parents are involved. Comparisons are made also between crosses involving the same pollen source but differing in the seed parents. Six and fourteen sets of comparisons, respectively, showing statistically significant differences were obtained. A comparison between reciprocal crosses showed that five pairs of reciprocals showed significant differences in seed setting.

The mean square for replication also is statistically significant at the 1 per cent level. Crosses made in the first two replications were made in the spring of 1959 whereas the last two replications were completed in the spring of 1960. The plants grown in 1960 were derived from cuttings of

Source of variation	Degree of freedom	Mean square	Computed F- value
Crosses	85	328.00	7.15***
Replications	3	2,846.81	62.05***
Error .	250	45.88	

Table 4a. Analysis of variance for seed setting obtained from sib-crosses among S<sub>1</sub> plants in Group IV

\*\*\* Exceeds the 1 per cent level

Table 4b. A partitioning of the sum of squares for seed yield among sib-crosses

Source of variation	Degree of freedom	Mean square	Computed F- value
Sib-crosses	83		
Cross combinations	41	405.29	7.66***
Reciprocals	1	12.96	
Combinations x reciprocals	41	52.87	1.36
Error	244	38.86	

\*\* Exceeds the 1 per cent level

plants grown in 1959. Twice as much seed setting was obtained in 1960 as compared to that obtained in 1959. It is indicated that the amount of seed setting following sib-mating is subject also to fluctuations owing to differences in environmental conditions occuring at the time crosses are made. The conditions in 1960 were more favorable to seed setting.

The sum of squares for crosses was broken down into three components (Table 4b). The mean square for combinations is statistically significant at 1 per cent level. The combinations x reciprocals component has a non-significant mean square. The results of the statistical analysis show that certain combinations are more compatible than others. The relative degree of seed setting among cross combinations was consistent in reciprocal crosses.

The average amount of seed set among the sib-crosses was 1.061 seeds per flower crossed. The average self-fertility of the 42 S<sub>1</sub> progenies and the S<sub>0</sub> parent was 0.246 and 0.910 seeds per flower, respectively. Apparently, loss of fertility is much slower following sib-mating than when selfing is per-formed.

Cyclic sib-crosses also were made for the S<sub>1</sub> plants belonging to Groups II and III which are both classified as low in self-fertility. The results are presented in Tables 6 and 7 in the Appendix. Only two replicated series of sibcrosses were made in the two groups since the amount of seed setting was found to be exceedingly low, from the practical point of view. The results obtained indicate that no additional advantage could be attained in sib-mating among self lines which are inherently very poor in self-fertility. The

problem encountered in making sib-crosses in Group II was the limited amount of pollen available from each S<sub>1</sub> plant. In making the actual crosses, however, pollen was collected in adequate amount, enough to cross-pollinate each female flower. Failure in seed setting was therefore the result of the incompatibility relationships between sibs rather than the lack of adequate cross-pollination.

The S<sub>1</sub> plants in Group III, although including a few plants which produced pollen in limited quantity, were, on the whole, better pollen producers than the plants in Group II. Seed setting in the sib-crosses also was exceedingly low.

It is believed that the populations of Groups II and III were homogeneous to an extent and were composed of  $S_1$  plants the majority of which were identical in the factors associated with self- or cross-incompatibility. Results of sib-mating therefore would be little different from those obtained by selfing.

# Results of Backcrossing

Table 9 in the Appendix shows the result of backcrossing of the  $S_1$  plants in Group I (1-42) to the parental clone from which they were derived. Seed setting ranges from 0.950 to 4.675 seeds per flower, with an average of 3.012 seeds. The analysis of variance is shown in Table 5. The mean square for crosses is statistically significant at the 1 per cent

level indicating that the S<sub>1</sub> progenies were different in the factors controlling fertility. The mean square for replications also is highly significant, showing that seed setting is partly dependent on environmental factors.

The analysis of variance for seed setting obtained in the backcrossing of the  $S_1$  progenies in Group III (201-242) is presented in Table 6. Significant differences in seed setting are obtained between backcrosses as shown by the statistically significant mean square. The range in seed set is from 0.70 to 3.625 seeds per flower, with an average of 2.131 seeds (Table 11, Appendix).

The analysis of variance for Group IV (301-342) is shown in Table 7. Statistical differences in seed set among backcrosses are obtained. Differences among replications are also

Source of variation	Degree of freedom	Mean square	Computed F- value
Crosses	39	276.14	11.31**
Replications	3	126.45	5.18**
Error	115	24.41	

Table 5. Analysis of variance for seed setting following backcrossing of S<sub>1</sub> plants to parent clone in Group I

\*\* Exceeds the 1 per cent level

Table 6. Analysis of variance for seed setting following backcrossing of S<sub>1</sub> plants to parent clone in Group III

Source of variation	Degree of freedom	Mean square	Computed F- value
Crosses	39	264.19	7.50**
Replications	3	33.06	0.94
Error	116	35.22	

\*\*\*Exceeds the 1 per cent level

Table 7. Analysis of variance for seed setting following backcrossing of S<sub>1</sub> plants to parent clone in Group IV

Source of variation	Degree of freedom	Mean square	Computed F- value
Crosses	39	550.51	13.09**
Replications	3	350.25	8.33**
Error	117	42.06	

\*\*Exceeds the 1 per cent level

statistically significant. The range in cross-fertility is from 0.950 to as high as 5.900 seeds per flower, the average being 2.793 seeds.

In Group II (101-142) only one replication of backcrosses is available. The average cross-fertility of the group is 0.813 seed per flower.

It is of interest to note that in all cases, the average seed set of a group following backcrossing exceeds that of selfing of the corresponding  $S_0$  parental clone (Table 8). There is indication that improvement in fertility among the  $S_1$  plants may be achieved by crossing them back to the parental clone from which they were derived.

## Correlation Studies

Correlation studies made for the  $S_1$  plants in Group I are shown in Table 9. As can be noted, the self-fertility of the  $S_1$  plants is correlated with the fertility of the same plants following backcrossing. It shows that both procedures were equally good in measuring genetic differences relating

Table 8. Seed set from selfing  ${\rm S}_{\rm O}$  parental clones and back-crossing  ${\rm S}_{\rm l}$  plants to parental clones

Numbe	Number of seeds per flower crossed or selfed					
	Group I	Group II	Group III	Group IV		
Self-fertility of parental clone	2.582	0.498	0.538	0.910		
Average cross- fertility of the S <sub>1</sub> plants back- crossed to parental clone	3.012	0.813	2.131	2.793		

to fertility among the  $S_1$  plants. Self-fertility and backcross fertility both are associated with the average crossfertility of the clones as male and female parents. There is, however, a lack of association when each of the  $S_1$ clones was considered only as a male parent in the process of sib-mating.

Table 9. Correlations of seed setting following selfing, backcrossing, and sib-mating of  $42 S_1$  plants in Group I (1 to 42).

	Eelf-fertility	Fertility following backcrossing	Fertility fol- lowing sib- mating (S <sub>1</sub> plants in- volved as fe- male parents)
Fertility following backcrossing	0.3702*		
Fertility following sib-mating (S <sub>1</sub> plan involved as female and male parents)	ts 0.3540 <sup>%</sup>	0.3810*	
Fertility following sib-mating (S <sub>1</sub> plan involved as female parents only)	ts 0.3285 <sup>*</sup>	0.5183*	
Fertility following sib-mating (S <sub>1</sub> plan involved as male parents only)	0.2843	0.1524	0.5063**

\*Exceeds the 5 per cent level

The correlation studies made for Group IV S<sub>1</sub> plants are presented in Table 10. The correlation value obtained between selfing and backcrossing is statistically non-significant, although it is just a little lower than the tabular value at the 5 per cent level. But there is a good association between self- and average sib-cross-fertility (as

Table 10. Correlations of seed setting following selfing, backcrossing, and sib-mating of 42 S<sub>1</sub> plants in Group IV (301-342)

	Self-fertility	Fertility following backcrossing	Fertility fol- lowing sib- mating (S <sub>1</sub> plants in- volved as fe- male parents)
Fertility following backcrossing	0.3042		

u u			
Fertility following sib-mating (S <sub>1</sub> plants involved as female and male parents)	0.4159***	0 <b>.</b> 6257**	
Fertility following sib-mating (S <sub>1</sub> plants involved as female parents only)	0.4861**	0.5671**	
Fertility following sib-mating (S <sub>1</sub> plants involved as male parents only)	0.2966	0.2506	0 <b>.</b> 7780 <sup>**</sup>

\*\*\*Exceeds the 1 per cent level

maternal and paternal parents) as shown by the significance of the computed r at 1 per cent level. A similar relationship in fertility between that of backcrossing and sib-mating is obtained. Statistically non-significant correlation values were obtained when each of the  $S_1$  clones was considered only as a pollen parent in the process of sib-mating.

In Group III, a significant correlation was obtained in seed setting between selfing and backcrossing, the r value being 0.5297.

The correlation values obtained seem to indicate that S<sub>1</sub> plants showing high self-fertility may generally be found to possess a high degree of backcross- or sib-cross-fertility preferably when used as female parents. This positive relationship between self- and cross-fertility is clearly demonstrated in Tables 11 and 12.

Table 11. Average self-, backcross-, and sib-cross-fertility (number of seeds per flower) of three sub-groups of 42 S<sub>1</sub> lines of Group I classified on the basis of self-fertility

Sub-group	Average self- fertility	Average backcross- fertility	Average sib- cross-fer- tility
Low self-fertili	lty 0.175	2.828	0.767
Intermediate sel fertility	Lf- 0.360	2.713	0.906
High self-ferti]	Lity 0.910	3.511	1.238

Table 12.	Average self-, backcross-, and sib-cross-fertility (number of seeds per flower) of three sub-groups of 42 S <sub>1</sub> lines of Group IV classified on the basis of self-fertility

Sub-group	Average self- fertility	Average backcross- fertility	Average sib- cross-fer- tility
Low self-fertil:	ity 0.076	2.046	0.811
Intermediate set fertility	lf- 0.173	2.980	1.084
High self-ferti	lity 0.487	3.338	1.286

### DISCUSSION

The low seed set obtained by selfing the  $S_1$  plants used in this study showed that a marked reduction in self-fertility had taken place through one generation of self-fertilization. Self-sterility was most pronounced in the two families derived from low self-fertility clones. Although a drastic loss in self-fertility had occurred in the  $S_1$  progenies of the high and intermediate groups, it is believed that a majority of the  $S_1$  plants could be selfed successfully at least one more generation.

Differences in seed setting among plants in inbred progenies, produced by selfing, backcrossing, and sib-mating indicated segregation of hereditary factors controlling fertility. Selection in the first and subsequent generations of selfing for individuals that are high in self-fertility, if this were desirable, might prove to be effective. Kirk (1927) recognized earlier the effectivity of selection for improved seed yield in the self generations of alfalfa. Bolton (1948) obtained one third generation line yielding over 60 pounds per acre.

Sib-mating is an inbreeding method which might be used as an alternative to straight selfing. As shown by Wright (1921) the approach toward homozygosis is somewhat slower through sib-mating than through selfing, and consequently the drastic drop in self-fertility characteristic of selfing might

be partially prevented or at least delayed. In the two low self-fertility groups in this study, however, sib-mating proved to be impractical because of a high degree of sib-incompatibility. On the other hand, sib-mating among  $S_1$  plants in the high and intermediate self-fertility groups resulted in a significant gain in fertility. A study made by Koffman (1959) has shown similar results.

Restoration of fertility following sib-mating rests on the following assumptions. As a result of selfing, plants tend to become more homozygous for the genetic factors associated with the expression of self-incompatibility, and may, therefore, be self-incompatible. However, the S<sub>1</sub> population undoubtedly is still heterozygous for many factors so that many of these self-sterile plants, with the operation of complementary and modifying factors, would still be capable of producing seeds when cross-fertilized by compatible sibs from the same population.

There are two observable components of self-incompatibility operating in alfalfa. One is associated with the inability of the pollen tube to reach the ovule and effect fertilization, as Cooper and Brink (1940), Brink and Cooper (1938), and Miller (1960) have observed. The other factor is that involving post-fertilization ovule collapse, as has been reported by Brink and Cooper (1938, 1939), Cooper and Brink (1940), and Miller (1960). The genetics involved in the two cases are not

understood at the present time. But it seems, as the present data suggest, that there are actually "plus" genes directly controlling fertility. This may be evidenced from the correlation studies made, which show that self-fertility of the  $S_1$  plants is positively associated with average backcrossand sib-fertility. A positive relationship between self- and cross-fertility has previously been shown by McAllister (1950) and Bolton (1948). Such significant correlations would unlikely be brought about by the operation of the S incompatibility gene systems. Whether or not such "plus" genes are related directly to the expression of embryo abortion is not known. It is believed that factors related to the production of functional gametes may have accounted for the positive association between self- and cross-fertility.

As the present correlation data indicate the degree of fertility is somewhat dependent on the maternal parent. Significant correlations were obtained in the  $S_1$  between selffertility and sib-fertility when the plants were utilized as maternal parents in making the crosses. This may explain why in some cases, reciprocal sib-crosses varied considerably in seed setting. Whitehead and Davis (1954) ascribed the variation in seed setting between reciprocal crosses to the differences in female fertility between the parental plants. It was postulated that alfalfa plants may differ in the number of highly functional ovules, a factor which would determine

female fertility. In Lotus sp., Bubar (1958) found that uniformity in the development of ovules within each ovary determines the self-fertility of a plant. The self-fertile plants were found to have a greater uniformity in the development of the ovules within each ovary than the self-sterile individuals.

From the 42 S<sub>1</sub> plants constituting a group, a total of 861 cross combinations are possible. The 42 cross combinations obtainable in the present experiment may be assumed to be a representative sample of the said theoretical number. Assuming that the 42 self plants represent a true population originating from a selfed non-inbred clone, it follows that the results in seed setting obtained in sib-mating would be similar to those obtainable when sib-mating occurs at random in the field. One point that should not be overlooked, however, is that seed setting under greenhouse conditions may not be essentially similar to that under natural field conditions. Seed setting would be influenced by the environmental conditions especially that of temperature and moisture prevailing in the area, and by the activity of pollinating insects.

Sib-mating as has been noted, is a form of inbreeding much slower than selfing in attaining homozygosity. This is especially true in alfalfa since it is a tetraploid. Eartlett and Haldane (1934) have estimated that in a tetraploid selfing must be done for 3.80 generations as compared to one generation

for a diploid, in order to halve heterozygosis. With brothersister matings, the number of generations of selfing is 8.72 and 3.26, respectively. Furthermore, alfalfa is characterized by a highly heterozygous nature maintained by the prevalent outbreeding system. Many generations of inbreeding, which may not be possible, would be required to produce approximately homozygous or true-breeding inbred lines. Stewart (1934), however, has observed remarkable uniformity in many observable characters after one generation of selfing.

Parent-progeny crosses made in this study have shown interesting results. There was a consistent and relatively high seed set in each of the groups studied resulting from backcrossing of each S1 individual to the parent from which it was derived, in comparison with that of sib-mating. Average cross-fertility in these S, backcrosses even exceeded the self-fertility of the respective non-inbred clones and approached the level of cross-fertility attained in outcrossing parent clones to an unrelated tester plant. No genetic explanation can be offered at this point. It should be remembered, however, that in making crosses only five flowers to a raceme were used as opposed to the greater number of flowers per raceme utilized in the process of selfing and the greater fertility indicated may be due, in part at least, to physiologic rather than genetic reasons. Furthermore, there were indications that the period during which all backcrosses were

made was favorable for seed setting.

Using the method of path coefficients (Wright, 1934), the coefficient of relationship between the S, individual and the non-inbred parental clone during the first backcrossing and that between full sibs may be computed and the two values should be equal. Assuming that a linear relationship exists between fertility and the degree of inbreeding, as previously shown by Fyfe (1957), seed setting in the two systems of mating should be the same. Yet seed setting resulting from backcrossing was higher than that following sib-mating. It should be noted though that in the case of backcrossing the pollen came from a non-inbred. There is reason to believe that the loss of fertility accompanying inbreeding in alfalfa is not only due to the action of the genetically controlled selfincompatibility mechanism but also to a consequent marked reduction in male potency or general level of vigor.

The possibility of using the backcross progenies especially among those involving self plants whose self-fertility is exceedingly low, as a starting point in an inbreeding program, deserves further study.

In the light of the preliminary results obtained in the present study, a procedure with respect to inbreeding of alfalfa is presented. A combination of selfing and sib-mating may be adopted in the inbreeding program. The objective would be to isolate reasonably homozygous lines which at the same

time possess some degree of fertility when selfed or sib-mated. Highly self-fertile non-inbred clones are initially selected from a population. These clones are selfed and  $S_1$  plants are grown. The  $S_1$  plants are artificially selfed, assuming that a procedure for extensive selfing is feasible, and seeds obtained from those with relatively high self-fertility are used for growing the next generation. Selfing and selection for high seed setting probably could be continued for a few subsequent generations depending on the self-fertility behavior of the self progenies. After this point, sib-mating among members of a line is followed. Sib-mating may be started early in the first self-generation if further selfing leads into sterility problems.

The possibility of restoring fertility by making intergeneration crosses among self plants may be further explored. This idea is based on the results obtained in the present backcross study of the low self-fertile groups. Self progenies in an advanced generation having low cross-fertility may be crossed to some related high self-fertile plants obtainable in the immediate preceding generation. If this is economically possible, self-fertility may be restored without sacrificing much for the loss of homozygosity in the other characters. Further study is needed along this line.

At the end of the inbreeding program isolated lines must have a restored fertility when either selfed or sib-mated.

These lines are then tested for combining ability, and those which combine well to give a high level of heterosis could be further evaluated as experimental  $F_1$  hybrids.

The procedures just discussed are applicable only where plants do not exhibit an extreme degree of sterility during the initial stages of inbreeding. In the two families studied, which were derived from clones of low self-fertility, one generation of selfing resulted in a drastic loss of fertility, and sib-mating used as a means of restoring fertility proved to be ineffective. If such is the case, then it would seem that a program of inbreeding would exclude those clones which are low in self-fertility. As previously shown by Wilsie (1951) and McAllister (1950) these may constitute as much as 30 to 40 per cent of a non-inbred population.

### SUMMARY AND CONCLUSIONS

A study was undertaken to consider the possibility of using advanced generations of inbred lines of alfalfa for commercial hybrid seed production. Four groups of  $S_1$  plants were used, one of high, one of intermediate, and two of low self-fertility. Selfing, sib-mating and backcrossing of  $S_1$ plants were done concurrently and a comparison of seed setting was made among these systems of mating, on the basis of the number of seed per flower selfed or crossed.

1. There was a marked reduction in self-fertility among the  $S_1$  plants in all groups. Expressed as a percentage of the self-fertility of the non-inbred parental clone, the average self-fertility of the high, intermediate, and two low self-fertility groups was 18.67%, 27.03%, 19.70% and 10.24%, respectively. There was a considerable range in selffertility within families in the first two groups.

2. Sib-mating was achieved by making cyclic crosses among the  $S_1$  plants. There was a considerable gain in fertility following sib-mating over that of selfing in the high and intermediate fertility groups. Seed setting resulting from sibbing was 2.01 and 4.31 as much as that of selfing in the two respective groups. The gain in fertility obtained in the two low-fertility groups was considered insignificant, for practical purposes.

3. Analysis of variance in the sib-crosses made for the

high and intermediate fertility groups showed statistical differences at the 1 per cent level in seed set among crosses and among cross combinations. There was a wide range of variation in seed setting among the sib-crosses and in some cases it reached the level of cross-fertility of two unrelated non-inbred clones. Factors for fertility were segregating in the  $S_1$  population and seed setting was conditioned by the compatibility relationship of any two self lines. Also, in some instances reciprocal crosses differed significantly in fertility.

4. Results of backcrossing showed statistical differences at the 1 per cent level in cross-fertility among the  $S_1$  plants. There was a consistent increase in seed setting resulting from backcrossing over that of sib-mating in all the groups. Seed setting was even as high, if not higher, than that of the corresponding selfed non-inbred parental clone.

5. The greater fertility obtained following backcrossing over that of sib-mating, despite the fact that the two systems represented the same degree of inbreeding, indicated that loss of fertility accompanying selfing had been the result not only of the operation of the genetic self-incompatibility mechanism but also of the consequent marked reduction in male potency or general level of vigor.

6. Statistically significant correlation values were

obtained either at the 5 or 1 per cent level, except for one, between self-fertility and cross-fertility following sibmating or backcrossing. The positive association was believed to have been brought about by the inherent differences among the  $S_1$  plants in the production of highly functional gametes, especially in the ovules. The factors involved are thought to be independent of those associated with the multiple oppositional allele series.

7. Consideration is given to the possibility of undertaking an inbreeding program for alfalfa based on techniques which provide a slower rate of inbreeding than that of continued selfing.

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## APPENDIX

Plant number	Number of flowers tripped	Number of pods that developed	Number of seeds that set	Seeds per flower tripped
1 234567890 11234567890 11234567890 11234567890 11234567890 332334567890 412585 8-2 Average	348 378 340 452 388 518 679 564 352 0 522 401 475 3720 5228 611 477 522 401 477 522 401 477 522 402 550 557 557 4366 557 557 4366 459 557 557 4366 459 557 557 4366 459 557 557 4366 459 557 557 557 4366 459 557 557 557 557 557 557 557 557 557 5	146 24 140 257 173 331 205 18 211 12 203 205 191 131 203 205 191 131 53 149 166 210 204 77 111 124 310 207 133 163 223 195 245 194 1592 195 195 195 195 195 195 195 195 195 195	$     \begin{array}{r}       151 \\       25 \\       112 \\       451 \\       218 \\       826 \\       158 \\       123 \\       439 \\       99 \\       246 \\       286 \\       310 \\       169 \\       58 \\       146 \\       146 \\       225 \\       283 \\       327 \\       86 \\       133 \\       156 \\       657 \\       188 \\       475 \\       203 \\       393 \\       137 \\       110 \\       199 \\       65 \\       211 \\       71 \\       93 \\       188 \\       726 \\       42 \\       152 \\       92 \\       86 \\       2,574 \\     \end{array} $	0.434 0.066 0.329 0.998 0.562 1.594 0.233 0.218 1.240 0.137 0.473 0.548 0.759 0.345 0.345 0.399 0.2548 0.398 1.220 0.398 1.227 0.398 1.229 0.398 1.240 0.345 0.398 1.2218 0.398 1.229 0.398 1.229 0.398 1.229 0.398 1.2275 0.398 0.2758 0.318 0.116 0.578 0.318 0.118 0.275 0.217 0.210 2.582 0.482

Table 1. Self-fertility of S1 plants in Group I

		T		
Plant number	Number of flowers tripped	Number of pods that developed	Number of seeds that set	Seeds per flower tripped
101 102 103 104 105 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120 121 123 124 125 126 127 128 129 130 131 132 133 134 135 136 137 138 139 140 141 142 618 C1 Average	830 189 425 52 302 454 102 252 285 416 450 408 134 - 239 358 789 243 346 124 134 - 239 358 789 243 346 124 137 232 281 59 432 272 487 15 357 272 334 452 334 455 272 3324 455 272 3324 455 272 3324 455 272 3324 455 272 324 59 455 272 324 59 455 272 324 59 455 272 324 455 201 172 201 172 201 172 201 172 201 172 201 172 201 172 201 172 201 172 201 172 201 172 201 172 201 172 201 172 201 172 201 172 880 of the S1 plant	$\begin{array}{c} 93\\ 69\\ 0\\ 9\\ 12\\ 1\\ 7\\ 2\\ 83\\ 22\\ 0\\ 4\\ -\\ 4\\ 10\\ 15\\ 0\\ 321\\ 0\\ 35\\ 6\\ 329\\ 0\\ 1\\ 0\\ 15\\ 17\\ 16\\ 43\\ 18\\ 0\\ 295\\ s\end{array}$	$     \begin{array}{r}             87 \\             2 \\             72 \\           $	$\begin{array}{c} 0.105\\ 0.011\\ 0.169\\ 0.000\\ 0.033\\ 0.020\\ 0.020\\ 0.020\\ 0.020\\ 0.020\\ 0.020\\ 0.020\\ 0.020\\ 0.020\\ 0.000\\ 0.049\\ 0.000\\ 0.030\\ -\\ -\\ 0.017\\ 0.034\\ 0.000\\ 0.030\\ 0.009\\ 0.000\\ 0.069\\ 0.000\\ 0.069\\ 0.000\\ 0.069\\ 0.000\\ 0.069\\ 0.000\\ 0.069\\ 0.000\\ 0.069\\ 0.000\\ 0.069\\ 0.000\\ 0.069\\ 0.000\\ 0.069\\ 0.000\\ 0.069\\ 0.000\\ 0.069\\ 0.000\\ 0.025\\ 0.067\\ 0.055\\ 0.000\\ 0.033\\ 0.044\\ 0.117\\ 0.055\\ 0.000\\ 0.000\\ 0.098\\ 0.051\\ \end{array}$

Table 2. Self-fertility of S<sub>1</sub> plants in Group II

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Plant number	Number of flowers tripped	Number of pods the developed	t seeds	Seeds per flower tripped
	$\begin{array}{c} 643\\ 472\\ 792\\ 727\\ 517\\ 560\\ 8\\ 8\\ 991\\ 756\\ 8\\ 8\\ 991\\ 776\\ 8\\ 9997\\ 760\\ 5\\ 6\\ 9997\\ 560\\ 570\\ 16\\ 9997\\ 560\\ 570\\ 16\\ 4\\ 9992\\ 5\\ 711\\ 4\\ 9992\\ 5\\ 713\\ 8\\ 524\\ 4\\ 758\\ 9\\ 1\\ 7\\ 5841\\ 6\\ 572\\ 5\\ 8\\ 572\\ 1\\ 4\\ 8\\ 5\\ 7\\ 1\\ 1\\ 6\\ 9\\ 9\\ 2\\ 5\\ 6\\ 5\\ 7\\ 1\\ 1\\ 4\\ 8\\ 5\\ 9\\ 1\\ 7\\ 1\\ 1\\ 1\\ 1\\ 1\\ 1\\ 1\\ 1\\ 1\\ 1\\ 1\\ 1\\ 1\\$	$ \begin{array}{c} 23\\22\\152\\23\\34\\68\\2\\11\\0\\8\\96\\66\\104\\81\\0\\20\\229\\105\\34\\-\\5\\8\\43\\107\\40\\50\\34\\-\\50\\34\\29\\46\\111\\93\\27\\42\\323\\101\\32\\162\end{array} $	$\begin{array}{c} 30\\ 11\\ 168\\ 20\\ 31\\ 81\\ 2\\ 6\\ 0\\ 8\\ 114\\ 83\\ 138\\ 87\\ 0\\ 0\\ 25\\ 308\\ 156\\ 35\\ -4\\ 76\\ 35\\ -4\\ 76\\ 544\\ 111\\ 34\\ 40\\ 44\\ 12\\ 84\\ 0\\ 29\\ 23\\ 49\\ 126\\ 76\\ 18\\ 36\\ 500\\ 115\\ 47\\ 260\end{array}$	$\begin{array}{c} 0.047\\ 0.023\\ 0.212\\ 0.028\\ 0.060\\ 0.105\\ 0.004\\ 0.009\\ 0.000\\ 0.000\\ 0.012\\ 0.292\\ 0.144\\ 0.189\\ 0.114\\ 0.000\\ 0.023\\ 0.125\\ 0.005\\ 0.023\\ 0.125\\ 0.062\\ 0.068\\ 0.005\\ 0.023\\ 0.126\\ 0.000\\ 0.041\\ 0.039\\ 0.058\\ 0.194\\ 0.145\\ 0.027\\ 0.089\\ 0.660\\ 0.268\\ 0.104\\ 0.538\\ 0.106\end{array}$

Table 3. Self-fertility of S<sub>1</sub> plants in Group III

Plant number	Number of flowers tripped	Number of pods that developed	Number of seeds that set	Seeds per flower tripped
301 302 304 305 306 306 307 309 3112 312 312 312 312 312 312 312 312 31	397 663 548 306 742 394 341 84 416 379 755 530 693 519 387 509 369 667 765 3752 782 717 5198 6205 6155 493 727 451 3945 227 468 561 738 727 451 3945 227 468 561 499 0 f the S <sub>1</sub> plant	$78 \\ 62 \\ 157 \\ 185 \\ 109 \\ 27 \\ 516 \\ 646 \\ 189 \\ 626 \\ 189 \\ 626 \\ 189 \\ 626 \\ 189 \\ 626 \\ 189 \\ 626 \\ 199 \\ 194 \\ 196 \\ 199 \\ 196 \\ 199 \\ 340 \\ 768 \\ 139 \\ 325 \\ 566 \\ 143 \\ 143 \\ 175 \\ 129 \\ 142 \\ 210 \\ 227 \\ 227 \\ 23$	79 78 102 315 120 559 749 39 749 39 52 75 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8	0.199 0.118 0.186 1.029 0.163 0.051 0.161 0.226 0.178 0.129 0.309 0.092 0.094 0.023 0.705 0.933 0.279 0.290 0.290 0.205 0.681 0.017 0.129 0.281 0.153 0.059 0.059 0.059 0.059 0.059 0.059 0.059 0.059 0.281 0.160 0.123 0.059 0.281 0.160 0.123 0.059 0.290 0.281 0.160 0.123 0.059 0.290 0.281 0.160 0.123 0.059 0.290 0.290 0.285 0.079 0.192 0.358 0.079 0.141 0.585 0.910 0.246

Table 4. Self-fertility of S1 plants in Group IV

		Repli	cation		Average num-
Sib-crosses	I	II	III	IV	ber of seed per flower
1 x $42$ 1 x 2 2 x 1 2 x 3 3 x 2 3 x 4 4 x 3 4 x 5 5 x 6 6 x 5 6 x 7 7 x 6 7 x 6 7 x 8 8 x 9 9 x 10 10 x 9 10 x 11 11 x 12 12 x 13 13 x 14 14 x 13 14 x 15 16 x 15 16 x 17 17 x 18 18 x 19 19 x 20 20 x 21 21 x 20 21 x	13162148817343255739065539029712093191248 217343255739065539029712093191248	32816289021476456457664112779272638845231119	02223984526064667726621490458281926920689	1 36248 7926827856810784516437130009653237010	0.125 0.250 0.675 0.275 0.375 1.000 2.025 1.000 2.225 1.300 1.975 0.675 0.600 0.400 0.4755 0.9255 0.9255 1.2255 0.975 1.2250 1.3255 1.34750 0.67525 1.5255 1.12500 0.9000 1.5255 1.175 1.2000 1.07500 0.9000

Table 5. Seed set data (per 10 flowers) for cyclic sib-crosses among  $S_1$  plants of Group I

		Repl	ication		Average num-
Sib-crosses	I	II	III	IV	ber of seed per flower
21 x 22 22 x 21 22 x 22 23 x 24 23 x 24 24 2554657687980 290 3011 312 x x x x x x x x x x x x x x x x x x x	58759686263818650892294137990744993755834	56668333541741467410350464483602222733065	11 37 50 302142957548766802569047207425274830 10 1125274830	14100466814359565999435311210479738801084170	0.875 0.675 1.000 0.750 1.0750 1.250 0.3255 0.3255 0.675 0.675 1.275 1.2755 1

Table 5 (Continued).

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		Rep	eplication Average number of sec				
Sib-crosses	I	II	III	IV	ber of seed per flower		
42 x 41 42 x 1	10 4	76	10 6	9 1	0.900 0.425		
592 A-3 x 652-25	30	19	16	21	2.150		
652-25 x 592 A-3	22	33	52	37	3.600		
Replication average	9.988	9.605	11.174	9.802			
Average of the sib-crosses				0.970			
L.S.D. (5%)	=				0.620		
L.S.D. (1%)	=				0.815		

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Table 5 (Continued).

Sib-crosses	Repli I	cation II	Average num- ber of seed per flower
101 x 142 101 x 102 102 x 101 102 x 103 103 x 102 103 x 104 104 x 103 104 x 105 105 x 104 105 x 106 106 x 107 107 x 108 108 x 107 108 x 107 108 x 109 109 x 108 109 x 100 110 x 101 111 x 112 112 x 111 112 x 113 113 x 112 113 x 114 114 x 113 114 x 115 115 x 114 115 x 116 116 x 117 117 x 116 117 x 118 118 x 117 118 x 119 119 x 120 120 x 121 121 x 122 122 x 121 122 x 123	$     \begin{bmatrix}             - & 3 \\             - & 3 \\             0 \\             - & 3 \\             0 \\             - & 3 \\             0 \\             0 \\         $		$\begin{array}{c} 000\\ 000\\ 000\\ 250\\ 000\\ 250\\ 000\\ 000\\$

Table 6. Seed set data (per 10 flowers) for cyclic sib-crosses among  $S_1$  plants of Group II

Sib emocran	Repli	Replication		
Sib-crosses	I	II	ber of seed per flower	
123 x 122	0	0	.000	
123 x 124	0	0	•000	
$124 \times 123$	-	0	.000	
$124 \times 125$	0	4 1 3 2 3 4 1	.200	
25 x 124	12	1	.100	
.25 x 126	2	3	.250	
.26 x 125	0	2	.100	
26 x 127	<u>4</u> .	5	• 350	
.27 x 126	-	4	.400	
.27 x 128	-	Ţ	.100	
28 x 127	160 <sup>-</sup>	0	.000	
28 x 129	· •	0	.000	
29 x 128	-	0	.000	
29 x 130	-	0	.000	
30 x 129	-	0	.000	
30 x 131	0	0	.000	
31 x 130	-	0	.000	
31 x 132	0	0	.000	
32 x 131	1	0	.050	
32 x 133	0	0 2 0 3 1 4 4	.100	
$33 \times 132$	U L	0	.000	
$33 \times 134$	4	3	.350	
34 x 133 34 x 135	1	$\frac{1}{\Sigma}$	.100	
	2	4	.300	
35 x 134 35 x 136	041221	. 4	.250	
	1		.100	
36 x 135 36 x 137	0	-	.000	
$37 \times 136$	0	0 0	.000	
$37 \times 138$			.000	
$38 \times 137$	2	1 6	•050	
28 = 120	0 2 1	1 6 5 2	.400	
38 x 139 39 x 138		2	.300	
39 x 140	С Б	1	.100	
40 x 139	1	11	.300	
$40 \times 139$ $40 \times 141$	0 5 1	0	.600 .000	
$41 \times 140$	ō	0	.000	
$41 \times 142$	-	0	.000	
$\frac{11}{12} \times 142$	-	n n	.000	
42 x 101	٦	),	.000	
C1 x 618 C1	รา้	70	6.050	
18 C1 x 5 C1	1 51 23	0 0 4 70 29	2.600	
		-7		
verage of sib-ci	COSSes		0.140	

Table 6 (Continued).

میں میں اور	Repl	ication	Average num-
Sib-crosses	I	II	ber of seed per flower
201 x 242 201 x 202 202 x 201 202 x 203 203 x 202 203 x 204 204 x 203 204 x 205 205 x 204 205 x 206 206 x 205 206 x 207 207 x 208 208 x 207 208 x 209 209 x 208 209 x 208 209 x 208 209 x 210 210 x 209 210 x 211 211 x 212 212 x 211 212 x 213 213 x 212 213 x 214 214 x 213 214 x 213 215 x 214 215 x 216 216 x 217 217 x 216 216 x 217 217 x 218 218 x 217 218 x 219 219 x 218 219 x 220 220 x 221 221 x 221 221 x 221 220 x 221 221 x 221 221 x 221 220 x 221 221 x 222	00022020060-42002200202020202020202020202020202020	2 0 0 0 3 6 7 1 8 1 - 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0	$ \begin{array}{c} 100\\ 000\\ 000\\ 100\\ 100\\ 100\\ 150\\ 050\\ -\\ -\\ -\\ -\\ -\\ -\\ -\\ -\\ -\\ -\\ -\\ -\\ -\\$
	-	-	

Table 7. Seed set data (per 10 flowers) for cyclic sibcrosses among  $S_1$  plants of Group III

	Replic	ation	Average num-
Sib-crosses	I	II	ber of seed per flower
222 x 221 222 x 223	1 8	-	.100
223 x 222	-	-	.800
223 x 224	0	5	.250
224 x 223	0	0	.000
224 x 225	0	0 2 2 2 1 0	.000
225 x 224 225 x 226	0 0	~ 2	.100 .100
226 x 225	õ	2	.100
226 x 227	0	1	.050
227 x 226	0	0	.000
227 x 228	0	1	.050
228 x 227 228 x 229	0	1 1 4 0 4 0	.050 .200
229 x 228	0	4	.000
229 x 230	ů Ö	Ŭ,	.200
230 x 229	0	Ó	.000
230 x 331	0	0	.000
231 x 230	0 4 3 2	1 0 2 4 0	.250
231 x 232 232 x 231	2	2	.000 .250
232 x 233	2	<u>ī</u>	.300
233 x 232	-		.000
233 x 234	õ	0	•000
$234 \times 233$	5	0	.250
234 x 235 235 x 234	2	ے ا	•550 •150
235 x 234 235 x 236	2	6	.400
236 x 235	0	2	.100
236 x 237	4	8	.600
237 x 236	059220430	0 2 1 6 2 8 1 15	.200
237 x 238 238 x 237			•750
238 x 239	1 2	1	.200 .150
239 x 238		Ē	.500
239 x 238 239 x 240	4	3 1 5 6 13	•500
240 x 239	- 4 12 2 7	13	•950
$240 \times 241$	12	-	1.200
241 x 240 241 x 242	7	-	.200 .700
$242 \times 241$	0		.000
242 x 201	0	0	.000
Average of the	e sib-crosses		.262

## Table 7 (Continued).

Sib-crosses		Repli	cation		Average num- ber of seed
510-cr08588	I	II	III	IV	per flower
301 x 342 301 x 302 302 x 301 302 x 303 303 x 304 304 x 305 305 x 304 305 x 306 306 x 305 306 x 307 307 x 308 308 x 307 308 x 309 309 x 310 310 x 311 311 x 312 312 x 311 312 x 311 312 x 312 313 x 314 314 x 315 315 x 316 316 x 317 317 x 318 318 x 319 319 x 318	566258551151 021223173421031903622050	9870791640156062284331002616755245037	319858274625107080509129110826664387243	1274601391372569401166941488678945859	1.425 $1.625$ $1.450$ $1.025$ $1.650$ $2.475$ $2.600$ $1.700$ $0.750$ $0.100$ $0.350$ $0.600$ $0.225$ $0.300$ $0.600$ $0.450$ $0.450$ $0.450$ $0.450$ $0.700$ $1.600$ $1.425$ $2.300$ $1.550$ $0.175$ $0.125$ $0.500$ $0.750$ $1.425$ $1.200$ $1.950$ $3.000$ $0.750$ $0.725$ $0.725$

Table 8. Seed set data (per 10 flowers) for cyclic sib-crosses among S1 plants of Group IV

<sup>a</sup>Computed

86

80<sup>4141</sup>

0.11		Replic	cation		Average num-
Sib-crosses	I	II	III	IV	ber of seed per flower
$319 \times 320$ $320 \times 319$ $320 \times 321$ $321 \times 322$ $321 \times 322$ $322 \times 321$ $322 \times 322$ $323 \times 322$ $323 \times 322$ $323 \times 3224$ $324 \times 3225$ $326 \times 3227$ $326 \times 3227$ $326 \times 3227$ $328 \times 3229$ $320 \times 3229$ $320 \times 3229$ $320 \times 3227$ $328 \times 3229$ $320 \times 3229$ $320 \times 3229$ $320 \times 3229$ $320 \times 3229$ $320 \times 3229$ $320 \times 3229$ $330 \times 3311$ $331 \times 332$ $3331 \times 3332$ $3332 \times 3334$ $334 \times 3335$ $335 \times 3336$ $337 \times 338$ $338 \times 339$ $339 \times 340$	22514645201237241754202011056346585569103 11056346585569103	167427431001401992230513137316395278072436	85168658571446520305020128628497000013129	70132644286079818600421106412298450053112	0.450 0.825 1.350 0.350 1.375 1.125 0.4500 0.5500 0.6250 0.4500 0.4500 1.4000 1.4000 1.4000 1.4000 1.4000 1.4200 1.4200 1.5500 0.5550 1.5500 1.7500 1.5500 1.7500 1.5500 1.7500 1.5500 1.7500 1.5500 1.7500 1.5500 1.7500

Table 8 (Continued).

0.11		Replication						
Sib-crosses	I	II	III	IV	ber of seed per flower			
340 x 339 340 x 341 341 x 340 341 x 342 342 x 341 342 x 301 652-25 C1	14 1 2 14 14 14	452255	9 5 7 12 21	1 8 11 4 17 21	0.450 0.475 0.550 0.300 0.950 1.275			
x 5 Cl 5 Cl x	6	8	54	60	3.200			
652 <b>-2</b> 5 Cl	42	38	67	68	5.375			
Replication average	6.210	6.560	16.71	15.95				
Average of t	1.061							
L.S.D. (5%)	0.939							
L.S.D. (1%)	=				1.234			

Table 8 (Continued).

S <sub>l</sub> female parent	I	Replic: II	ation III	IV	Average num- ber of seed per flower
ـــــــــــــــــــــــــــــــــــــ	E	ackcrossed	to 585	B-2 C1	
1234567890 111234567890 111234567890 1222222222222222222222222222222222222	212 90480 76560 380 182 0 341 516 7731 16 332 332 435 331 516 7731 16	17 <sup>a</sup> 42076255291786787843937433039 12132218767843931217433039	24415251556622180798088033821639	17 3255 312 432 13 432 14 33 14 32 14 33 14 33 14 33 14 33 14 33 14 33 14 33 14 33 14 33 14 33 14 33 14 33 14 33 14 33 14 33 14 33 14 3 3 14 33 14 14 14 14 14 14 14 14 14 14 14 14 14	1,975 4,200 1,875 3,975 3,655 2,55 3,025 5,025 3,025 5

Table 9. Seed set data (per 10 flowers) of S<sub>1</sub> plants in Group I after backcrossing to the S<sub>0</sub> parental clone

<sup>a</sup>Computed

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C. formal a		Average num-			
S <sub>1</sub> female parent	I	II	III	IV	ber of seed per flower
33 34 35 37 38 39 40 41 42	17 31 30 39 39 29 19 27 25	16 30 27 28 32 21 24 21 25	18 20 38 20 41 25 35 25 30	17 26 27 38 38 28 31 34 17	1.700 2.675 3.050 3.125 3.750 2.575 2.725 2.675 2.425
Replication average	3.052	2.760	3.060	3.178	
General average					3.012
L.S.D. (5%) =					0.692
L.S.D. (1%) =					0.916

Table 9 (Continued).

S <sub>l</sub> female parent	Average number	of seed per flower
$ \begin{array}{c} 101\\ 102\\ 103\\ 105\\ 106\\ 107\\ 108\\ 109\\ 110\\ 111\\ 112\\ 113\\ 115\\ 116\\ 117\\ 118\\ 119\\ 120\\ 121\\ 122\\ 123\\ 124\\ 125\\ 126\\ 127\\ 128\\ 129\\ 130\\ 131\\ 132\\ 133\\ 134\\ 135\\ 136\\ 137\\ 139\\ 140\\ 142 \end{array} $	Backcrossed to	618 C1 0.7 0.0 1.6 0.3 1.0 0.2 1.1 0.8 1.5 1.2 0.2 1.0 1.2 0.3 2.1 0.0 2.1 0.0 1.7 1.4 0.0 1.7 1.4 0.0 1.7 1.4 0.0 0.9 0.9 0.0 0.8 0.0 0.8 0.0 0.8 0.0 0.8 0.0 0.8 0.0 0.8 0.0 0.8 0.0 0.8 0.0 0.8 0.0 0.8 0.0 0.8 0.0 0.9 0.0 0.8 0.0 0.0
Average		0.813

Table 10. Seed set data of  $S_1$  plants in Group II after backcrossing to the  $S_0$  parental clone

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S <sub>l</sub> female parent	I	Replic II	cation III	IV	Average num- ber of seed per flower
	]	Backcrosse	ed to 631-	101 C1	
201 202 203 204 205 206 206 207 206 207 208 210 211 212 213 216 217 219 222 2256 228 229 231 232 232 233 234	9133533715347758883205450580543121 1231728883205450580543121	17 92 29 43 63 90 59 31 840 57 40 18 80 51 02 867 27	1451539702223578943289212130573423332 333289212130573423332 12122573423332	331150 10550358236606124002699083 12312232221223221233 12512322233 125123223 125123223 125123223 125123223 125123223 1251233 125123 125123 125125 125123 12512 125123 12512 125123 12512 125	1.825 $1.150$ $1.925$ $2.350$ $3.000$ $1.275$ $.750$ $1.475$ $1.275$ $1.375$ $3.375$ $2.900$ $2.125$ $2.900$ $2.925$ $2.900$ $2.925$ $2.900$ $2.925$ $2.750$ $3.575$ $1.850$ $1.725$ $1.250$ $1.400$ $2.700$ $2.300$ $1.650$ $1.575$ $0.700$ $3.200$ $3.300$ $1.150$ $2.400$

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Table 11. Seed set data (per 10 flowers) of S<sub>1</sub> plants in Group III after backcrossing to the S<sub>0</sub> parental clone

<sup>a</sup>Computed

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6 . Aswa]a	and and a set of the set	Average num-			
S <sub>1</sub> female parent	I	II	III	IV	ber of seed per flower
235 236 237 238 239 240 241 242	28 30 16 21 20 34 26 23	24 19 19 29 12 45 22 33	28 24 16 7 13 33 15 31	28 33 26 16 22 33 18 29	2.700 2.650 1.925 1.825 1.675 3.625 2.025 2.900
Replication average	2.035	2.192	2.072	2 2.222	
General average					2.131
L.S.D. (5%) =					0.831
L.S.D. (1%) =					1.100

Table 11 (Continued).

	-				
S <sub>l</sub> female parent	I	Replic II	cation III	IV	Average num- ber of seed per flower
		Backcrosse	ed to 5 C1		
301 302 303 305 306 307 306 307 309 311 312 314 316 317 3190 321 3224 3224 3226 3290 3312 3234 3267 3290 3312 3334 3356 337 338 38	412311121411252132242113332122413372	32079079546817497353581439994559063370 213 25411324521 3131111412342	3242221234314633414621594576158777744	424322232459139723146639447686014004 <b>1</b>	3.625 1.875 3.800 1.575 2.850 1.5725 2.42500 1.5525500 1.55255000 1.5555000 1.5555000 1.5555000 1.5555000 1.5525555000 1.552555000 1.52255000 1.52255000 1.525555000 1.525555000 1.525555000 1.525555000 1.525555000 1.525555000 1.5255555000 1.52555555000 1.525555000 1.5255555000 1.52555555000 1.5255555000 1.5255555000 1.525555550000 1.52555550000 1.52555555550000 1.5255555555550000 1.5255555500000 1.52555555555500000 1.52555555555555500000 1.52555555555555555555555555500000000000

Table 12. Seed set data (per 10 flowers) of  $S_1$  plants in Group IV after backcrossing to the  $S_0$  parental clone

		Average num-			
S <sub>l</sub> female parent	I	II	III	IV	ber of seed per flower
339 340 341 342	19 9 26 17	<b>21</b> 15 22 19	33 25 25 19	27 28 34 17	2.500 1.925 2.675 1.800
Replication average	2.668	2.435	3.008	3.062	
General average					2.793
L.S.D. (5%) =					0.908
L.S.D. (1%) =					1.202

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Table 12 (Continued).