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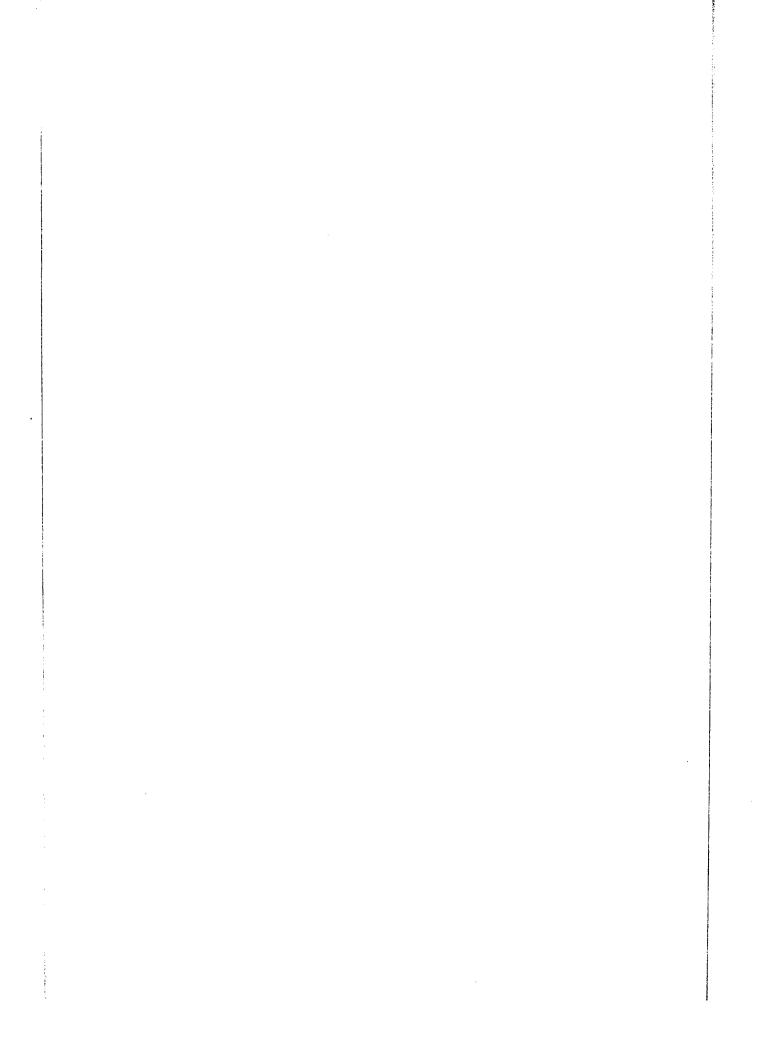
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PHYSIOLOGIC SPECIALIZATION IN PUCCINIA CORONATA

AVENAE (COHDA) ERIKS. AND HENN.

By

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A Thesis Submitted to the Graduate Faculty

for the Degree of

DOCTOR OF PHILOSOPHY

Major Subjects - Plant Pathology and Crop Breeding

Approved:

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TABLE OF CONTENTS

-2-

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	Page
INTRODUCTION	3
PERTINENT LITERATURE	4
MATERIALS AND METHODS	13
Selection of possible differential hosts	15
Types of crown rust infection	17
Field test of varieties and selections	18
IDENTIFICATION OF PHYSIOLOGIC FORMS	21
Cultures collected during 1927 and 1928	22
Cultures collected during 1929 and 1930	25
Key for identification of physiologic forms	29
Nature of physiologic forms identified	30
PREVALENCE AND GEOGRAPHIC DISTRIBUTION OF PHYSIOLOGIC FORMS IDENTIFIED) 34
Physiologic forms collected on Rhammus	51
Physiologic forms collected on Avena	53
RESPONSE OF VARIETIES, SELECTIONS, AND SPECIES	56
GRAMINEOUS HOST RANGE OF CERTAIN PHYSIOLOGIC FORMS .	72
DISCUSSION AND CONCLUSIONS	78
SUMMARY	81~
ACKNOWLEDGMENTS	83
LITERATURE CITED	84
EXPLANATION OF PLATES	91

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PHYSIOLOGIC SPECIALIZATION IN PUCCINIA CORONATA

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AVENAE (CORDA) ERIKS. AND HENN.

INTRODUCTION

The presence of physiologic specialization in the rusts was first demonstrated in 1894 by Eriksson and Henning (29), and since that time many rusts have been shown to consist of two or more entities, each having a specific host relationship. Two developments have come about in our understanding of these specialized entities, first that major physiologic units exist within such species of rust as <u>Puccinia graminis</u> Pers. and <u>P</u>. <u>coronata</u> Corda and are distinguished by their ability to infect certain host species; second, that there are minor units which are separated by their specific reaction on varieties. The former are considered as physiologic varieties and the latter as physiologic forms.

In the present investigation an attempt has been made to identify, and determine the prevalence and distribution of physiologic forms of <u>P. coronata avenae</u> (Corda) Eriks. and Henn. present in the oat-growing regions of the United States and Ganada during the years 1927, 1928, 1929, and 1930. The relationship of the occurrence of these forms to the specific host from which they were collected, and the host ranges of the most important forms has also been studied.

PERTINENT LITERATURE

Our knowledge of physiologic specialization in the rust fungi begins with Schroeter (59). In 1879 he calls attention to the presence of physiologic variation in certain rusts on Carex. Similar variation in Puccinia graminis Pers. and other rust fungi were reported by Dietel (18), in 1887. It remained for Eriksson and Henning (29) in 1894 to first definitely demonstrate physiologic specialization based on pathogenicity. They showed that there were present within Puccinia coronata Corda at least four physiologic varieties and within P. dispersa Eriks. two such varieties, which were distinguished by their ability to infect certain hosts. Later in the same year Eriksson (24) described six physiologic varieties in P. coronata; four in P. dispersa; five in P. graminis, and five in P. glumarum (Schm.) Eriks. and Henn. Three physiologic varieties were added to P. coronata by Klebahn (41, 42, 43) and three by Eriksson (25, 28), while Muhlethaler (49), using a different host relationship as the basis of his classification, described twelve physiologic varieties. Eriksson (26) later added two physiologic varieties to P. dispersa and reised all of these to specific rank. One physiologic variety was added to P. graminis by Eriksson (27) and three by Jaczewski (36), while in the United States Stakman and Piemeisel (65) have added the physiologic variety P. graminis tritici conapacti and suggest the establish-

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ment of <u>P. phlei-pratensis</u> Eriks. and Henn. as an additional variety under <u>P. graminis</u>. This same phenomenon has been observed and studied in many additional species of rust by Magnus (44), Rostrup (57), Klebahn (42), Dietel (19), Ward (68), Bandi (9), Probst (56), Jordi (37, 38), and others.

That it is possible to further divide certain of these physiologic varieties described by Eriksson and Henning (29), Eriksson (24), Klebahn (42), and others, into physiologic forms distinguished by their specific reaction on varieties or species, was first shown by Stakman and Piemeisel (65). Working with the physiologic variety <u>Puccinia graminis tritici</u> Eriks. and Henn., Stakman and Levine (62) were able to distinguish 37 physiologic forms on the basis of the differential reaction of 12 varieties of wheat. The existence of four physiologic forms within <u>P. graminis avenae</u> Eriks. and Henn. was demonstrated by Stakman, Levine and Bailey (63). Later Bailey (8) described a fifth form, and in 1928 Gordon (31) and Gordon and Bailey (32) reported a sixth physiologic form occurring in Canada.

The presence of physiologic forms within the rust species is not limited to <u>P. graminis</u>. Mains and Jackson (46) were able to distinguish 12 physiologic forms of <u>P. triticina</u> Eriks. by their specific reaction on 11 varieties of wheat. Scheibe (58) working in Germany discovered four forms of <u>P. triticina</u>, one of these being similar to one described by Mains and Jackson.

-5-

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Mains (45) reports the presence of two physiologic forms in <u>P</u>. <u>dispersa</u>, two in <u>P</u>. anomala Rostr., and four in <u>P</u>. sorghi Schw. Stakman and Christensen (60) earlier reported the presence of three and possibly five physiologic forms of <u>P</u>. sorghi. Later Stakman, Christensen and Brewbaker (61) recognized seven physiologic forms of <u>P</u>. sorghi by their specific reaction on eight selfed lines of corn. Hungerford and Owens (35) reported indication of the presence of two or more specialized forms of <u>P</u>. glumarum tritici Eriks. and Henn. Bailey (7) demonstrated the presence of at least three, and possibly four, forms of <u>P</u>. <u>helianthi</u> Schw.

It is clear that with the increase in our knowledge of physiological specialization, at least in <u>P. graminis</u>, we have gone from a rather wide grouping as proposed by Eriksson and Henning (29), Eriksson (24), and others to a more narrow grouping as set out by Stakman and Piemeisel (65). The work that has been done on <u>P. coronata</u> seems to tend in the same course. The development of our knowledge of specialization in this species begins with de Bary (11), when he discovered in 1865 that the aecial stage of <u>P. coronata</u> occurred on <u>Rhamnus cathartica</u> L. and <u>R. frangula</u> L. Nielsen (52) in 1877 secured infection on <u>Lolium perenne</u> L. with aeciospores from <u>Rhamnus cathartica</u> and in turn infected oats with the urediniospores from <u>Lolium</u> <u>perenne</u>. Then Cornu (13) in 1880 infected oats directly with

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acciospores from Rhamnus cathartica. Plowright (54) in 1889 suggested the presence of two species within Puccinia coronata. His conclusions were based mainly upon the fact that teliospores from Lolium perenne would not infect Rhamnus frangula, while teliospores from Dactylis glomerata L. and Festuca sylvatica Vill. readily produced aecia on Rhamnus frangula. Klebahn (39) reported in 1892 that there were two species of crown rust. That species which bore teliospores on Dactylis glomerata, Festuca sylvatica and other grasses, and bore aeciospores on Rhammus frangula, he called Puccinia coronata. While the species which produced teliospores on Lolium perenne, Avena sativa L., Festuca elatior L., Arrhenatherum elatius (L.) Beauv. and other grasses, and produced acciospores on Rhamnus cathartica and other Rhammus species, he called Puccinia coronifera Klebahn. The conclusions of Klebahn were supported in later investigations by Klebahn (40, 41, 42), Eriksson (25, 28), Eriksson and Henning (30), and Muhlethaler (49).

Eriksson and Henning (29) were probably the first to recognize the presence of physiologic varieties in crown rust. They divided <u>Puccinia coronata</u> into three series and certain of these were subdivided into "Formen". Series I, with aecial stage on <u>Rhamnus cathartica (Puccinia coronifera Kleb.</u>), was divided into two "Formen": <u>Avenae</u> and <u>Alopecuri</u>; series II, with aecial stage on <u>R. frangula (Puccinia coronata Kleb.</u>), consisted of

-7-

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one "Formen" occurring on Dactylis glomerata and Festuca silvatica; and series III, an aecial stage on Rhamnus dahurica (Puccinia coronata var. himalensis Barcl.), consisted of one "Formen". Two physiologic variaties Calamagrostidis and Melica were not included in the series because their accial stage was unknown. Later Eriksson (24) divided crown rust into four series and certain of these series were subdivided into "forme species." Series I (Puccinia coronifera Kleb.) was divided into four "forme species": Avenae, Alopecuri, Festucae, and Lolii; series II (P. coronata (Corda) Kleb.) contained only one "forme species": Calamagrostis; series III (P. coronata var. himalensis Barcl.) contained no "forme species"; and series IV (aecial stage unknown) contained one "forme species": Melicae. Additional "forme species" were added to series I and II by Klebahn (42) and Eriksson (25). In 1911 Muhlethaler (49) presented a somewhat different arrangement. He divided crown rust into five series: I. P. coronifera Kleb.; II. P. himalensis (Barcl.) Diet.; III. P. Alpinae-coronata nov. sp.; IV. P. coronata (Corda) Kleb.; V. P. coronata Corda s. lat. Series I contained nine "forme species", series IV three "forme species", series IV three "forme species", and series V one "forme species."

Treboux (65), working with crown rust in southern Russia, did not secure the differential reaction on Rhamnus species

-8-

reported by Klebahn (42), Eriksson (24), and Muhlethaler (49). Using aeciospores secured from <u>R</u>. frangula he was able to infect <u>Avena sativa</u>, a host belonging to <u>P</u>. coronifera Kleb., and using aeciospores secured from <u>Rhamnus cathartica</u> he was able to infect <u>Agrostis stolonifera</u>, <u>Calamagrostis arundinacea</u>, and <u>Phalaris arundinacea</u>, all of which are hosts of <u>Puccinia</u> <u>coronata</u> (Corda) Kleb. Troboux (66) concluded that the existence of separate species of crown rust upon either <u>Rhamnus</u> <u>cathartica</u> or <u>R</u>. frangula is doubtful.

In the United States, Arthur and Fromme (5), Arthur and Holway (6), Carleton (12), Melhus, Dietz and Willey (47), Melhus and Durrell (48), Lietz (21), and others prefer to retain the name <u>Puccinia coronata</u> Corda. The marked differentiation on Rhamnus species reported by Klebahn (41) has not been reported in America. Arthur and Holway (6) were able to produce infection on <u>Avena sativa</u> using acciospores secured from <u>Rhamnus</u> <u>lanceolata</u>. Carleton (12) used acciospores produced on <u>R. lanceolata</u> and produced infection on <u>Phalaris caroliniana</u>. The host range of <u>Puccinia coronata</u> has been extended, both on the alternate hosts and on the gramineous hosts, by Arthur (2, 3, 4), Melhus and Durrell (48), Melhus, Dietz and Willey (47), and Dietz (21). Melhus, Dietz and Willey (47) using teliospores produced on <u>Avena sativa</u> were able to secure pycnia on <u>Rhamnus frangula</u>.

-9-

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Dietz (21) secured normal aecia on <u>R. dahurica</u> using teliospores produced on <u>Avena sativa</u>. The results secured by investigators in the United States do not justify the division of crown rust into the various species reported in Europe by Rlebahn (39), Eriksson (24), and Muhlethaler (49).

Melhus, Dietz and Willey (47) studied four "biologic forms" (physiologic varieties) of crown rust: <u>P. coronata avenae</u>, <u>P. coronata lolii</u>, <u>P. coronata calamagrostis</u>, and <u>P. coronata holci</u>. They determined the reaction of various gramineous species to each of these physiologic varieties. The gramineous host ranges of these four physiologic varieties overlap somewhat, but the reaction of each host was specific for each physiologic variety.

Dietz (21) found that the alternate host range of crown rust was not limited to the genus Rhamnus, or to the family Rhamnaceae. He secured aecial infection of crown rust on <u>Berchemia scandens</u> (Hill) Trel. of the family Rhamnaceae, and <u>Lepargyrea canadensis</u> of the family Eleagnaceae, in addition to thirteen species of Rhamnus. Dietz (21) secured a marked specialization in reaction on the alternate hosts with different varieties of crown rust. This specialization of varieties and the wide alternate host range of crown rust is further evidence, Dietz believes, that the crown rust organism should be considered as one species.

Hoerner (34) was the first to report the presence of physic-

-10-

logic specialization within the variety <u>P. coronata avenae</u>. He distinguished four physiologic forms on the basis of their reaction on Iowa 73 (Ruakura) and Iowa 96 (Green Russian):

Form 1. infects both variaties normally. Form 2. infects both varieties weakly. Form 3. infects Iowa 73 weakly and Iowa 96 normally. Form 4. infects Iowa 73 normally and Iowa 96 weakly.

Popp (55) identified 22 cultures of <u>P. coronata avenae</u> collected from five provinces of Canada. Using Iowa 96 (Green Russian), Sterilis Selection, and Minnesota 539 (White Russian) as differential hosts, he was able to distinguish four physiologic forms as follows:

Form	1.	(Eight cultures) infects Sterilis Selection	n weakly;
		Minnesota 539 (White Russian) normally, and	l Iowa 96
		(Green Russian) normally.	

- Form 2. (Eight cultures) infects all three varieties weakly.
- Form 3. (Two cultures) infects Sterilis Selection normally; Minnesota 539 (White Russian) weakly, and Iowa 96 (Green Russian) weakly.
- Form 4. (Four cultures) infects all varieties more or less normally.

Parson (53) obtained 15 collections of crown rust from the United States and Canada. Using four varieties as differential hosts, he was able to distinguish five physiologic forms as follows:

Ruakura - resistant (Hoerner's Form 3)	
Avena sterilis nigra - resistant	form 3
A. sterilis nigra - susceptible	
Red Rustproof - resistant	form 5
Red Rustproof - susceptible	form 4
Ruakura - susceptible (Hoerner's Form 1)	
Green Mountain - resistant	form 2
Green Mountain - susceptible	form 1

Parson determined the reaction of each of his forms on 27 varieties, selections, and species of oats, and found that none were resistant to more than three physiologic forms.

Murphy (50) secured 45 collections of <u>Puccinia coronata</u> <u>avenae</u>, of which 32 were collected on Avena and 13 on Rhamnus. An equal number of cultures isolated from these collections were each tested on pure line selections of 33 varieties and species of oats. Eight of these acting as differential hosts disclosed the following nine physiologic forms:

```
Belar - resistant
 Red Rustproof (C.I. 1079<sup>1</sup>) - resistant
    College Algerian (C.I. 2052) - resistant . . . form 3
   College Algerian (C.I. 2052) - susceptible . . form 8
 Red Rustproof (C.I. 1079) - susceptible
    Cowra - resistant. . . . . . . .
                                               . . form 9
    ••••form 7
Belar - susceptible
  Iowa No. 69 - resistant
    Avena strigosa - resistant
     Anthony (C.I. 2143) - resistant .
Anthony (C.I. 2143) - susceptible
                                       •••••form 4
                                         •••• form 2
    Avena strigosa - susceptible
 Iowa No. 69 - susceptible
    Iowa No. 102 - resistant . .
                                                  form 6
    Iowa No. 102 - susceptible . . . .
                                                 . form 5
```

The 13 aecial collections on five species of Rhamnus were identified as follows: four collections on <u>Rhamnus cathartica</u> were form 3; six collections on <u>R. lanceolata</u> were form 5; one each from <u>R. tinctoria</u>, <u>R. spp</u>. (from Montana), and <u>R. infectoria</u> were forms 7, 8, and 9, respectively.

1 C.I. = Bureau of Plant Industry, Cereal investigation accession number.

MATERIAL AND METHODS

Leaves infected with <u>Puccinia coronata avenae</u> were collected on naturally infected oat plants and on Rhamnus naturally and artificially infected, during the years 1927, 1928, 1929, and 1930. For each collection, the name of collector, date, place, and the name of the host were recorded. During transit, crown rust spores retain their viability much longer when the infected leaves are allowed to dry immediately after collection and are placed in an ordinary manile envelop. Aecial collections, due to their short viability, were immediately used to infect oat plants, and the resulting uredinial stage continued, or the infected leaves stored in the refrigerator. All cultures were maintained on the variety Iomine (C.I. 2327) during 1928. However, this variety showed partial resistance to certain forms and Markton (C.I. 2053) was substituted for it during 1929 and 1930.

Single uredinium cultures were isolated by inoculating Markton or Iomine plants sparsely with spores from a particular collection. About six or seven days after inoculation and before any uredinia had broken through the epidermis, a plant was selected which bore a single uredinium. This plant was then placed under a covered lamp chimney (Plate I, fig. 2) and the uredinicepores allowed to mature. Because of the possibility of the presence of more than one physiologic form in a collec-

-13-

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tion, two single-uredinial isolations were usually made from each collection. Each culture had as its source a single uredinium.

Seedling plants were usually inoculated between 4 and 6 p.m. The first leaf of each plant was gently drawn between the moistened forefinger and thumb, then the entire plant was moistened with a constant-pressure sprayer. The plants were then moved to the room containing the stock cultures and each plant was individually inoculated with the desired culture by applying a small amount of crown rust spores to the moistened surface of the first leaf with a flattened needle. The plants were again sprayed and placed in a moist inoculation chamber where they were kept for approximately 14 hours. In order that the plants might not dry too quickly or otherwise become injured, the chambers were opened at about 8 a.m. and the plant retained in the open chamber until about 4 p.m.

Where types of infection were to be determined, the plants were placed in muslin compartments (Plate I, fig. 1) similar to those described by Melhus, Dietz and Willey (47); while those used for stock cultures were held under lamp chimneys. The tops of the chimneys were covered with a thin layer of cotton held between two pieces of cheese-cloth and this all fastened with a rubber band (Plate I, fig. 2). Each lamp chimney would cover the contents of a three-inch flower pot. These pots were

-14-

variety used as a differential host by Parson (53).

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At the beginning of the investigation, approximately 200 variaties, selections, and species, reported as being resistant to crown rust, were inoculated with ten cultures which furnished a basis for the selection of possible differential hosts. From these investigations, a set of 33 differential hosts was selected. This preliminary study gave evidence that certain of the commercial variaties and so-called pure-line selections on hand were not homozygous for reaction to all crown rust cultures. Because of this heterozygous reaction, a single-panicle selection was made of each of the possible differential hosts at the beginning of the experiment. Unless otherwise stated, pure-line selections were used throughout these investigations for differentiating physiologic forms.

The differential hosts were inoculated with each culture of crown rust. When pure lines were used as possible differential hosts, five plants of each were inoculated and their reaction recorded. Ten or more plants were inoculated when the possible differential host was not a pure line. Whenever a variety of species appeared heterozygous for resistance to a particular culture, a larger number of plants were inoculated and the predominating reaction recorded.

-16-

Types of Crown Rust Infection

In recording the reaction of different varieties, selections, and species to crown rust, it was necessary to adopt a series of rust manifestations which would describe the classes of host reactions observed. The types of crown rust infection observed are shown in Plate II, fig. 2, and a description follows:

Immine

Completely resistant

Highly resistant

Moderately resistant

Moderately susceptible

Completely susceptible 4.

I. - No macroscopic evidence of infection.

- 0. No uredinia formed, necrotic areas present.
- 1. Urcdinia few, small, always in necrotic areas, also more or less necrotic areas produced without the development of uredinia.
- 2. Uredinia fairly abundant, small to medium size, always in necrotic or very chlorotic areas.
- 3. Uradinia abundant, madium in size, and surrounded by chlorotic areas.
- 4. Uredinia abundant, large, no necrosis or chlorosis immediately surrounding the uredinia.

The reaction of all plants to crown rust was recorded 10 and 14 days after inoculation. During the interval between the time of inoculation and recording, the plants were kept under optimum conditions for the normal development of the oat plants. The type of infection on a particular pure line inoculated with a particular physiologic form is quite constant when environmental conditions are uniform and favorable. During cloudy weather, artificial light is necessary in order to secure normal types of infection. A "green-island" type of infection may be developed on a completely susceptible plant by reducing the light intensity for a period of two days. Excessive drought, wilting, excessive high or low temperature, and abnormal nutrition also tend to produce subnormal types of infection.

In the present investigation, a difference of at least two types of infection was considered necessary to establish a new form, that is, a type 0 contrasted with a type 2, a type 1 with a type 3, or a type 2 with a type 4. It was unusual for the reaction of a pure line to a particular physiologic form to vary more than one type under normal greenhouse conditions.

Field Test of Varieties and Selections

A knowledge of the field reaction of varieties studied under greenhouse conditions is very desirable. This is especially true of those varieties used as differential hosts, and of those additional varieties used in determining the host range of a particular physiologic form. During 1926, 1927, and 1928, a uniform oat crown-rust nursery was grown at Ames, Iowa, and Experiment, Georgia. This uniform nursery contained pure-line selections of 100 varieties which were selected for their known reaction to crown rust. During 1929 a new uniform

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oat crown-rust nursery containing pure-line selections of 100 varieties, species, and selections was grown at eight different experiment stations in the contral and southern portions of the United States. The locations of these murse-ies and the names of the cooperators were as follows:

Location	:	Cooperator
Ames, Iowa Knoxville, Tennessee Experiment, Georgia Tifton, Georgia A. & M. College, Mississ Denton, Texas Stillwater, Oklahoma Manhattan, Kansas	ipp1	S.H. Essary R.P. Bledsoe W.J. Davis L.E. Miles P.B. Dunkle J.C. Ireland J.H. Parker and C.O. Johnston

During 1929, 660 additional varieties, selections, and species were grown at Manhattan, Kansas, and Ames, Iowa. Three hundred of these were introduced foreign varieties supplied by T.R. Stanton, Office of Cereal Crops and Diseases, Bureau of Plant Industry, Washington, D.C.

The percentages of crown rust infection on the different pure lines, varieties, selections, and species included in the oat crown-rust nurseries were determined according to the scale illustrated by Durrell and Parker (23). Types of crown rust are often very difficult to determine under field conditions, because of the maturity of the oat plant and the presence of

-19-

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uredinia in different stages of development. Whenever a variation was observed, the range from the most resistant to the most susceptible was recorded.

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IDENTIFICATION OF PHYSIOLOGIC FORMS

One hundred seventy-one collections of <u>Puccinia coronata</u> <u>avenae</u> were secured during 1927, 1928, 1929, and 1930, on varieties and species of Avena, and species of Rhammus. Two hundred forty-five rust cultures were isolated from these collections. The distribution of these collections according to the time and the state or province from which they were collected is given in table 1.

Table 1. Distribution of collections of Puccinia coronata avenae during 1927, 1928, 1929, and 1930, in states and provinces.

State or			collec		Total	
Province	: 1927	: 1928	: 1929	: 1930 :	collections	
Alabama			l		1	
California		5			1 5	
Florida		5 1 4		12	13	
Georgia		4	4			
Illinois	l				8 1	
Iowa	2	16	29	21	68	
Kansas	2		10		12	
Minnesota			2		2	
Mississippi		4	4		8	
Missouri	l	1 1 1 1 1	4		6	
Nebraska		1			l	
North Dakota	1	l	1		3	
Ohio		1			1	
Oklahoma	1	1	3		5	
Ontario		1			1	
Oregon		1	3		4	
Quebec	1				l	
South Dakota			1		l	
Tennessee		1	1 6		7	
Texas	1	6	4 3		11	
Virginia			3		3	
Nest Virginia		1	8		9	
Total	10*	45	83 1927 w	33 Jere furni	171 shed by Mr.	

"Nine collections secured in 1927 were furnished by Mr. L.D. Leach, who made a preliminary study of physiologic specialization and identified three physiologic forms (unpublished data). The number of collections secured on different hosts during the years 1927, 1928, 1929, and 1930 is given in table 2. Table 2. Host and number of collections of crown rust secured during 1927, 1928, 1929, and 1930.

		*	ومخارك وجبي يستوجد والتقار	Number co	llection	18
ماريد اول در او «او در .	Host	;	1927	: 1928	: 192	29 : 1930
Avena s	species		10	32	66	3 12
	e cathartica		-	4	1	5
14	chlorophora		•••		-	- 2
11	infectoria			1	13	5 5
17	lanceolata			6		- 2
11	montana			1		5 3
H	tinctoria		-	ī	-	- 4
Total			10	45	83	3 33

All the collections on Rhammus species were from artificially inoculated plants, except one on <u>Rhammus lanceolata</u> secured during 1928, and three on <u>Rhammus cathartica</u> (two during 1928 and one during 1929). The collections on Avena species were all taken from naturally infected plants in the field.

Cultures Collected During 1927 and 1928

Fifty-five cultures of crown rust were isolated from an equal number of collections taken during 1927 and 1928. A uniform set of 33 pure lines, selected from different variaties, selections, and species of oats was separately inoculated with each of these cultures. Nine physiologic forms were identified among these 55 cultures by the differential reaction of eight of the 33 pure lines. In table 3 is given the name of each of the 33 pure-line-selected: variaties, selections, and species, used as possible differential hosts, and the average type of

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Pure-line selections of: varieties, selections,	: :Averag	e type		infoct	ion with	phy	siologic	form	number:
and species	: 1 :	2:	3	: 4	: 5 :	6	<u>: 7 :</u>	8	: 9
			~		4		0	^	
Belar (C.I. 2760)	4	4	0	3	4	3	0	0	Ţ
Red Rustproof (C.I. 1079)	4	4	0	3	4	4	4	1	4 A
College Algerian (C.I. 2052)	4	4	0	4	4	4	4	4	4 <u>4</u> . ▼
Cowra (C.I. 2761)	4	3	0	3	4	4	3	0	1. A
Iowa 69 (C.I. 2463)	0	0	4	0	4	4	4	4	4
Avena strigosa (C.I. 1782)	4	2	0	0	3	3	4	3	4
Anthony (C.I. 2143)	0	4	4	0	4	4	4	4	4
Iowa 102 (C.I. 2464)	0	4	4	0	4	0	4	4	4
Avena brevis (C.I. 2762)	4	1	4	1	4	4	4	4	4
Red Rustproof (C.I. 775)	4	4	4	4	4	4	4	4	4
Sunrise (C.I. 982)	4	1	0	2	4	3	2	1	4
Black Algerian (C.I. 204)	4	3	1	4	4	4	2	0	3
Nortex (C.I. 2382)	4	4	0	4	4	4	4	4	4
Ruakura (C.I. 2052)	3	4	1	4	4	4	4	1	0
Early Burt (C.I. 2763)	4	3	0	4	4	4	2	4	4
Red Rustproof (C.I. 1805)	4	4	0	4	4	4	4	4	4
Green Mountain (C.I. 1892)	ō	ī	4	0	4	4	4	4	4
Fulghum (C.I. 650-203)	4	4	4	4	4	4	4	4	4
Burt (C.I. 2054)	4	4	4	4	4	4	4	4	4
White Tartar (C.I. 1640)	$\tilde{4}$	4	4	$\bar{4}$	4	4	4	4	4
Red Rustproof (C.I. 1640)	4	4	ō	4	4	4	4	4	4
Green Russian (C.I. 1978)	Â	4 4	4	4	4	4	4	4	4
	Õ	้า	Ā	ō	4	4	4	4	4
Iowa 46	ŏ	4	- 4	ĩ	4	ī	$\overline{4}$	4	4
Iowa 77 (C.I. 2813)	4	÷ 3	- <u>-</u> - /	4	4	4	ŝ	3	$\overline{4}$
Iomine (C.I. 2827)	生 人	3 3	14 1	4	4	4	4	ă	Ā
Hundred Bushel (C.I. 2797)	4	3	بار		-1			<u> </u>	-

Table 3. Reaction of 33 pure-line selections to the nine physiologic forms of <u>Puccinia coronata avenae</u> identified from collections made during 1927 and 1928.

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-23-

Table 3. Concluded.

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Pure-line selections of : varieties, selections,		rage	type	of	infec	tion	wit	n phy	s101	ogic	for	m m	mber
and species	: 1	:	2 :	3	: 4	?	5	: 6	:	7 :	<u>8</u> .		9
Early May (C.I. 2828)	4		4	1	4		4	4		4	4	• ··	4
A.C. No. 10 (C.I. 2829)	4		4	2	4		4	4		4	3		4
Guyra (C.I. 2768)	4		4	3	4		4	4		3	3		3
Narrigal (C.I. 2798)	- 3		3	1	3		4	3		3	2		3
Sidney (93)	4		4	1	4		4	4		4	4		1
Folden (C.I. 2806)	4		4	4	4		4	4		3	3		3
Mortgage Lifter (C.I. 2804)	4		4	3	4		4	4		4	3		4

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infection of each to the nine identified physiologic forms.

The nine physiologic forms discovered during 1927 and 1928 may be identified by arranging the first eight pure lines in table 3 in a dichotomous key (50).

Cultures Collected During 1929 and 1930

The first 17 pure-line-selected: varieties, selections, and species given in table 3 were retained as possible differential hosts for use during 1929 and 1930. Eight additional pure lines, seven varieties, and one species were added. Two of the foreign varieties, "Avena victoria" (C.I. 2764), and "Avena capa" (C.I. 2765), were obtained from Mr. Jose M. Scasso of Moron, Argentina, while "Avena 1095a" (C.I. 2766) and "Avena 64s" (C.I. 2767) were obtained from Doctor Alberto Boerger, Senior Director del Instituto Fitotecnico, Departmento Colonia, Uruguay. These four varieties were reported resistant to crown rust by Mr. Scasso*. The other three varieties Hay (C.I. 1622), Schoolmam (C.I. 2057), and Victoria (C.I. 2401) were included because of their unusual resistance during a natural epiphytotic of crown rust in the summer of 1929 at Ames, Iowa, and Manhattan, Kansas. Avena strigosa glaberscens (C.I. 2630) was apparently immane during the same epiphytotic at both stations.

*These varieties were reported as being resistant to crown rust in Argentine, by Mr. Scasso, in a personal letter to the writer dated April 20, 1929.

-25-

One hundred ninety cultures of crown rust were isolated from 116 collections during 1929 and the spring of 1930. Uniform groups of 25 pure lines, seven varieties and one species of oats were inoculated separately with each of these cultures and eight physiologic forms were identified. Four of these were forms previously identified in collections secured during 1927 and 1928, and four were first isolated in 1929.

In table 4 is given the reaction of pure lines, numbers 1 to 17, to physiologic forms numbers 1, 3, 5, 6, 10, 11, 12, and 13 collected during the years 1927, 1928, 1929, and 1930, and the reaction of pure lines, varieties, and species, numbers 18 to 33, for the same physiologic forms collected during 1929 and 1930. As physiologic forms 2, 4, 7, 8, and 9 were not recovered during 1929 and 1930, the reaction of only pure lines numbers 1 to 17 is recorded.

	1928, 1929, and 1930.					- Ollar	- 100			0.01	TTR:	4061	•	
	Number and name of pure-	2		Av	erage) ty		fin	oct	ion	with			
	line selections, varie-	:						for				•••	1.1.4	
	ties, and species	: 1	: 2	: 3							:10:	11	12:	13
1 2 3 4 5 6 7 8 9 0 11 2 3 4 5 6 7 8 9 0 11 2 3 4 5 6 7 8 9 0 11 2 3 4 5 6 7 8 9 0 112 15 6 7 8 9 0 112 15 6 7 8 9 0 112 15 15 15 15 15 15 15 15 15 15 15 15 15	Pure-line selections Belar (C.I. 2760) Red Rustproof (C.I. 1079) College Algerian (C.I. 2052) Cowra (C.I. 2761) Iowa 69 (C.I. 2463) Avena strigosa (C.I. 1782) Anthony (C.I. 2143) Iowa 102 (C.I. 2464) Avena brevis (C.I. 2762) Red Rustproof (C.I. 775) Sunrise (C.I. 982) Black Algerian (C.I. 204) Nortex (C.I. 2382) Ruakura (C.I. 2025) Early Burt (C.I. 2763) Red Rustpfoof (C.I. 1805) Green Mountain (C.I. 1892) Bathurst (C.I. 1810) Glenn Innis (C.I. 980)	****	4 4430244141344341	000404444010000404	34430000142444440	************	3444434044344444444	0443444442244244	01404344441041444	144144444340444	0000040044000100104	4444040444444044144	0000040044000000004	-27* -27*
20 21 22 23 24 25	Varieties White Russian (C.I. 2460) White Russian (C.I. 2461) Green Russian (C.I. 2342) Green Russian (C.I. 2344) Avena brevis (C.I. 1783) Markton (C.I. 2053)	044 404 04		444 4 4 4 0 4		44444	044444				04444	044444	044444	14444 444

Table 4. Reaction of 33 pure-line selections, varieties, and species to 13 physiologic forms of <u>Puccinia</u> coronata avenae identified during 1927, 1928, 1929, and 1930.

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412-1-1	Number and name of pure- line selections, varie-	* *	Ave			infoct. form nur	on with			, and a second
	ties, and species	:1:	2:3	4:5	:6:	7:8	9 :10:	11:	12:	13
	Varieties							•		
26	Hay (C.I. 1622)	4	4	4	4		4	4	4	4
27	Schoolmam (C.I. 2057)	4	4	4	4		4	4	4	4
28	Victoria (C.I. 2401)	0	. 0	0	0		I	0	I	0
29	"Avena victoria" (C.I. 2764)	0	0	0	0		I	0	I	0
30	"Avena capa" (C.I. 2765)	0 3	0	3	4		I	0	0	0
31	"Avena 1095a" (C.I. 2766)	4	0	4	4		0	1	0	0
32	"Avena 64s" (C.I. 2767)	3	0	3	4		0	0	0	0
33	Species Avena strigosa glaberscens (C.I. 2630)	0	0	0	0		4	0	I	0

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-28-

Key for Identification of Physiologic Forms

Thirteen physiologic forms were identified among the 245 cultures studied by using the differential reactions of eight pure lines and one species, <u>Avena strigosa glaberscens</u> (C.I. 2630). <u>A. strigosa glaberscens</u> (C.I. 2630) was not pure-line selected, however, it always gave a homozygous reaction to physiologic forms 1, 3, 5, 6, 10, 11, 12, and 13. The eight pure-line-selected: varieties, selections, and species; and one species not pure-line-selected, may be arranged in a dichotomous key as follows:

Belar (C.I. 2760) - resistant Red Rustproof (C.I. 1079) - resistant Iowa 102 (C.I. 2464) - resistant Avena strigosa glaberscens (C.I. 2630) -. . . form 12 Avena strigosa glaberscens (C.I. 2630) susceptible form 10 Iowa 102 (C.I. 2464) - susceptible College Algerian (C.I. 2052) - resistant Anthony (C.I. 2143) - resistant form 13 Anthony (C.I. 2143) - susceptible . . . form 3 College Algerian (C.I. 2052) - susceptible. form 8 Red Rustproof (C.I. 1079) - susceptible 9 Cowra (C.I. 2761) - susceptible form 7 Belar (C.I. 2760) - susceptible Anthony (C.I. 2143) - resistant Avena strigosa (C.I. 1782) - resistant. . . form 4 Avena strigosa (C.I. 1782) - susceptible Iowa 102 (C.I. 2464) - resistant. form 1 Iowa 102 (C.I. 2464) - susceptible. . . . form 11 Anthony (C.I. 2143) - susceptible Iowa 102 (C.I. 2464) - resistant form 6 Iowa 102 (C.I. 2464) - susceptible. . . . Iowa 69 (C.I. 2463) - resistant form 2 Iowa 69 (C.I. 2463) - susceptible form 5 Physiologic forms 1, 3, 5, 6, 10, 11, 12, and 13, may be identified by the differential reaction of the pure-line-selected varieties: Belar (C.I. 2760), Anthony (C.I. 2143), Iowa 102 (C.I. 2464); and the species <u>Avena strigosa glaberscens</u> (C.I. 2630). The reaction of each one of these differential hosts along with that of Victoria (C.I. 2401) and a pure-line selection of Markton (C.I. 2053), to each of the eight physiologic forms identified from collections secured during 1929 and 1930 is illustrated in plates III-VI.

Nature of Physiologic Forms Identified

The 13 physiologic forms identified, differ greatly in their ability to infect normally the possible differential hosts given in table 4. A descriptive formula may be given to each form by writing the number (given in table 4, column 1) representing each of the 33 possible differential hosts that show a resistant reaction to it. These formulae would then appear as follows:

Physiologic form number Formula 1 5, 7, 8, 17, 20, 24, 28, 29, 33. 2 *5, 6, 9, 11, 17. 1, 2, 3, 4, 6, 11, 12, 13, 14, 15, 16, 18, 24, 3 23, 29, 30, 31, 32, 33. 4 *5, 6, 7, 8, 9, 11, 17. 28, 29, 33. 5 6 8, 20, 28, 29, 33. 7 *1, 11, 12, 15. *1, 2, 4, 11, 12, 14. 8 9 *1, 4, 14. 1, 2, 3, 4, 5, 7, 8, 11, 12, 13, 14, 15, 16, 17, 18, 20, 28, 29, 30, 31, 32. 10 5, 7, 14, 17, 20, 28, 29, 30, 31, 32, 33. 11 1, 2, 3, 4, 5, 7, 8, 11, 12, 13, 14, 15, 16, 17, 18, 20, 28, 29, 30, 31, 32, 33. 1, 2, 3, 4, 5, 7, 11, 12, 13, 14, 15, 16, 17, 20, 12 13 28, 29, 30, 31, 32, 33.

The 13 physiologic forms, as shown by their ability to infect normally the 33 possible differential hosts given in table 4, rank from the most virulent to the least as follows: 5, 6, 9, 7, 1, 11, 2, 8, 4, 3, 13, 10, 12.

Physiologic form 5 is a virulent form. In addition to the possible differential has ts given in tables 3 and 4, approximately 400 additional pure lines, varieties, selections, and species have been inoculated with this form. With the exception of the varieties, Victoria (C.I. 2401) and "Avena victoria" (C.I. 2764), and the species <u>Avena strigosa glaberscens</u> (C.I. 2630), no resistant pure lines, varieties, selections, or species have been found. Form 6, apparently little less virulent than

*These physiologic forms were not identified in collections secured during 1929 and 1930. For that reason their reaction to numbers 18-33 is not known.

-31-

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form 5, was tested only with the possible differential hosts given in tables 3 and 4.

Physiologic forms 9, 7, 1, 11, 2, 8, and 4 are intermediate in virulence, while forms 3, 13, 10, and 12 are comparatively weak forms. Forms 10 and 12 differ only in reaction on <u>Avena strigosa glaberscens</u> (C.I. 2630).

Under similar conditions, cultures identified as physiologic forms 3, 10, 12, and 13 tended to form teliospores earlier than cultures identified as forms 1, 2, 4, 7, 8, 9, and 11. While cultures identified as forms 5 and 6 rarely showed any telicspore formation. Teliospore formation seems to be characteristic of narrowly specialized forms. It did not seem to be correlated with any particular pure lines, varieties, species, or selections, and it was not limited to plants showing a resistant type of infection. Certain plants with a type 4 infection consistently showed early teliospore formation with certain cultures. However, early teliospore formation occurred more frequently on plants showing a type 1 or type 2 infection. Different cultures belonging to the same physiologic form and growing under similar conditions would sometimes vary greatly in their ability to produce teliospores on certain pure-line selections and varieties. Certain cultures identified as form 12 consistently formed teliospores 10 to 12 days after inoculation on pure-line selections and varieties, numbers: 9, 10, 17, 20, 21,

-32-

and 26. While other cultures also identified as form 12 did not form teliospores on these same hosts until 21 to 33 days after inoculation. Apparently it would be possible to subdivide certain physiologic forms on the basis of their relative ability to form teliospores on certain hosts.

Plate VII illustrates the relative ability of two cultures to form teliospores. Two panieles of oats were selected, one from Fulghum (C.I. 650-203) and the other from Dookie Agricultural College No. 10 (C.I. 2829). Seed from these panieles were planted at the same time. The resulting seedling plants were inocalated and held under the same conditions until after the photograph in plate VII was secured. Two cultures of crown rust representing forms 3 and 5 were separately inoculated to individual plants from each of the two panieles. Sixteen days after inoculation the culture identified as form 3 had formed abundant telia on both selections, while the culture identified as form 5 had formed only uredinia. The relative ability of these two cultures to form telia was typical of all other cultures identified as forms 3 and 5.

-33-

PREVALENCE AND GEOGRAPHIC DISTRIBUTION OF PHYSIOLOGIC FORMS IDENTIFIED

Two hundred forty-five cultures of crown rust were isolated from 171 collections secured during the years 1927, 1928, 1929, and 1930. Each of these was identified as belonging to one of the 13 physiologic forms described in table 4. In table 5 is given the place of collection, name of collector, name of host from which the original collection was secured, and date of this original collection, for each of the 245 cultures identified. This information is grouped according to the physiologic form to which the culture belonged and in order of the date of collection.

Physiologi form	: .c: : Place collected	: : Collector	: : : Host	: Date : Collected
Puccinia		L.D. Leach	Cultivated oats	8-1-27
	Ames, Ia.	do	Volunteer oats	9-14-27
coronata	Clyde, Kan. Manhattan, Kan.	C.O. Johnson	do	9-18-27
orm 1		Fred Griffe	do	10-11-27
CTHL T	Stillwater, Okla.			2-6-28
	College Station, Tex.			4-14-28
	Hollywood, Calif.	J.M. Raeder	Volunteer oats	
	Redondo, Calif.	do	do	4-16-28
	San Diego, Calif.	do	do Ded Duratamont	4-23-28
	San Antonio, Tex.	do	Red Rustproof	5-12-28
	Napa, Calif.	L.D. Leach	Cultivated oats	7-2-28
	Experiment, Ga.	H.C. Murphy	Iomine	5-15-29
	do	do	do do	đo
	do	do	Sunrise (C.I. 982)	do
	Tifton, Ga.	do	Iomine	5-16-29
	do	do	do	do
	A.& M. College, Miss.	do	Anthony (C.I. 2143)	5-22-29
	do	do	Avena brevis (C.I. 1	
	, d o	do	Sunrise (C.I. 982)	do
	đo	do	do	đo
	Greenville, Tex.	đo	Nortex (C.I. 2382)	5-30-29
	Denton, Tex.	đo	Iomine	6-1-29
	do	do	do	do
	Lawton, Okla.	đó	Colburt (C.I. 2019)	6-4-29
	Manhattan, Kan.	do	Iomine	6-11-29
	do	do	do	do
	do	do	Anthony (C.I. 2143)	do
	do	do	Sunrise 73	do
	do	do	Rustless (C.I. 724)	do
	Columbus, Kan.	C.O. Johnston	Cultivated oats	6-15-29

Table 5. Source, date of collection and cultures of <u>Puccinia coronata</u> avenae identified.

Table 5. Continued.

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hysiologi	: .C:			: Date
form	: Place collected	: Collector	Host	collected
	Columbus, Kan.	C.O. Johnston	Cultivated oats	6-15-29
	Moron, Kan.	do	do	6-16-29
	do	do	do	do
	Parsons, Kan.	do	do	do
	Knoxville, Tenn.	T.R. Stanton	Green Russian	•
	đo	do	(C.I. 2344) do	6-19-29
				do
	do	do	White Tartar (C.I. 351)	do
	do	do	Biffen's White Wond	er
			(C.I. 2011)	do
	Blacksburg, Va.	do	Cultivated oats	6-21-29
	do	đo	do	do
	Colby, Kan.	C.O. Johnston	do	6-21-29
	Roanoke, Va.	Hutton and Stakm	an do	6-22-29
	do	do	đo	đo
	Harlan, Ia.	P.W. Rohrbaugh	do	do
	Booneville, Ia.	do	do	6-25-29
	Linn, Mo.	do	do	' đo
	do	đo	đo	do
	Nettleton, Mo.	do	do	đo
	Ottumwa, Ia.	do	do	6-27-29
	do	do	do	do
	Manhattan, Kan.	H.C. Murphy	Avena strigosa (C.I 1782)	• do
	do	do	do	do
	do	do	Sunrise (C.I. 982)	do
	do	do	Sunrise Sel. 73	do
	Sioux Falls, S.D.	do	Kherson	7-3-29
	La Vorne, Minn.	do	do	do
	Mankato, Minn.	do	Silvermine	7-5-29

-36-

Physiolog	: :			t
form	: Place collected	: Collector	: Host	: Date :collected
· •••	Iowa Falls, Ia.	H.C. Murphy	Kherson	7-6-29
	Ames, Ia.	do	Sunrise Sel. 73	7-17-29
	do	do	Cultivated oats	8-4-29
	do	do	do	do
	Corvallis, Ore.	E.N. Bressman	Iogold (C.I. 2329)	8-24-29
	do	do do	do	d o
	do	do	Cultivated oate	do
	Morgantown, W.Va.	H.C. Murphy	Black Mesdag (C.I.	uo
	morganoonn, a.va.	meo • marpiny	1877)	9-17-29
	đo	do	Gopher x Black Mesdag	
	do	do	Hybrid oats	do
	Gainesville, Fla.	A.H. Eddins	Cultivated oats	3-3-30
	Leesburg, Fla.	W.B. Tisdale	Red Rustproof	3-14-30
	Gainesville, Fla.	A.H. Eddins	Cultivated oats	3-25-30
	do	do	do	00-00-00 d0
Puccinia				
coronata	Libson, N.D.	S.M. Dietz	Volunteer oats	8-26-27
avenae				
form 2		يسور ب فالقرب في المراجع الألف بالمراجع المالية المراجع المراجع المراجع المراجع المراجع المراجع المراجع المراجع	and a start of the	
Puccinia	Ste Anne de la			
coronata	Pocatiere, Que., Can.	H.B. Humphrey	Victory (near Rhammus	3
avenae			cathartica	8-23-27
form 3	Jackson Co., Ia.	J.M. Steddens	Rhamnus cathartica	5-23-28
	Millard, Nebr.	P.W. Rohrbaugh	do	5-27-28
	Waverly, Ohio	J.W. Baringer	Cultivated oats	7-13-28
	Montrose, W.Va.	S.H. Murphy	Iogold (C.I. 2329)	7-16-28
	Kingston, Ont.	W.P. Raleigh	Cultivated oats	7-29-28
	Amos, Ia.	H.C. Murphy	Iowa 102 (C.I. 2464)	9-23-28
	do	do	do	do
	do	do	Rhamnus infectoria	
			(aecial cup No. 10)	4-26-29

Table 5. Continued.

-37-

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Physiolog form	ic: Flace collected	: Collector	: : : Host	: Date
1011	; FIACO COLLOCION		: HOSU	:collected
- · · •	Harlan, Ia.	P.W. Rohrbaugh	Cultivated oats	6-22-29
	Farmington, Ia.	do	do	6-25-29
	Nettleton, Mo.	do	do	do
	Ames, Ia.	H.C. Murphy	Oats near Rhamus	
	•		cathartica	7-12-29
	đo	do	ob	đo
	do	do	Markton (C.I. 2053)	7-17-29
	do	đo	do	do
	Ryder, N.D.	A.A. Ziegler	Rhamnus cathartica	8-2-29
	đo	đo	do	do
	do	do	do	do
	đo	do	do	đo
	Morgantown, W.Va.	H.C. Murphy	Hybrid 17-10-48	9-17-29
	do	do	do	do
	dó	do	Gopher x Black Mesdag	do
	Ames, Ia.	do	Iowa 102 (C.I. 2454)	10-2-29
	do	do	do	âo
	do	do	Iowa 444 (C.I. 2331)	do
	do	do	Rhamnus cathartica	3-27-30
	do	do	do	đo
	do	do	do	3 - 29 -30
	đo	do	do	do
	do	do	do	3-30-30
	do	do	Rhamnus infectoria	3-30-30
·····	do	do	do	do
uccinia	Urbana, Ill.	L.D. Leach	Volunteer oats	9-14-27
oronata				
venae				
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-38-

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Table 5. (Continued.
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Physiologi				: Date
form	: Place collected	: Collector	: Host	:collected
Puccinia	Fulton, No.	L.D. Leach	Volunteer oats	9-18-27
coronata	Mason City, Ia.	H.C. Murphy	d do	10-13-27
avenae	Del Rio, Tex.	J.M. Raeder	Cultivated oats	5-9-28
form 5	Nashville, Tenn.	S.M. Dietz	do	5-17-28
	Gainesville, Fla.	A.H. Eddins	do	5-18-28
	Afton, Ia.	F.H. Mendell	Rhamnus lanceolata	5-21-28
	Experiment, Ga.	S.M. Dietz	Winter Turf	5-23-28
	Tifton, Ga.	đo	Cultivated oats	5-24-28
	A. & M. College, Miss.	do	do	5-26-28
	Denton, Tex.	do	Anthony (C.I. 2143)	5-27-28
	do	đo	Cultivated oats	5-28-28
	Gainesville, Tex.	do	đo	5-29-28
	Stillwater, Okla.	đo	do	5-30-28
	Watonsville, Calif.	L.D. Leach	Cultivated oats	5-31-28
	Experiment, Ga.	J.S. Hadden	Guyra	6-9-28
	đo	đo	Laurel Skinless	do
	A. & M. College, Miss.	D.C. Neal	Iowa 102 (C.I. 2464)	do
	do	do	Iowa 69 (C.I. 2463)	do
	do	do	Iowa 77	đo
	Ames, Ia.	H.C. Murphy	Rhamnus lanceolata	6-15-28
	do	do	do	d o
	do	do	do	6-17-28
	do	do	do	do
	do	do	do	6-18-28
	Astoria, Ore.	H.B. Humphrey	Cultivated Oats	6 -25-28
	Canton, Mo.	P.W. Rohrbaugh	do	6-29-28
	Sac City, Ia.	Dietz and Murphy	do	7-7-28
	Hinton, Ia.	do	do	7-8-28
	Devil's Lake, N.D.	H.B. Humphrey	Avena fatua	7-27-28
	Ames, Ia.	H.C. Murphy	Iowa 102 (C.I. 2464)	9-23-28
	do	do	do	đo

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-39-

Table 5. Continued.

Physiologic:	•	•	i. Dokt
form : Place collected	Collector	Host	: Date collected
Ames, Ia.	H.C. Murphy	Rhamnus infectoria	•••••
		(aecial cup No. 4)	4-26-29
do	do	do, (aecial	
		oup No. 8)	do
Knoxville, Tenn.	đo	Iomine	5-13-29
do	do	do	do
do	do	Anthony (C.I. 2143)	đo
do	do	do	do
Experiment, Ga.	do	do	5-15-29
do	do	do	do
Tifton, Ga.	do	do	5-16-29
do	do	do	do
Auburn, Ala.	do	Cultivated oats	5-20-29
do	đo	do	do
A. & M. College, Miss.	, do	Iomine	5-22-29
do	ob	do	do
do	do	Anthony (C.I. 2143)	do
do	do	Avena brevis	
		(C.I. 1783)	đo
đo	do	Sunrise (C.I. 982)	do
Denton, Tex.	do	Anthony (C.I. 2143)	6-1-29
đo	do	do	do
do	do	Sunrise (C.I. 982)	do
Lawton, Okla.	do	Colburt (C.I. 2019)	6-4-29
Stillwater, Okla.	do	Iomine	6 -6- 29
do	do	do	do
do	do	Anthony (C.I. 2143)	do
do	đo	do	do
Manhattan, Kan.	do	do	6-11-29
do	do	Sunrise Sel. 73	do
Knoxville, Tenn.	T.R. Stanton	White Tartar (C.I. 5	

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-40-

P.7.3 P.6.5 (177 P.5.4)

Table 5.	Continued.
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Physiologi	:		•	Date
form	: Place collected	Collector	: Host	:collected
··· •	Knoxville, Tenn.	T.R. Stanton	Biffen's White Wonder	•
	•		(C.I. 2011)	6-19-29
	do	do	Iowa 77	đo
	do	do	do	đ.o
	Colby, Kan.	C.O. Johnston	Cultivated oats	6-21-29
	Hamburg, Va.	Hatton and Stak-		
		man	đo	6-22-29
	do	do	do	do
	Booneville, Ia.	P.W. Rohrbaugh	do	6-25-29
	Manhattan, Kan.	H.C. Murphy	Sunrise (C.I. 982)	6-27-29
	do	do	Sunrise Sel. 73	do
	La Verne, Minn.	do	Kherson	7-3-29
	Mankato, Minn.	do	Silvermine	7-5-29
	Ames, Ia.	do	Iomine	7-17-29
	do	đo	Victoria (C.I. 2401)	do
	do	do	Sunrise Sel. 73	đo
	Corvallis, Ore.	E.N. Bressman	Cultivated oats	8-24-29
	Morgantown, W. Va.	H.C. Murphy	Black Mesdag	9-17-29
	Ames, Ia.	do	Iowa 102 (C.I. 2464)	10-2-29
	do	do	do	do
	do	do	Volunteer oats	do
	đo	do	do	do
	do	do	do	do
	Gainesville, Fla.	A.H. Eddins	Cultivated oats	3-3-30
	do	do	do	do
	Leesburg, Fla.	W.B. Tisdale	Red Rustproof	3-14-30
	do	do	do	do
	Gainesville, Fla.	A.H. Eddins	Cultivated oats	3-25-30
	do	do	do	do
	Ames, Ia.	H.C. Murphy	Rhamnus lanceolata	3-30-30
	do	do	do	đo

-41-

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Table 5. Continued.

Physiolog	: a:	e e	•	: Date
form	: Place collected	: Collector	: Host	:collecte
	Gainesville, Fla.	A.H. Eddins	Cultivated oats	4-3-30
	do Amo a Ta	do II () Viermahar	do Thanna ta da ata ata	do
Puccinia	Ames, Ia. Denton, Tex.	H.C. Murphy P.B. Dunkle	Rhamnus infectoria Volunteer oats	4-5-30
coronata	Iowa Falls, Ia.	H.C. Murphy	Kherson	11-6-27 7-6-29
venae	Ames, Is.	do	Sunrise (C.I. 982)	7-17-29
form 6	do	do	do	do
Puccinia	Ames, Ia.	H.C. Murphy	Rhamnus tinctoria	6-15-28
coronata				0-10-00
venae				
form 7				
Puccinia	Ames, Ia.	H.C. Murphy	Rhamnus spp. (from	6-15-28
coronata		-	Mont:)	
avenae				
form 8				
Puccinia	Ames, Ia.	H.C. Murphy	Rhamnus infectoria	6-15-28
coronata				
torm 9				
Puccinia	Farmington, Ia.	P.W. Rohrbaugh	Cultivated oats	6-22-29
coronata	Montrose, W.Va.	H.C. Murphy	Volunteer oats	9-1-29
venae	do	do	đo	đo
<u>orm 10</u>	Morgantown, W.Va.	do	Gopher (C.I. 2027)	9-17-29
	do	đo	do	do
	do	do	Hybrid oats	do
	do	đo	Hybrid 17-10-1	do
	Ames, Ia.	do	Iowa 444 (C.I. 2331)	10-2-29
	Conesville, Ia.	D.V. Layton	Volunteer oats	10-6-29
	do	do	do	do
	Ames, Ia.	H.C. Murphy	Rhamnus tinctoria	3-27-30
	do	do	Rhamnus spp. (from Mont.)	

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Physiolog		:	· · · · · · · · · · · · · · · · · · ·	: : Date
form	: Place collected	: Collector	: Host	:collected
Puccinia	Parsons, Kan.	C.O. Johnston	Cultivated oats	6-16-29
coronata	Bowling Green, Mo.	P.W. Rohrbaugh	do	6-26-29
avenae	do Taritar alla Ta	do	do	do
form 11	Indianola, Ia.	do	do	6-29-29
	do Kartana ///	do M. G. Marmilian	do Malaata aata	do
	Montrose, W. Va.	H.C. Murphy	Volunteer cats	9-1-29
	do	do	do	do
	Morgantown, W. Va.	do	Hybrid 17-10-1	9-17-29
Puccinia	Ames, Ia.	H.C. Murphy	Rhamnus spp. (from	4 00 00
coronata	•	-	Mont.)	4-26-29
avenae	do	do	Rhamnus infectoria	-
form 12	_	-	(aecial cup No. 1)	do
	do	do	do (aecial cup No.3)	do
	do	do	do (aecial cup No.5)	do
	do	do	do (aecial cup No.6)	do
	do	do	do (aecial cup No.7)	do
	do	do	do (aecial cup No.9)	do
	do	do	do (aecial cup No.11)	do
	do	do	do (aecial cup No.12)	
	do	do	do (aecial cup No.13)	do
	do	do	Rhamnus spp. (from	
			Mont.)	4-27-29
	do	do	do	do
	do	do	do	3-27-30
	do	do	Rhamnus infectoria	do
	do	do	Rhamnus tinctoria	do
	do	do	do	3-30-30
	do	do	Rhamnus spp. (from	
			Mont.)	do
	do	do	Rhamnus tinctoria	đo

-43-

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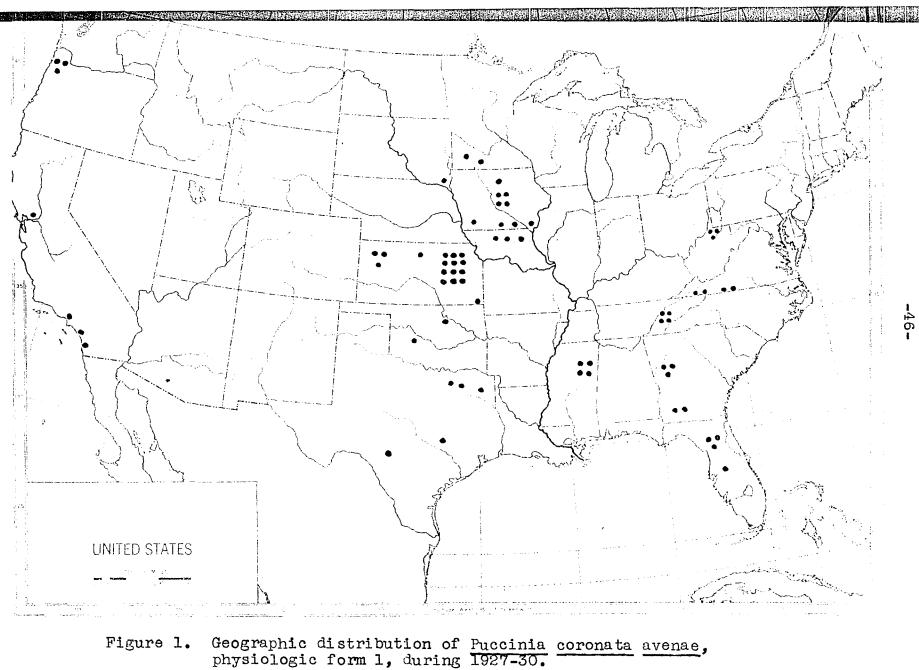
Physiologi form	.c: Place collecte	d Collector	: : : Host	: Date : Date :collected
Puccinia coronata	Ames, Ia.	H.C. Murphy	Rhamnus infectoria (aecial cup No. 2)	4-26-29
avenae	đo	do	Rhamnus infectoria	4-5-30
form 13	do do	d o d o	Rhamnue chlerophera do	4-5-30 do

Seventy cultures collected during the years 1927, 1928, 1929, and 1930 wore identified as physiologic form 1. This form is apparently widely distributed throughout the United States and common in occurrence. (Fig. 1). Form 1 was not identified in cultures collected from Rhamnus and its distribution may be independent of an alternate host. It apparently hibernates in the south and probably moves north during the spring and summer.

Physiologic form 2 was identified in only one culture, collected in North Dakota in 1927 by Dr. S. M. Dietz. It may have originated on Rhamnus and it is apparently a comparatively rare form.

Thirty-three cultures were identified as physiologic form 3. Although a comparatively weak form and limited to the central and northeastern oat-growing regions of the United States, form 3 is important because few of the northern oat varieties are resistant to it. The principal source of collection of form 3 was on <u>Rhamnus cathartica</u>, both naturally and artificially infected. It was also collected on oats in Iowa, Ohio, West Virginia, Ontario, and Quebec. However, all of the cultures collected from oats were either, from plants growing near infected <u>R</u>. <u>cathartica</u>, or from a region where this species is common. Therefore, it seems probable that form 3 is disseminated from <u>R</u>. cathartica.

-45-



<u>Rhammus cathartica</u> bushes are widely distributed in this northern cat growing region and will probably continue to furnish a source of inoculum of form 3. Dietz (20) has shown that <u>R. cathartica</u> is effective in disseminating crown rust. <u>Rhammus</u> infectoria, another host for this form, is not native in America.

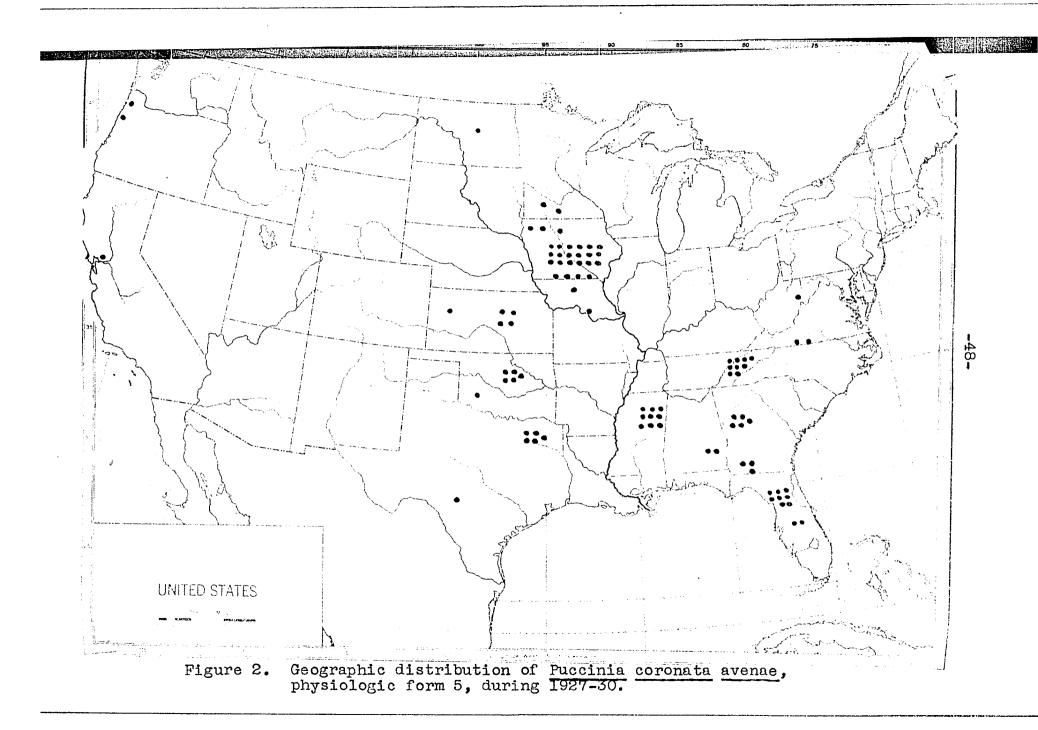
Physiologic form 4 was collected only once, at Urbana, Illinois, by Mr. L.D. Leach. It is somewhat similar to form 2 in virulence.

The most prevalent, widespread, and virulent form studied is number 5. Ninety-one of the 245 cultures collected during 1927, 1928, 1929, and 1930, were identified as belonging to this form. It is common in occurrence throughout all regions of the United States from which collections were secured (Fig.2). This form was collected on oats and on both <u>R. lanceolata and <u>R.</u> <u>infectoria</u>. The former, which is widely distributed in the central portion of the United States, probably disseminates this form. Form 5 hibernates in the south and probably moves northward during the spring and summor.</u>

Physiologic form 6, collected only on oats, was identified in cultures: once from Denton, Texas, in 1927; once from Iowa Falls, Iowa, in 1930, and twice from Ames, Iowa, in 1930.

Physiologic forms 7, 8, and 9, collected respectively from R. tinctoria, R. spp. (from Montana), and R. infectoria, were each identified once.

-47-



Physiologic form 10, identified in 12 cultures, ten from Avena in West Virginia and Iowa, and two from Rhamnus, one from <u>R. infectoria</u> and one from <u>R. spp.</u> (from Montana), is apparently a comparatively rare form.

Physiologic form 11, a comparatively rare form, was identified in eight cultures collected in Kansas, Missouri, Iowa, and West Virginia. This form was not collected on Rhamnus.

Physiologic forms 12 and 13 came only from Rhamnus. Form 12 was identified in 18 cultures isolated from collections secured from <u>R. infectoria and R. tinctoria</u>. Form 13 was identified in four cultures coming from <u>R. infectoria</u>, <u>R. chlorophora</u>, and <u>R. spp.</u> (from Montana).

The prevalence of each of the 13 forms during the years 1927, 1928, 1929, and 1930 and their distribution on Rhamnus and Avena hosts is given in table 6.

			cultur						•
Physiologic	: 19			928		929	: 19		: Total
form number	Avena	Rhamm	ls Avena	Rnamnu	stavena	Knumnu	s:Avena	Rhamnus	
l	4	-	6	-	56		4	-	70
2	1	-	-	-		-	-	-	1
3	1	-	3	4	13	5	-	7	3 3
4	1	-			-		-	-	1
5	2	**	23	6	47	2	8	3	91
6	1	-	-	-	3	-		-	4
7	-	-	-	1	-			*	1
8	**			l	-	-	-		1
8	-	•	-	l				-	1
10			-	-	10	-	-	2	12
11	-				8		-	-	8
12	-	-	-	-		12	-	6	18
13	-		-	**	-	1		3	4
l'otal									
per species	10		32	13	137	20	12	21	245
Total	-	-							
p er yea r	1	0	45		1	57	33	3	245

Table 6. Distribution of 13 physiologic forms of <u>Puccinia</u> coronata avenae on Avena and Rhamnus hosts during 1927, 1928, 1929, and 1930.

Eight of the 13 forms were each identified in more than one culture. In order of their prevalence, these forms are 5, 1, 3, 12, 10, 11, 15, and 6. Forms 1, 2, 4, 6, and 11 were collected only from Avena species. While forms 7, 8, 9, 12, and 13 were collected only from Rhamnus species. Forms 3, 5, and 10 were collected on both Rhamnus and Avena.

Physiologic Forms Collected on Rhamnus

The distribution of the forms obtained from Rhamnus according to the species from which they were collected is given in table 7.

Table 7. Physiologic forms identified and number of cultures collected from different species of Rhamnus during 1928, 1929, and 1930.

-	Rhamnus host	:	Nu				iltu umb		10	ient	ified	
		: 3	:	5 :	7	:	8	:	9	: 10	: 12	: 13
R.	cathartica	13		-			-	,	-	-	-	-
R.	chlorophora	-			-				•	-	-	2
R.	infecto ria	3		3	-		-		1	-	10	2
R.	lanceolata	-		8	-		-		-		••	-
R.	spp. (from Montana)	-			-		-		-	-	-	2
R.	tinctoria	-		-	1		-			1	3	-

In these data certain species of Rhamnus produced only one physiologic form of crown rust, while other species produced as many as five. Both naturally and artificially infected plants of <u>Rhammus lanceolata</u> have consistently produced only physiologic form 5. While naturally and artificially infected plants of <u>R. cathartica</u> have consistently produced only form 3. A total of five physiologic forms have been isolated from <u>R.</u> <u>infectoria</u>. In the light of the investigations conducted by Dietz (21), Craigie (14, 15, 16, 17, 18), Hanna (33), Newton, Johnson and Brown (51), Stakman, Levine and Cotter (64), and Allen (1), it seems probable that certain of these forms, particularly those of rare occurrence, may have arisen as a result of hybridization.

On September 23, 1928, a collection of volunteer Iowa 102 (C.I. 2464) oat plants was secured at Ames, Iowa. These plants heavily infected with crown rust, both in the telial and uredinial stages. Nine different single-uredinium cultures were identified, from the urediniospore material collected, using the possible differential hosts given in table 4. Six of these cultures were found to be physiologic form 5 and three physiologic form 3. Early in March, 1929, the telial material of this same collection was used to inoculate plants of <u>R. infectoria</u> and <u>R.</u> <u>spp</u>. (from Montana). Abundant infection was secured on both species. A total of 16 unopened accial cups, 13 from <u>R. infectoria</u> and 3 from <u>R. spp</u>. (from Montana), were carefully picked with sterilized forceps. The acciospores from each of these accial cups were used to inoculate Markton plants and the single-uredinium

-52-

isolations were identified in the usual manner. The thirteen cultures obtained from the same number of aecial cups from <u>R</u>. <u>infectoria</u> were identified as follows: nine were form 12; two were form 5; one was form 3, and one was form 13. The three cultures from <u>R</u>. <u>spp</u>. (from Montana) were identified as form 12.

The above evidence, although circumstantial, strongly indicates that new physiologic forms are being produced on certain species of Rhamnus. This may partly account for the number of new forms collected only on Rhamnus. The fact that the teliospores were collected from a pure-line selection of Iowa 102 (C.I. 2464), which is highly resistant to form 12, is still further evidence that this form was probably not present in the original collection and that it may have originated on <u>R. infectoria</u> and <u>R. spp.</u> (from Montana).

Physiologic Forms Collected on Oats

Crown rust of oats probably does not hibernate farther north than the northern limit of over-wintering of volunteer oats. Those physiologic forms that hibernate in this southern region should be available for collection before those forms that originate on Rhamnus have appeared. Physiologic form 1 was collected as early as February 6 at Denton, Texas, during 1928, and both forms 1 and 5 were collected March 30, 1930, at Gainesville, Florida. The occurrence of the thirteen physiologic

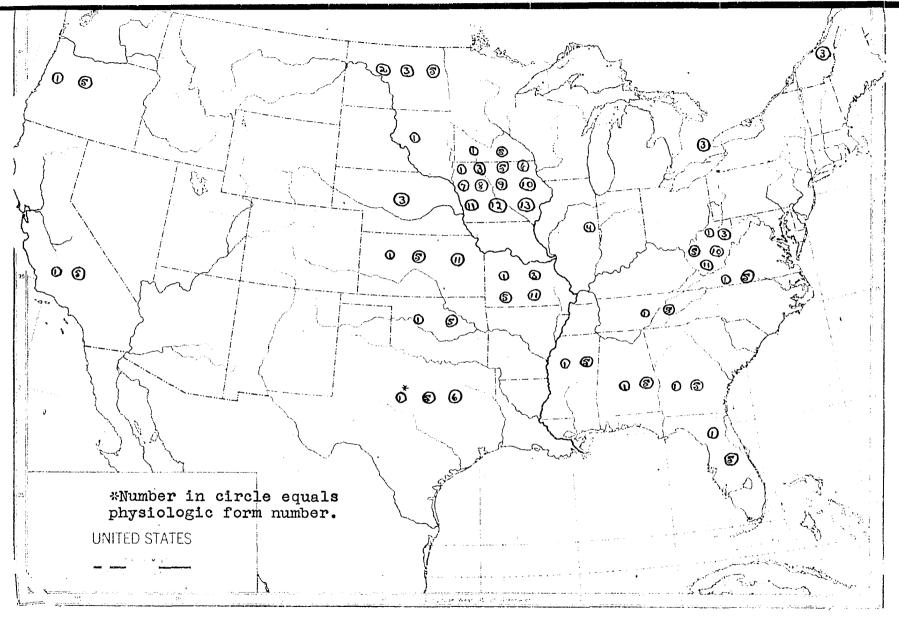


Figure 3. Physiologic forms of Puccinia coronata avenae collected from different states and Canada during the period 1927-30.

-54 -

RESPONSE OF VARIETIES, SELECTIONS, AND SPECIES

It is very desirable to know the reaction of the most important agronomic variables of oats to the most widely distributed physiologic forms. The forms identified most frequently were numbers: 1, 3, 5, 10, 11, and 12.

A uniform oat crown rust nursery was grown at Ames, Iowa, and Experiment, Georgia, during 1926, 1927, and 1928, and again in 1929, with a few changes, at Ames, Iowa, Experiment, Georgia, Knoxville, Tennessee, Tifton, Georgia, A. & M. College, Mississippi, Baton Rouge, Louisiana, Denton, Texas, Stillwater, Oklahoma, and Manhattan, Kansas. The per cent of crown rust infection on each of the varieties, selections, and species included in these various nurseries was determined. In addition the seedling reaction of these same varieties, selections, and species, to physiologic forms 1, 3, 5, 10, 11, and 12 was determined under greenhouse conditions.

In table 8 is given the average type of infection on each of these varieties, selections, and species with physiologic forms 1, 3, 5, 10, 11, and 12, and the percentage of crown rust infection observed on these same varieties, selections, and species, under field conditions at various locations in the United States. The nurseries omitted either escaped infection or the observations were untimely for satisfactory comparisons.

-56-

Variety, solection,	: : : C.I.	7	800	dlin	type ; read	or stion ic fo	126	r Tapor	TPARE	*	Statjfappinget jarrage tek	Denglinis tiggi viviline = tiget annual	various	of crown locations n:A. & M.
Or.	: munder	•	ŰV		inder Inder		a 642	: Geor	VIA		G. JOW		: Kanaas	1 Missi
specios		I	13				12				-1927a		: 1029	1
lebama Hybrid	971	4	4	4	4	4	4	73	10	70	30	55	30	
lbion	729	4	4	4	4	4	4					25	50	
loot White (64)**		4	4	4	O	4	0	40	t	70	40	26	50	•
lgerian	3570	4	1	4	4	4	4	20	3	20	80	10	15	
do	640	4	0	4	0	4	0					20	30	
âo	2052	4	0	4	1	1 4	1					8	30	
đo	989	4	4	4	4	4	4					25	25	
lgerian (69)		4	0	4	4	4	4	35	t	25	66	15	25	
leerian (140)		4	4	4	1	4	1					15	55	
lgerian Cape	1002	4	0	4	0	1	8					5	35	
lgorian z Calcutta	2807	4	影	4	4 0 2	4	4	5	5		20	20	40	
nthony	2143	0	4	4	0	0	0					15	46	£
Argentine	993	4	4	4		2	2					5	60	
UTONA	831	4	4	4	4	4	4					60	65	
wnless Culted	1776	4	4	4	4	4	4					25	60	
wnless Konarch	1879	4	4	4	4	4	4					15	35	
wnless Probstier	1888	4	4	4	4	4	4					15	40	
Awnless Kustproof	1776	4	4	4	4	4	4					35		
Avena abbysinica (225)	0	4	4	4	4	4					18	50	
brevis	1783	Ö	0	4	4	4	4					5	5	
L. brevis (70)	2762	З	0	4	4	3	4					10	10	
. brovis (220)		4	3	4	3	é	8					t	10	
T. strigosa (226)		4	4	4	4	4	4					t	25	
do	1782	4	0	3	4	4	\$					5	15	
A. strig. Elaborscone	2030	0	0	0	4	0	0					0	Ö	
7. steriis (220)	•	4	4	4	4	4	4					60	45	

Table 8. Heaction of certain variaties, selections and species of outs to six physiologic forms of functinia coronate avenue, and the per cent of orown rust infection on these variaties under field conditions. .

lections and species of outs to six onsta uvenue, and the per cent of crown under field conditions.

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of otion ic for	1.12 1	: : Caor	inont		s, Iow	1	various la	of crown rust 1 Cations during A. & M. Colleg Mississippi	the years	i indica	ted Stillwater
. 11	15	:10276	1928	1998%	19272	10150	1929	19//9	: <u>1929</u>	1929	1929
4	4	73	10	70	30	55 25	30 50	50	5	20	80
4 4	04	40 20	t 1	70 20	40 20	25 10	50 15	70 10	30 t	35 10	16 20
4	0 1 4					20 5 25	30 30				
1 4 4	4	35	t	25	55	15 15	25 25 35	10	t	15	16
1 4	8 4	5	¢n		20	5 20	చర 40	t	5	t	t
0 2 4	024					15 5 50	45 60 65	25	40	20	10
4	444					23	6 0 35				
4 4						15 15 35 15	40				
4 4 3	4 4 4 4					5 10	50 5 10	0	0	0	Ó
6 4	2440					t t 5	10 25	-		v	0
4 0 4	6 0 4					0 40	15 0 46				

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Table 8. Continued.

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Varioty,	1 1	S	est].	ing	roact	tion		a			بيردخية برزيرتي والككار		various		
selection,	: C.I.	00 1	DEIY	8101	ogae	20Pm		: Export	lend	1		\$	Fanhatta	nia*	
or Spocies	:munber			naunb		1 11	. 19	- 0901 - 10901	STR DE	• • • • • • • • • • • • • • • • • • •		i:1929:	<u>Kansas</u> 1929	nun	163
	and the second sec				and the state of the						and a start of the starter				
"Avena capa"	2765	3	0	3	I	0	0								
"Avena victoria"	2764	0	Ö	0	I	0	ð								
"Avena 048"	2767	3	0	3	0	0	0								
"Avena 1095a"	2766	4	0	4	0	1	0								
Avoine de Barco	1789	4	4	4	0	0	0						10		
Batharst	1810	6	0	4	0	4	0	15	t	55	5	8	10		
Belar Sel. (1)		4	0	4	0	4	0					8 t 5 5	10		
Bolar (65)	276 0	4	0	4	0 1 4	4	0	20	1	13	10	5	15		
Bolar (S2)	2770	4	1	4	1	4	1	15	1	20	80		15		
Belyak	1630	4	4	4	4	4	4					35	50		
	206-151	4	8	4	Ö	4	0					40	50		
Biffen's White Winte		4	4	4	4	4	4	60	10	85	\$15	30	50		
do	2012	4	4	4	4	4	4	60	5	85	60	45	60		
Black_Algerian	204	4	1	4	O	4	0	40	5	15	20	5	5		
đo	640-1	4	4	4	Ö	4	o	40	â	00		15			
do	2051	4	3	4	4	4	4	40	2	20	60	5	30		
Black Bell I	1767	4	ġ	4	4	4	4					15	50		
Black Bell II	1308	4	4	4	4	đ	4					25	50		
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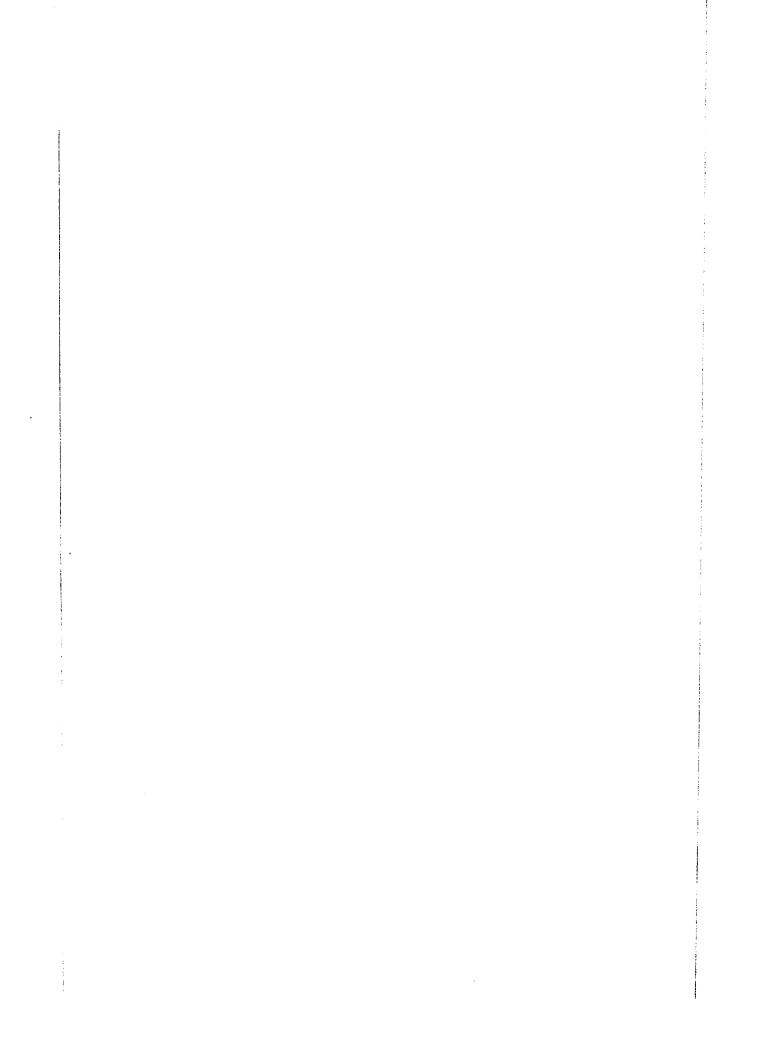
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Whe data for 1920 and 1927 were collected by Dr. 3.M. Dietz, Botany Department, Iowa State College, Ames, Iowa, and by L. D. Leach, Flant Pathology Department, California Agricultural Experiment Station, Davis, California. **Numbers in parenthesis refer to 1929 row numbers at Ames, Iowa. .

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A total of 316 varieties, selections, and species were inoculated with physiologic forms 1, 3, 5, 10, 11, and 12. Physiologic form 5 was again the most virulent form, only two varieties and one species being resistant to it. Victoria (C.I. 2401) and "Avena victoria" (C.I. 2764), apparently identical or similar varieties, were both completely resistant to form 5. <u>Avena strigosa glaberscens</u> (C.I. 2630) was also completely resistant to form 5.

Sixteen varieties and selections, distributed among the following species: Avena abbysinnica, A. brevis, A. byzantina, A. sativa, A. strigosa, and A. strigosa glaberscens, were resistant to physiologic form 1. Next to form 5 this was the most virulent form tested.

Form 11 was of medium virulence. Thirty-one varieties, celections, and species were resistant to it. Ten of the varieties, selections, and species resistant to form 1 were also resistant to form 11.

Forms 3, 10, and 12 were comparatively weak forms. A total of 77, 76, and 74 variaties, selections, and species (including those in common) were, respectively, resistant to these forms. Forms 10 and 12 are very similar in their reaction on the 316 variaties, selections, and species tested. Two of the three variaties that are resistant to forms 1, 5, and 11 are also resistant to these two forms. The oat variaties belonging to the species <u>A</u>. <u>byzantina</u>, and especially the Red Rustproof varieties, are more resistant to form 3 than to the other five forms tested.

Only the two varieties Victoria (C.I. 2401) and "Avena victoria (C.I. 2674) were resistant to the six forms tested. <u>Avena strigosa glaberscens</u> (C.I. 2630) was resistant to all except form 10, and Calcutta x Cape (C.I. 2802) was resistant to all except form 5. Twenty varieties were resistant to four of the six forms used. Ninety-nine of the 316 varieties, selections, and species tested were resistant to one or more physiologic forms.

Because of the prevalence and wide distribution of form 5, it is difficult to correlate seedling reaction in the greenhouse with percentage of infection in the field. Those varieties and species that were resistant to form 5 were uniformly resistant in all field tests. Many varieties that were uniformly susceptible to all physiologic forms in the greenhouse show a low percentage of infection under certain field conditions. However, a low percentage of infection is apparently not always due to physiologic resistance. Usually the type of infection on these plants would be "completely susceptible", and when artificially inoculated with spores produced on themselves they would produce a larger percentage of infection, depending upon the amount of inoculum used. The resistance in such instances is probably functional instead of physiologic.

A number of the Red Rustproof varieties produced a relatively low percentage of infection under field conditions, while adjoining plants of another variety would produce a high percentage of infection. Numerous collections from the Red Rusproof and adjoining plants were all identified as either physiologic form 1 or 5. When Red Rustproof plants of these same varieties, of equal age were inoculated under greenhouse conditions with either form 1 or form 5 they produced a high percentage of infection. But under field conditions with an abundance of the same inoculum, they were apparently partially resistant, at least at certain locations. A variety would often show a low percentage of infection at one station and a high percentage of infection at another where the same forms were apparently present in equal abundance. It would seem that this is not true physiologic resistance and that it is affected by environmental conditions.

Certain variaties and species were outstanding for their resistance to crown rust under field conditions. Victoria (C.I. 2401) was observed during 1929 at Ames, Io.a, and at Manhattan, Kansas. At Manhattan, Kansas, it showed a trace of crown rust infection of type O-1, while at Ames, Iowa, there was a trace of type O. The unusual field resistance of Victoria (C.I. 2401), combined with its resistance to eight of the 13 physiologic

-70-

forms described, should make this variety very valuable for hybridization and selection.

Avena strigosa glaberscens (C.I. 2630) showed a trace of type O infection at Ames, Iowa, and Manhattan, Kansas, during 1929. It was "completely resistant" or "immune" to all except one of the eight physiologic forms identified during 1929 and 1930.

Other varieties and species showing a comparatively low percentage of crown rust infection under field conditions were: White Russian (C.I. 2460 and 2461), Green Russian (C.I. 1040 and 2344), Glenn Innis (C.I. 980), Belar (C.I. 2760), <u>Avena</u> <u>brevis</u> (C.I. 2762), Red Rustproof (C.I. 1079 and 1089), Schoolmam (C.I. 2057), Sunrise (C.I. 982 and 1799), Algerian (C.I. 3579), and Nortex (C.I. 2382). While none of these varieties or species were equal to Victoria (C.I. 2401) in resistance during 1929, they were notable for their low percentage of infection as compared to the average commercial variety.

GRAMINEOUS HOST RANGE OF CERTAIN

PHYSIOLOGIC FORMS

An attempt was made to determine the gramineous host range of physiologic forms 1, 3, 5, 10, 11, and 12. As many as possible of the various gramineous species that have been reported as hosts for <u>Puccinia coronata</u> were secured. The seedling reaction of each of these, and other species, to the above mentioned physiologic forms was determined.

The reaction of certain species of grass to a particular physiologic form of crown rust was not nearly as constant as was the reaction of oat varieties, selections, and species. Abnormal types of reaction, and much variation on individual plants and between different plants of the same species were observed.

In table 9 is given the reaction of 79 species, belonging to the tribes Agrostideae, Andropogoneae, Aveneae, Chlorideae, Festuceae, Hordeae, and Phalarideae, to each of the physiologic forms mentioned above.

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Table 9. Concluded.

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Forty of the 79 species were apparently immune to each of the six physiologic forms tested. Eighteen species (including 15 Avena species) gave a susceptible reaction to each of the six forms. Three of these (Achroyden aureum Boehmer, Anthoxanthum odoratum L., and Festuca octoflora Walt.) are wild grass species. Anthoxanthum odoratum and Festuca octoflora are common in occurrence throughout the United States, while Achroyden aureum is a native of Europe and is naturalized only in southern California. Dactylis glomerata L. gave a susceptible reaction to physiologic form 1, and minute uredinia were produced when it was inoculated with forms 3 and 5. The other forms (10, 11, and 12) produced only necrotic areas on this species. Minute uredinia were produced on Poa annua L. by each of the six physiologic forms used. Phleum pratense L. was quite variable in reaction. Uredinia were produced on this species by all forms, except form number 11 and repeated trials failed to produce uredinia when it was used. There was much variation in the production of chlorotic and necrotic areas on the different species. Apparently this type of infection is greatly influenced by environment and the age and condition of the host tissue. Newly developed leaves often gave a more susceptible reaction than older leaves.

It is evident from the data given in table 9 that certain of the species tested differ in their reaction when inoculated

-76-

with different physiologic forms. However, outside of the Avena group, this evidence of physiologic specialization is slight. Before physiologic forms could be accurately differentiated using gramineous hosts, other than Avena, it would be necessary to secure pure lines of the various species.

Melhus, Dietz and Willey (47), using <u>Puccinia coronata</u> <u>avenae</u>, obtained normal infection on <u>Alopecurus pratensis</u> L. and <u>Arrhenatherum elatius</u> (L.) Mert. and Koch., while in the present investigations the maximum infection observed on these species was type O. It is possible that they were using a physiologic form to which these species were particularly susceptible, or it may be that the variance in results is due to variation within the two gramineous species.

The following species produced uredinia, when inoculated with some one or more of the six physiologic forms used:

Achroyden aureum	Avena sativa diffusa
Anthoxanthum odoratum	Avena sat iva mu tica
Avena abysinnica	Avena sativa vegetalis
Avena barbata	Aven a strigosa
Avena brevis	Avena strigosa glaberscens
Avena nuda	Avena miestia
Avena nuda brevis	Dactylis glomerata
Avena sat iva	Festuca octoflora
Avena sativa aristata	Phleum pratense
Avena sat iva aurea	

-77-

DISCUSSION AND CONCLUSIONS

Forms 1, 3, and 5 seem to be common, occurring regularly from year to year. In addition there are evidently a number of comparatively rare forms of more or less infrequent occurrence. The source of these rare forms is not definitely known, but certain of them apparently originate on Rhamnus. If so, it is very probable that these and other new forms will continue to appear from year to year and it is altogether possible that new forms more virulent than form 5 will appear and that they will become widespread in distribution. However, the chances of such forms being produced are probably no greater for the future than they have been for the past.

Apparently the number of physiologic forms of crown rust present in certain localities is not constant from year to year. New forms seem to appear and certain of the old forms disappear. This is especially true in those regions where the alternate host functions. By collecting from different species of Rhamnus it may be possible to indefinitely continue the discovery of new physiologic forms.

The discovery of new differential hosts suggests the idea that they may also be a limiting factor in the discovery of new physiologic forms. It seems very probable that a more careful search would disclose additional differential hosts and that it might even be possible to sub-divide some of the physiologic forms now known. Also, the discovery of certain varieties possessing unusual resistance to the different physiologic forms and under field conditions, encourages further search for varieties resistant to all physiologic forms.

The varieties Victoria (C.I. 2401) and "Avena victoria" (C.I. 2764) are outstanding for their resistance to the eight physiologic forms secured during 1929 and 1930. Using resistant varieties of this type, it should be possible to produce, by hybridization, a hybrid selection resistant to crown rust, stem rust, loose smut, and covered smut. Such varieties as logold (C.I. 2329), Anthony (C.I. 2143), and Hajira (C.I. 1001) should furnish resistance to all physiologic forms of Puccinia graminis avenae now present in the United States. While Markton (C.I. 2053), Black Mesdag (C.I. 1877), and other smut-resistant varieties and selections, should furnish resistance to loose and covered smut. Appropriate crosses between these varieties, and between their hybrids should be of great value both from the standpoint of producing high yielding, resistant selections, and for a study of the inheritance of resistance to these various diseases. By crossing Victoria (C.I. 2401) or "Avena victoria" (C.I. 2764) with the different differential hosts, it should also be possible to determine the number of factors involved in the inheritance of resistance to different physiologic forms of crown rust.

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-79-

The investigation of physiologic specialization in crown rust should be continued, and apparently the relation of the alternate host to physiologic specialization and the production of new physiologic forms should be especially emphasized.

In Europe Klebahn (42) first divided <u>Puccinia coronata</u> Corda into two species <u>P. coronata</u> (Corda) Kleb. and <u>P. coronifera Kleb. He believed that <u>Rhamnus frangula</u> acted as a differential host for these two species. Dietel (19) raised Barclay's (10) variety <u>himalensis</u> to a species rank, thereby adding a third species. The aecial stage of <u>Puccinia himalensis</u> (Barcl.) Diet. was on <u>Rhamnus dahurica</u>. Then Mühlethaler (49) added a fourth species, <u>Puccinia alpinae-coronata</u> Muhl. with an aecial stage on <u>Rhamnus purshiana</u>.</u>

In America, Melhus, Dietz and Willey (47) and Dietz (22) have shown that certain species of Rhamnus act as differential hosts for the physiologic varieties of <u>Puccinia coronata</u> Corda.

The data secured in the present investigation indicates that certain species of Rhamnus may also act as differential hosts for physiologic forms of the physiologic variety <u>P. coronata avenae</u>.

Therefore, it is only logical to retain all physiologic varieties and forms of crown rust under the one morphologic species <u>Puccinia coronata</u> Corda. Physiologic specialization in the species is highly developed and is evident on both the Rhamnus and gramineous hosts.

-80-

SUMMARY

Two hundred forty-five cultures of <u>Fuccinia coronata avenae</u> were obtained from 171 collections made during the years 1927, 1928, 1929, and 1930. One or the other of two uniform sets of 33 possible differential hosts were individually inoculated with each of these 245 cultures.

Thirteen physiologic forms were distinguished among these cultures by the differential reaction, of seedlings, of eight pure-line-selected: varieties, selections, and species of oats, and one species that was not pure-line-selected.

Physiologic form 5 was the most virulent form described, only the two varieties Victoria (C.I. 2401) and "Avena victoria" (C.I. 2764), and the species <u>Avena strigosa glaberscens</u> (C.I. 2630) being resistant to it. Forms 5, 1, and 3 were the most widely distributed forms, the former two predominating in the south.

Evidence is presented which indicates that certain physiologic forms may originate on certain species of Rhamnus.

The seedling reaction of 316 variaties, selections, and species to physiologic forms 1, 3, 5, 10, 11, and 12 is given. The percentage of crown rust on certain of these same variaties under field conditions at seven locations in the United States is also recorded.

The variety Victoria (C.I. 2401) was outstanding for its

-81-

resistance to eight physiologic forms collected during 1929 and 1930, and to epiphytotic of crown rust under field conditions at Ames, Iowa, and Manhattan, Kansas, during 1929.

A study of the gramineous host range of physiologic forms 1, 3, 5, 10, 11, and 12 is reported. The reaction of all species, outside the Avena group, to each of these physiologic forms is very similar.

ACKNOWLEDGMENTS

The writer wishes to express his appreciation to Dr. S.M. Dietz for timely assistance given throughout the investigation, and to Dr. I.E. Melhus and Professor L.C. Burnett for cooperation, and assistance in preparing this manuscript. Appreciation is expressed also to Dr. H.B. Humphrey, Bureau of Plant Industry, Washington, D.C., under whose direction these investigations were conducted, and to Mr. T.R. Stanton, Bureau of Plant Industry, Washington, D.C., who furnished seed of the new and foreign varieties and species of cats studied. The writer is indebted to Mr. L.D. Leach for the nine collections of crown rust secured during 1927, and much additional material and information furnished at the beginning of the investigations. To the cooperators mentioned earlier in this manuscript and to those men who so kindly sent in collections of crown rust, the writer wishes to express his sincere gratitude.

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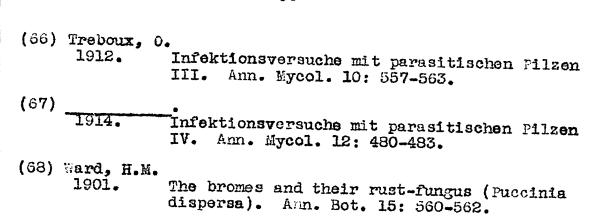
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EXPLANATION OF PLATES

Plate I

- Fig. 1. Muslin compartments in which all seedling plants were grown, to determine types of crown rust infection.
- Fig. 2. Covered lamp chimneys used to retain crown rust cultures.

Plate II

- Fig. 1. Tanks made of galvanized steel with false bottom and glass top used as inoculation chambers.
- Fig. 2. Types of crown rust infection:
 - I. No macroscopic evidence of infection.
 - 0. No uredinia formed, necrotic areas present.
 - 1. Uredinia few, small, always in necrotic areas, also more or less necrotic areas produced without the development of uredinia.
 - 2. Uredinia fairly abundant, small to medium size, always in necrotic or very chlorotic areas.
 - 3. Uredinia abundant, medium in size, and surrounded by chlorotic areas.
 - 4. Uredinia abundant, large, no necrosis or chlorosis immediately surrounding the uredinia.

Plate III

- Fig. 1. Puccinia coronata avenae (Physiologic form 1)
 - 1. Belar (C.I. 2760) type 4.
 - 7. Anthony (C.I. 2143) type O.
 - 8. Iowa 102 (C.I. 2464) type O.
 - 11. Avena strigosa glaberscens (C.I. 2630) type O.
 - 21. Victoria (C.I. 2401) type 0.
 - 28. Markton (C.I. 2053) type 4.

Fig. 2.	Puccinia coronata avenae (Physiologic form 3).
	1. Belar (C.I. 2760) - type O.
	7. Anthony (C.I. 2143) - type 4.
	8. Iowa 102 (C.I. 2464) - type 4.
	11. Avena strigosa glaberscens (C.I. 2630) - type O.
	21. Victoria (C.I. 2401) - type O.
	28. Markton (C.I. 2053) - type 4.
	Plate IV.
Fig. 1.	Puccinia coronata avenae (Physiologic form 5).
	1. Belar (C.I. 2760) - type 4.
	7. Anthony (C.I. 2143) - type 4.
	8. Iowa 102 (C.I. 2464) - type 4.
	11. Avena strigosa glaberscens (C.I. 2630) - type .O.
	21. Victoria (C.I. 2401) - type O.
	28. Markton (C.I. 2053) - type 4.
Fig. 2.	Puccinia coronata avenas (Physiologic form 6).
	1. Belar (C.I. 2760) - type 3.
	7. Anthony (C.I. 2143) - type 4.
	8. Iowa 102 (C.I. 2464) - type O.
	11. Avena strigosa glaberscens (C.I. 2630) - type O.
	21. Victoria (C.I. 2401) - type O.
	28. Markton (C.I. 2053) - type 4.

Plate V

Fig. 1. Puccinia coronata avenae (Physiologic form 10). 1. Belar (C.I. 2760) - type 0. 7. Anthony (C.I. 2143) - type O. 8. Iowa 102 (C.I. 2464) - type O. 11. Avena strigosa glaberscens (C.I. 2630) - type 4. 21. Victoria (C.I. 2401) - type I. 28. Markton (C.I. 2053) - type 4. Puccinia coronata avenae (Physiologic form 11). Fig. 2. 1. Belar (C.I. 2760) - type 4. 7. Anthony (C.I. 2143) - type O. 8. Iowa 102 (C.I. 2464) - type 4. 11. Avena strigosa glaberscens (C.I. 2630) - type O. 21. Victoria (C.I. 2.01) - type O. 28. Markton (C.I. 2053) - type 4. Plate VI Puccinia coronata avenae (Physiologic form 12) Fig. 1. 1. Belar (C.I, 2760) - type O. 7. Anthony (C.I. 2143) - type O. 8. Iowa 102 (C.I. 2464) - type 0. 11. Avena strigosa glaberscens (C.I. 2630) - type I. 21. Victoria (C.I. 2401) - type I. 28. Markton (C.I. 2053) - type 4.

Fig. 2. Puccinia coronata avenae (Physiologic form 13).

1. Belar (C.I. 2760) - type O.

7. Anthony (C.I. 2143) - type 1.

8. Iowa 102 (C.I. 2464) - type 4.

11. Avena strigosa glaberscens (C.I. 2630) - type O.

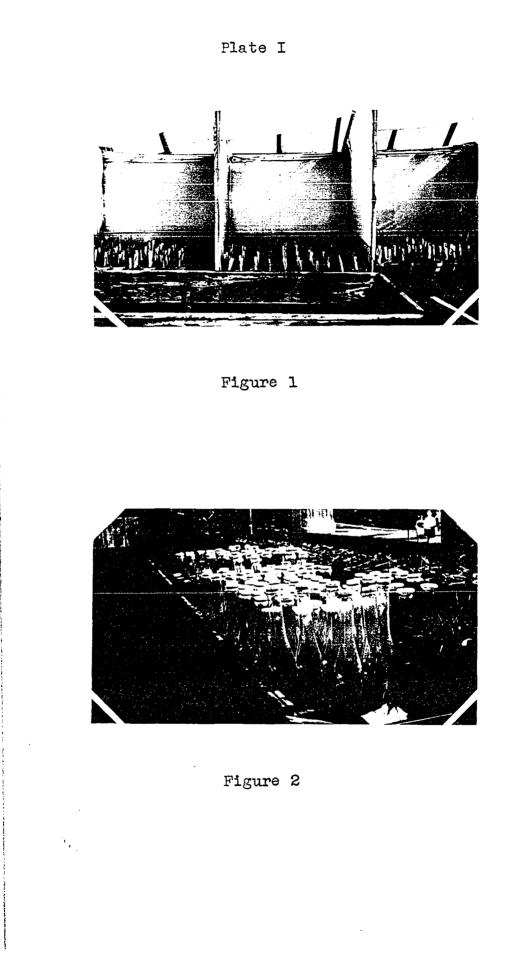
21. Victoria (C.I. 2401) - type O.

28. Markton (C.I. 2053) - type 4.

Plate VII

Teliospore formation of physiologic forms 3 and 5 on seedling plants from two panicles of oats. All infection 16 days old and produced under identical conditions.

- 1. Fulghum (C.I. 650-203) abundant telia formation, physiologic form 3.
- 2. Fulghum (C.I. 650-203) abundant uredinia formation (no telia), physiologic form 5.
- 3. D.A.C. # 10 (C.I. 2829) abundant telia formation, physiologic form 3.
- 4. D.A.C. # 10 (C.I. 2829) abundant uredinia formation (no telia), physiologic form 5.





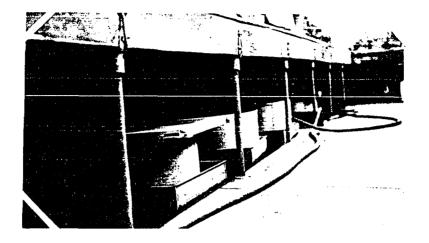


Figure 1

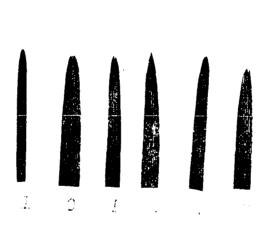


Figure 2

